Phialophora and some similar morphologically little-differentiated anamorphs of divergent ascomycetes

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Abstract: Phialophora is a little-differentiated genus of more or less pigmented, phialidic hyphomycetes. With the addition of numerous species, it has become highly polyphyletic, comprising anamorphs of discomycetes, pyrenomycetes and loculoascomycetes. The core of the genus consists of anamorphs of Capronia in the Herpotrichiellaceae, Chaetothyriales. Some taxa have already been segregated from Phialophora into genera such as Lecythophora (Coniochaetaceae) and Phaeoacremonium (Magnaporthaceae?). Ascomycete orders in which phialophora-like fungi have been placed are reviewed, and some further segregation is proposed. For the common anamorphs of the discomycete family Dermateaceae, the old generic name Cadophora is available and should be used for Ph. fastigiata and related taxa (one new combination). For anamorphs of Gaeumannomyces and Magnaporthe (Magnaporthaceae), a new genus, Harpophora, is proposed with four new combinations. Criteria that allow a correlation with the suggested broad-scale subdivision are outlined.

Key words: Cadophora, Capronia, Gaeumannomyces, Harpophora, Lecythophora, Magnaporthe, Phaeoacremonium, teleomorph connections.

Introduction

Like Acremonium Link and Verticillium Nees among the hyaline phialidic hyphomycetes, Phialophora Medlar is a model of a poorly defined, little differentiated and highly polyphyletic genus of more or less pigmented hyphomycetes that requires disentangling. Some species are very common saprobes in soil, wood pulp and other plant material, others are more specialized plant pathogens; human pathogenicity is known for a few species. Whilst it is still extremely difficult to differentiate species among these anamorphs, it seems possible to distinguish and name a few groups of higher ranks that can be correlated with teleomorph orders or families. Some first steps in this direction are proposed here.

The most comprehensive monograph of *Phialo-phora* so far produced is that by Schol-Schwarz (1970). The genus was vaguely characterized by more or less pigmented hyphae and conidiophores, solitary or aggregated, flask-shaped phialides, usually with a flaring collarette. Several additions and clarifications of species were published subsequently, e.g.

by Cole & Kendrick (1973) and Sivasithamparam (1975). Gams & Holubová-Jechová (1976) described a new, obviously also heterogeneous section, Catenulatae W. Gams, based on the anamorph of Lasiosphaeria hirsuta (Fr.) Ces. & De Not. This section was characterized by often catenate conidia with distinctly truncate conidial bases; it included several species of varying pigmentation. A key to some phialophora-like genera in the broadest sense was given by Crous et al. (1996).

A first step towards excluding defined groups of taxa from *Phialophora sensu* Schol-Schwarz was made by Gams & McGinnis (1983), who recognized *Lecythophora* Nannfeldt (Fig. 2 d) for anamorphs of *Coniochaeta* (Sacc.) Cooke [in Schol-Schwarz the *Ph. hoffmannii*-group + *Ph. lignicola* (Nannf.) Goidànich].

Coryne Nees, anamorph of the discomycete genus Ascocoryne J.W. Groves & D.E. Wilson (1967), was included in *Phialophora* by Schol-Schwarz (1970) but can easily be distinguished, as discussed below. 188 W. GAMS

Phaeoacremonium W. Gams et al. (Fig. 2 f) was introduced by Crous et al. (1996) for species similar to Phialophora parasitica Ajello et al., colonizers of vascular systems in woody plants and from medical origins, with slender, aculeate phialides and non-flaring collarettes.

Several similar fungi were not included in *Phia-lophora* previously, but should be considered here:

Phialocephala W.B. Kendrick (1961) is characterized by dark conidiophore stipes and densely penicillate conidiophores bearing conidia in slimy masses.

Cladorrhinum Sace. & Marchal (Fig. 2 b) was reintroduced by Von Arx & Gams (1967) for phialophora-like anamorphs with almost exclusively intercalary phialides with widely flaring collarettes, formed in a reticulate system of aerial hyphae.

Exophiala J.W. Carmichael is now clearly defined for black-yeast anamorphs of Capronia Sacc., with imperceptably annellate, narrow conidiogenous openings (de Hoog, 1977; Masclaux et al., 1995; and many subsequent publications). Wangiella McGinnis (1977) was introduced for a similar, closely related fungus that has conidiogenous cells with multiple annellidic openings, as well as separately developing phialides.

Pseudogliomastix W. Gams (Fig. 3 a) was introduced by Gams & Boekhout (1985) for the anamorph of Wallrothiella subiculosa Höhnel, which was formerly classified in Gliomastix Guég. It is characterized by pigmented, branched conidiophores and somewhat darker conidia, the walls of which are pigmented throughout and not only pigment-incrusted, as they are in some species of Acremonium section Gliomastix.

Redelimitation and reclassification of phialophora-like anamorphs

In this section, the various components of *Phialo-phora* are distributed over the respective orders and families of ascomycetes, with some necessary renaming. In cases where the anamorphs are rarely encountered in culture separate from the teleomorph, little need is felt for formal anamorph binomials or separate anamorph genera.

LEOTIALES

Helotiaceae

Species of Ascocoryne J.W. Groves & D.E. Wilson 1967 have anamorphs now classified in Coryne (Fig. 1 f), with hyaline, more or less penicillate conidiophores and phialides that lack any sign of visible collarette. They have a strong tendency to form sporodochium-like or gelatinous conidiomata in vivo and

in vitro (Fig. 1 f; Roll-Hansen & Roll-Hansen, 1979; Seifert, 1989). Claussenomyces Kirschst. is normally connected with synnematous anamorphs in Dendrostilbella Höhn. (Seifert, 1985) but some species form anamorphs in vitro that are indistinguishable from Coryne (G. Marson, unpublished).

Dermateaceae

The commonest saprobic species (-aggregates) classified in Phialophora are Ph. fastigiata (Lagerb. & Melin) Conant (Fig. 1 a) and Ph. malorum (Kidd & Beaumont) McColloch (Fig. 1 b, c). Le Gal & Mangenot (1960, 1961) demonstrated their relationship with discomycete teleomorphs in Mollisia (Fr.) P. Karst., Pyrenopeziza Fuckel and related genera. Greenleaf & Korf (1980) isolated a Phialophora that is similar to Ph. fastigiata (Lagerb. & Melin) Conant from Mollisia dextrinophila Korf (Fig. 1 d). Other recent ascospore isolates of such fungi did not produce phialophora-like anamorphs (M.M. Nauta, pers. comm.). While the vegetative hyphae in these species are more or less pigmented, the phialides are pale to hyaline, particularly the collarettes. For these species, the genus Cadophora Conant is available, which certainly now warrants recognition. The correct binomials should therefore be used as follows:

Cadophora fastigiata Lagerb. & Melin Cadophora melinii Nannf.

Cadophora repens Davidson

Cadophora malorum (Kidd & Beaum.) W. Gams, comb. nov. ≡ Sporotrichum malorum Kidd & Beaum., Trans. Brit. Mycol. Soc. 10: 111. 1974 ≡ Phialophora malorum (Kidd & Beaum.) McColloch, Mycologia 36: 589. 1944. The synonyms listed by Schol-Schwarz (1970) under this name still require critical revision.

Other binomials available in Cadophora are not adopted at this time, pending a more thorough study on their identity.

Anamorphs of the closely related genus *Tapesia* (Pers.) Fuckel (= *Mollisia* (Fr.) P. Karst, *nom. cons.*) were placed by Aebi (1972) in the similar genus *Cystodendron* Bubák (Fig. 1 g), characterized by dark, densely penicillate and more or less sporodochial conidiophores and phialides with pronounced tubular collarettes.

Pseudopezicula tracheiphila (Müll.-Thurg.) Korf & Zhuang was associated with the anamorph Phialophora tracheiphila (Sacc. & D. Sacc.) Korf (Korf et al., 1986). The vascular pathogen causing brown stem rot in soya beans, Phialophora gregata (Allington & Chamberlain) W. Gams (Fig. 1 e; Chen et al., 1996; Yamamoto et al., 1992), also has discomycete affinities according to 18S and 28S rDNA sequences (Paulin



Fig. 1. Phialophora-like anamorphs of discomycetes. a. Cadophora fastigiata, CBS 226.30. b. and c. Cadophora malorum, CBS 260.32 and 258.89. d. Anamorph of Mollisia dextrinospora, CBS 401.78. e. Phialophora gregata, CBS 184.70. f. Coryne anamorph of Ascocoryne sarcoides, CBS 247.80. g. Cystodendron dryophilum, from type of Tubercularia dryophila in Herb. B. h. Myrioconium sp., CBS 465.71. i. Anamorph of Cistella hymeniophila, CBS 886.73 and herb. H.A. 4378 in CBS.

& Harrington, this volume). The generic classification of the anamorphs of these species awaits reconsideration.

Hyaloscyphaceae

Raschle (1977) illustrated several phialophora-like anamorphs of *Hyalopeziza* Fuckel, *Urceolella* Boud., and *Unguicularia* Höhn. species, without a detailed analysis. The connection of *Ph. rhodogena* (Mangenot) W. Gams from section *Catenulatae* (Fig. 1 i) with the teleomorph *Cistella hymeniophila* (P. Karst.) Korf was established by Helfer (1991). This species, like *Ph. brachyconia* W. Gams, *Ph. hyalina* W. Gams, and *Ph. brevicollaris* W. Gams, differs from the remaining members of the section by its hyaline hyphae and phialides. A separate generic classification may be envisaged.

Sclerotiniaceae

The common microconidial forms of many Sclerotiniaceae classified in Myrioconium Syd. (Fig. 1 h) have generally hyaline structures with densely, irregularly penicillate conidiophores, phialides with a pronounced collarette and slimy, subglobose conidia. A similar, not necessarily congeneric fungus, has been classified in Phialophora as Ph. alba van Beyma.

CALICIALES

Mycocaliciaceae

Mycocalicium schefflerae Samuels & D.E. Buchanan (1983) was shown to have a lecythophora-like palely pigmented anamorph with broad hyphae and many intercalary phialides.

SORDARIALES

Sordariaceae

Podospora austrohemisphaerica Lundqvist et al. (1999) was described as having densely penicillate conidiophores and phialides with a widely flaring collarette, resembling Ph. cyclaminis van Beyma; conidia are globose, individually hyaline but brownish in mass. In other species of the genus, the phialides are shorter and broadly flask-shaped, all producing guttulate conidia (Bell & Mahoney, 1995). According to Lundqvist et al. (l.c.), twenty species of Podospora Ces. have so far been linked with phialophora-like anamorphs, others with Cladorrhinum. It is noteworthy to observe that these conidia do not germinate in vitro and apparently have spermatial functions, suggesting little need for a formal anamorph name.

Wallrothiella subiculosa Höhnel, [anam. Pseudogliomastix protea (Sacc.) W. Gams & Boekhout (1985) (Fig. 3 a)], seems to occupy a basal position in the Sordariales, but its family classification is still uncertain (Réblová & Winka, 2000); an affinity to the Niessliaceae has been assumed, but, according to 28S rDNA sequences, the genus falls outside the Hypocreales (Rehner & Samuels, 1995). According to conidiophore and conidial pigmentation, Phialophora geniculata van Emden 1975 (Fig. 3 b), a species common in tropical alkaline soils (Nagai et al., 1995) (syn. Ph. taiwanensis Matsushima 1983), is very close to the anamorph of W. subiculosa and may be congeneric.

Batistiaceae

Batistia annulipes (Mont.) Cif. was connected with the synnematous anamorph genus Acrostroma Seifert in cultural work by Samuels & Rodrigues (1989). This fungus is a synnematous counterpart of Phialophora section Catenulatae.

Lasiosphaeriaceae

Lasiosphaeria hirsuta (Fr. : Fr.) Ces. & De Not., L. hispida (Tode: Fr.) Fuckel (Fig. 2 a) and L. ovina (Fr. : Fr.) Ces. & De Not. of Phialophora section Catenulatae have pigmented mycelium and phialophora-like conidiophores with mostly catenate conidia. The teleomorphs of Cladorrhinum (Fig. 2 b) are found in Apiosordaria von Arx & W. Gams and Cercophora Fuckel in this family (Mouchacca & Gams, 1993). An anamorph more typical of Phialophora with conidial heads is seen in the similar Eosphaeria uliginosa (Fr.) Höhnel (Petrini et al., 1984). Barrina polyspora Ramaley (1997), with its polysporous asci, might belong to this family as well. Its anamorph resembles the lecythophora-like anamorph of Munkovalsaria rubra Aptroot et al. in the Dacampiaceae. Because cultures of these anamorphs are probably rarely encountered separately from their teleomorphs, no nomenclatural formalization is envisioned at this time.

Chaetosphaeriaceae

This family was introduced for *Chaetosphaeria* Tul. & C. Tul. and related genera by Réblová et al. (1999) and comprises mainly anamorphs in *Chloridium* Link, *Dictyochaeta* Speg., *Menispora* Pers., *Kylindria* DiCosmo et al. and *Cacumisporium* Preuss (see Réblová, this volume). *Phialophora phaeophora* W. Gams (section *Catenulatae*) (Fig. 2 c) is the anamorph of *Chaetosphaeria pygmaea* (P. Karst.) O. Const. (Constantinescu et al., 1995). This species could be interpreted as a miniature *Chloridium*.

Coniochaetaceae

Lecythophora Nannf. (Fig. 2 d) has been repeatedly correlated with teleomorphs in Coniochaeta (e.g. Checa

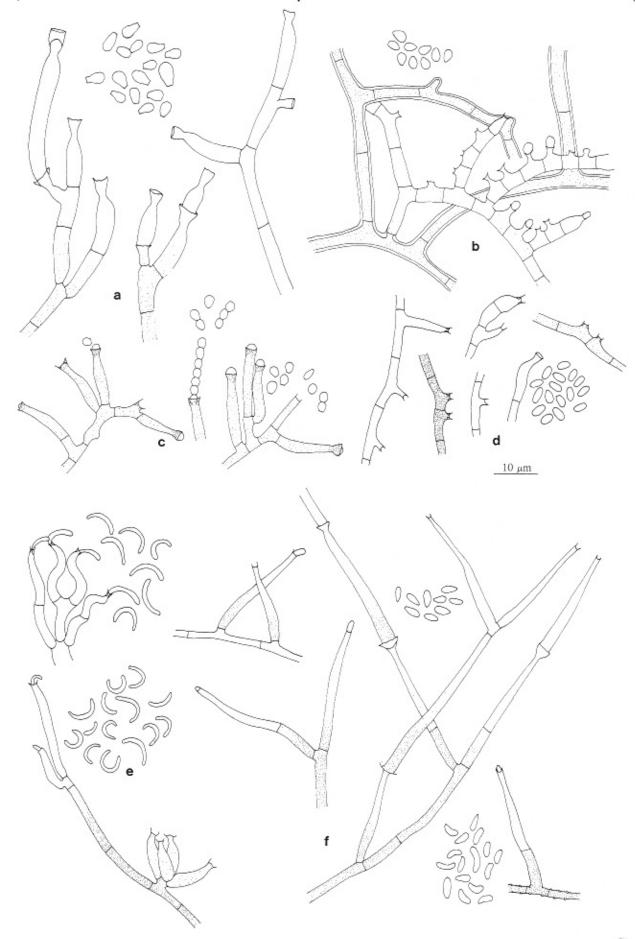


Fig. 2. Phialophora-like anamorphs of pyrenomycetes. a. Anamorph of Lasiosphaeria hispida, CBS 955.72. b. Cladorrhinum samala, type specimen, IMI 113129. c. Phialophora phaeophora anamorph of Chaetosphaeria pygmaea, CBS 699.74. d. Lecythophora lignicola, CBS 267.33 e. Harpophora radicicola, CBS 296.53 and 350.77. f. Phaeoacremonium parasiticum, CBS 984.73.

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Checa et al., 1988; Romero et al., 1999). The genus is characterized and differentiated from *Phialophora* by more or less hyaline undifferentiated fertile hyphae, with mostly intercalary phialides, with very short lateral necks but mostly visible periclinal wall thickenings below the hardly flaring collarettes. The different anamorphs of distinct *Coniochaeta* species have not yet been differentiated in culture.

Helminthosphaeriaceae

Veramycina elegans Subram. is a phialocephala-like synanamorph of Oedemium minus (Link) S. Hughes, anamorph of Chaetosphaerella phaeostroma (Durieu & Mont.) E. Müll. & C. Booth in the Helminthosphaeriaceae (Réblová, 1999), but this fungus does not grow in culture.

DIAPORTHALES

Magnaporthaceae

A distinct group of species formerly included in Phialophora comprises the anamorphs of Gaeumannomyces von Arx & D.L. Olivier and Magnaporthe R.A. Krause & R.K. Webster, which have mostly sickle-shaped phialoconidia (Fig. 2 e). The colonies are fast-growing and often have darker and broader 'runner hyphae' (see also Cunningham, 1981). In several species, narrower and strongly curved conidia were found to be non-germinable spermatia, while broader, less curved conidia germinated readily in vitro (Wong & Walker, 1975). The habit of the fastgrowing, thin colonies is a valuable criterion for recognizing this group (Deacon, 1973, 1974; Landschoot & Jackson, 1989; Scott & Deacon, 1983; Walker, 1980, 1981; Wong & Walker, 1975). The introduction of a new genus for this group is appropriate because the anamorphs are morphologically sufficiently distinct. Only a few anamorph species have so far formally been named in this complex, besides the well-known plant-pathogenic teleomorphs. A further addition is the closely related Cephalosporium maydis Samra et al., for which so far no adequate generic classification has been available (Bryan et al., 1995; Ward & Bateman, 1999). According to ITS sequences, Phialophora radicicola Cain (Fig. 2 e) is a central species in this complex, well suited to serve as type of the new anamorph genus Harpophora. Cephalosporium maydis, together with Magnaporthe poae Landschoot & Jackson and Gaeumannomyces incrustans Landschot & Jackson are genetically somewhat more distant (Ward & Bateman, 1999).

HARPOPHORA W. Gams, gen. nov.

Genus hyphomycetum, anamorphoses Magnaporthacearum complectens. Coloniae celeriter crescentes, plus minusve olivaceo-brunneae, hyphis latioribus radiantibus. Phialides Phialophorae similes, plus minusve pigmentatae, collare divergens ferentes. Conidia cylindrica, valde curvata, fere hyalina. Species typica Harpophora radicicola (Cain) W. Gams.

A genus of hyphomycetes comprising phialidic anamorphs of the *Magnaporthaceae*. Colonies growing rapidly, more or less olivaceous-brown, typically showing some broader radiating 'runner hyphae' and narrower lateral hyphae. Phialides resembling those of *Phialophora*, more or less pigmented, with a conspicuous divergent collarette. Conidia borne in slimy heads, cylindrical with strong curvature, hyaline, sometimes not germinable *in vitro*. Type species *Harpophora radicicola* (Cain) W. Gams.

Etym.: Greek harps = sickle, in reference to the conidial shape, pherein = to bear and alluding to the similarity with Phialophora.

Harpophora radicicola (Cain) W. Gams, comb. nov. (Fig. 2 e).

Phialophora radicicola Cain, Canad. J. Bot. 30: 340. 1952.

Harpophora maydis (Samra et al.) W. Gams, comb. nov.

Cephalosporium maydis Samra, Sabet & Hingorani, Phytopathology 53: 404. 1963.

Harpophora graminicola (Deacon) W. Gams, comb.

≡ Phialophora radicicola var. graminicola Deacon, Trans. Brit. Mycol. Soc. 63: 322. 1974 ≡ Phialophora graminicola (Deacon) John Walker, Mycotaxon 11: 90. 1980.

Harpophora zeicola (Deacon & D.B. Scott) W. Gams, comb. nov.

≡ Phialophora zeicola Deacon & D.B. Scott, Trans. Brit. Mycol. Soc. 81: 256. 1983.

Phaeoacremonium Crous et al. (1996) (Fig. 2 f) is characterized by pigmented conidiophores that gradually become paler in the upper part, slender, aculeate phialides with hardly flaring collarettes, and hyaline, mostly allantoid conidia. Yan et al. (1995) already demonstrated that the type species, formerly classified as Ph. parasitica Ajello et al., was unrelated to other species of Phialophora using ITS rDNA sequences. Its affinity with the Magnaporthaceae was shown by Dupont et al. (1998). One equally common colonizer of woody vascular bundles in grapevine, the causal agent of Petri grapevine decline called Phaeoacremonium chlamydosporum W. Gams et al., falls outside this complex. It has been transferred to a separate genus, Phaeomoniella Crous & W. Gams in the Chaetothyriales (Fig. 4 c; Crous & Gams, 2000). This fungus is also morphologically distinguished by very darkly pigmented conidiophore stipes, which contrast sharply with the pale phialides and never curved conidia.

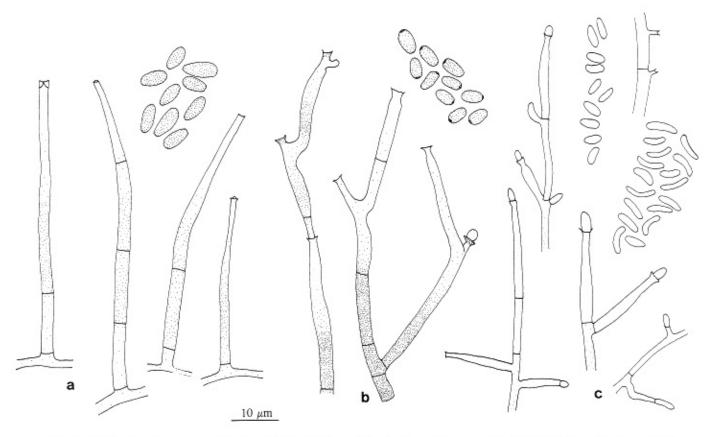


Fig. 3. Phialophora-like anamorphs of uncertain affinity. a. Pseudogliomastix anamorph of Wallrothiella subiculosa, CBS 656.94. b. Phialophora geniculata, CBS 673.83. c. Margarinomyces bubakii, CBS 269.33.

OPHIOSTOMATALES

Ophiostomataceae

While most anamorphs in this family have polyblastic conidiogenesis, a few species of *Ophiostoma* have *Phialocephala* anamorphs. A similar genus, *Sporendocladia* G. Arn. ex Nag Raj & W.B. Kendrick was delimited by Wingfield *et al.* (1987) because of 'ring wall building' in the development of basally truncate, catenate conidia. A phylogenetic analysis of this group is still missing.

HYPOCREALES

Clavicipitaceae

The classification of the tolypocladium-like *Phialophora endoparasitica* Barron & Szijarto (1982) and *Ph. tribrachiata* Barron & Szijarto (1984) in the *Clavicipitaceae* is still speculative, lacking teleomorph or DNA sequence evidence. These species parasitize rotifers and form hyaline colonies. In any case, they are unrelated to other species of *Phialophora*.

Acremonium, a genus with Hypocreales affinities, need not be further considered in this context, except

for A. atrogriseum (Panasenko) W. Gams, which was recently again described as a new species, *Phialophora intermedia* Iwatsu et al. (1988). The correct classification of this fungus with slightly swollen phialides and somewhat pigmented mycelium has not yet been resolved.

Ceratostomataceae (Melanosporaceae), family of uncertain affinity (Hypocreales or Sordariales?, Rehner & Samuels, 1995).

The genus Proteophiala Ciferri (1957) (and not the sometimes cited genus Harziella Cost. & Matr., an illegitimate homonym, introduced for a very distinct hypocrealean mycoparasite growing on Lepista nuda) is best suited to accommodate the short-lived phialidic anamorphs of many Ceratostomataceae: Species of Melanospora Corda, Microthecium Corda (Hawksworth & Udagawa, 1977), Persiciospora P.F. Cannon & D. Hawksw., Sphaerodes Clem. (Horie et al., 1986), Pteridiosperma Penz. & Sacc. (Udagawa & Muroi, 1981), and synanamorphs of Harzia Cost., Olpitrichum G.F. Atk. and Papulaspora Preuss. This genus is distinguished by its completely hyaline hyphae, conidiophores and conidia, and the flask-

shaped phialides borne on apically more or less inflated conidiophores.

SPATHULOSPORALES (order of uncertain affinity) In Hispidocarpomyces galaxauricola Nakagiri (Hispidocarpomycetaceae), Nakagiri (1993) found penicillate spermatiophores with very long tubular collarettes, and in Retrostium amphiroae Nakagiri & Tad. Ito (1997) (Spathulosporaceae) the collarettes were somewhat shorter and more phialophora-like.

DOTHIDEALES

Dothideaceae

The tree root-inhabiting and poorly sporulating species, *Phialophora finlandica* C.J.K. Wang & Wilcox and *Phialocephala fortinii* C.J.K. Wang & Wilcox (sterile mycelium known as *Mycelium radicis atrovirens* Melin) (Wang & Wilcox, 1985; Harney *et al.*, 1997), are related to *Hortaea* Nishim. & Miyaji in the *Dothideales* (Rogers *et al.*, 1999). Other species of *Phialocephala* have teleomorphs in *Ophiostoma* (see above).

Dacampiaceae

Munkovalsaria rubra Aptroot et al. (Aptroot, 1995) is associated with a lecythophora-like anamorph with relatively broad hyphae. No binomial anamorph name is needed for this very characteristic fungus, which has the unique odour of m-cresol in culture.

Unclassified family

Tirisporella beccariana (Ces.) E.B.G. Jones et al. (1996) has very broad hyphae and a phialophora-like anamorph with cup-shaped collarettes and elongate dacryoid conidia, otherwise resembling *Ph. olivacea* W. Gams in section *Catenulatae*.

CHAETOTHYRIALES

Herpotrichiellaceae

Molecular studies by Yan et al. (1995), Untereiner & Naveau (1999), de Hoog et al. (1999) and others have shown that the type species of Phialophora, Ph. verrucosa Medlar, and its closest relative, Ph. americana (Nannf.) S. Hughes (Fig. 4 a), belong to the Herpotrichiellaceae, Chaetothyriales. The latter species was connected to the teleomorph Capronia semi-immersa (Candoussau & Sulmont) Untereiner & Naveau (Untereiner & Naveau, 1999). These species are commonly found on decayed wood and both can cause chromoblastomycosis and other diseases in humans, as reviewed by de Hoog et al. (1999). Morphologically, they are characterized by short, lageniform, darkly pigmented phialides, with distinct, usually even darker, flaring collarettes (Fig. 5, left column).

This combination of morphological features distinguishes these and two newly described taxa (de Hoog et al., 1999) from all others classified in the genus. Other Capronia teleomorphs with Phialophora connections are Capronia acutiseta Samuels, C. epimyces M.E. Barr and C. dactylotricha Untereiner et al. (Untereiner, 1995; Untereiner et al., 1995; Untereiner, this volume); the latter two also have Exophiala synanamorphs with percurrent proliferation. This is the group of Phialophora species that comprises most of the medically relevant members and it will continue to form the core of a naturally delimited genus Phialophora.

Exophiala is a monophyletic though highly diversified group, also connected to teleomorphs in Capronia (Uijthof, 1996; Uijthof et al., 1997; Untereiner & Naveau, 1999; Rogers et al., 1999); it is characterized by dark conidiogenous cells with inconspicuous annellidic elongation (Fig. 5, top left). Structures appearing similar to polyphialides in E. dermatitidis have the same annellidic mechanism, though extremely condensed, seen when using SEM, as well as the occasional occurrence of phialides with balloon-shaped collarettes. McKemy et al. (1999) still recognized the genus Wangiella McGinnis (1977) for this species and E. heteromorpha because of the additional presence of phialides, but in both species conidiogenous cells with percurrent elongation (though inconspicuous) predominate. Haase et al. (1999), using SSU rDNA sequencing, found that E. dermatitidis, E. heteromorpha and E. mansonii form a weakly separated subclade within the Herpotrichiellaceae, for which no clear-cut phenetic criterion is available. Therefore, it seems preferable to retain all these closely related taxa in one genus, Exophiala.

Cladophialophora Borelli (1980) is intimately related with the other Herpotrichiellaceae (Masclaux et al., 1995; Gerrits van den Ende & de Hoog, 1999). It is distinguished by the production of a synanamorph with chains of cladosporium-like, hardly seceding blastoconidia, as well as phialophora-like conidia.

UNCLASSIFIED

Margarinomyces: Van Beyma (1943) recognized Margarinomyces Laxa as distinct from Phialophora, because the phialides of its species commonly lack flaring collarettes. Schol-Schwarz (1970) merged the two genera, because in the type species, M. bubakii Laxa (Fig. 3 c), collarettes can sometimes be observed. A feature generally overlooked in this fungus is the 'acropleurogenous' pattern of branching; i.e., a prostrate fertile hypha ends with a phialidic opening, then, tracing it backwards, one or several intercalary phialides follow with lateral necks, before further back discrete, lateral phialophora-type phialides with a basal

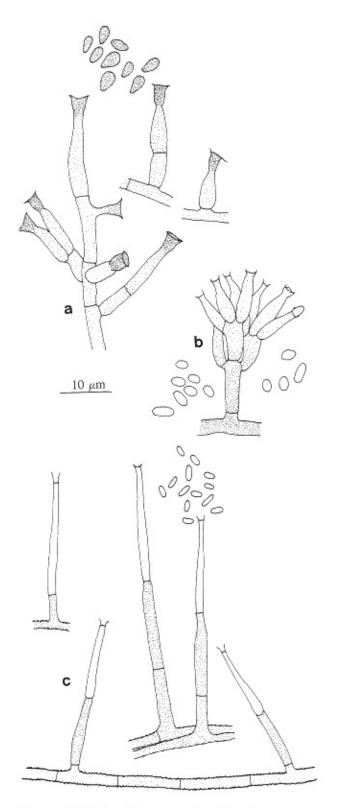


Fig. 4. Phialophora-like anamorphs of loculoascomycetes. a. *Phialophora americana*, CBS 281.35. b. *Phialophora* ? finlandica, CBS 166.90. c. *Phaeomoniella chlamydospora*, CBS 239.74.

septum are formed. This feature is unusual in this complex and seems to warrant the recognition of a

redefined genus *Margarinomyces*. This pattern is also observed in the hyaline counterpart, *Phialemonium* W. Gams & McGinnis (1983). But the affinity of these two anamorph genera among the ascomycetes has not yet been elucidated.

The species *Phialophora richardsiae* (Nannf.) Conant and *Ph. mustea* Neerg, are characterized by their dimorphic conidia (darkly pigmented and hyaline) and saucer-shaped collarettes. The former species has been sequenced by Yan *et al.* (1995) and found to occupy a distant position from other *Phialophora* species, but no conclusion about its phylogenetic affinities was made.

The causal agent of carnation wilt, *Ph. cinerescens* (Wollenw.) van Beyma, has densely penicillate hyaline phialides with pronounced vase-shaped collarettes and somewhat pigmented conidia. It may also have a remote relationship.

Not all species described by Schol-Schwarz (1970) and subsequent authors in *Phialophora* can be reclassified, at present, and this survey is admittedly incomplete. To revise the enormous amount of material available will require major efforts. At the moment, a broad genus *Phialophora sensu lato* continues to have its value as a kind of dustbin for poorly classified fungi or simply as a morphological term. Less differentiated phialophora-like synanamorphs commonly occurring as synanamorphs of other hyphomycete genera are not treated here.

Criteria for distinction of anamorph groups

Conidiogenesis is phialidic in all these fungi, with conidia being blown out in basipetal succession from a single conidiogenous locus within a collarette, with pronounced periclinal wall thickening resulting from successive conidium formation.

The most suitable criteria for further subdivision are found in the pigmentation of the mycelium and the fertile hyphae. In species of *Phialophora sensu stricto*, all structures are darkly pigmented. In *Cadophora* and *Harpophora* species, old hyphae are dark, younger parts near-hyaline, with conidiophores and phialides being intermediate. Colonies of *Harpophora* species grow faster and have sickle-shaped conidia.

In addition, colony habit and growth rate are valuable in recognizing members of the *Chaetothyriales*. Their colonies are very dark olivaceous, compact, black-yeast-like, and slow-growing. The strongest contrast occurs with the *Harpophora* anamorphs of *Gaeumannomyces* species, which have less pigmented and fast-growing colonies, while other groups are intermediate.

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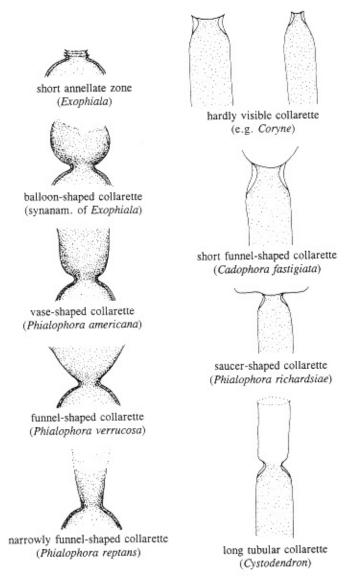


Fig. 5. Shapes of phialides in phialophora-like taxa (left column after de Hoog et al., 1999).

Phialide shape: Short flask-shaped phialides or intercalary phialides with sessile collarettes are characteristic of species of *Phialophora sensu stricto*. In most other taxa discussed, the phialides are more or less cylindrical.

Intercalary phialides: In most taxa of *Phialophora* sensu lato, intercalary phialides with lateral collarettes occur occasionally as well as the dominant terminal phialides. In species of *Lecythophora* and *Cladorrhinum*, intercalary phialides predominate, whereas terminal or lateral discrete phialides comprise a minority. An acropleurogenous pattern is characteristic of discrete groups of anamorphs: behind a terminal phialide follows a series of intercalary cells with

subterminal lateral phialidic necks, before discrete lateral phialides are formed further back. This is particularly characteristic of *Margarinomyces bubakii* in the pigmented and *Phialemonium* species in the non-pigmented phialophora-like hyphomycetes.

Shape of the collarette (Fig. 5): In species of Phialophora sensu stricto, de Hoog et al. (1999) distinguished four shapes of collarettes, which all are somewhat constricted at the phialide tip (Fig. 5, left column). In addition, the extremes of either saucershaped or perfectly tubular collarettes occur, as well as less differentiated shapes (Fig. 5, right column). In Margarinomyces and Coryne species, flaring collarettes are usually absent.

Conidial shape and pigmentation: In most phialophora-like taxa, conidia are elongate. But (sub-)globose to dacryoid conidia occur in *Ph. cyclaminis*, *Ph. richardsiae*, *Myrioconium* spp, *Cladorrhinum* spp., and anamorphs of *Podospora* spp., *Tirisporella beccariana*, etc. In addition to the species of *Phialophora sensu stricto*., the occurrence of pigmented conidia is a valuable feature in identifying *Ph. cinerescens*. Dimorphic, strongly pigmented and hyaline, conidia are characteristic of the still unclassified species *Ph. richardsiae* and *Ph. mustea* Neergaard.

Conidial germinability may be a further criterion. In Cladorrhinum foecundissimum Sacc. & Marchal, conidia do not germinate, but in some other species of the genus they do. For the anamorphs of Podospora, Lundqvist et al. (1999) stated that conidia have a spermatial function, as they do in the members of the Spathulosporales listed. In Harpophora species, germinable conidia are observed next to non-germinable spermatia, apparently not sharply morphologically delimited (Wong & Walker, 1975).

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Literature cited

AEBI, B., 1972 — Untersuchungen über Discomyceten aus der Gruppe *Tapesia-Trichobelonium*. — Nova Hedwigia 23: 49–112.

APTROOT, A., 1995 — Redisposition of some species excluded from *Didymosphaeria* (Ascomycotina). — Nova Hedwigia 60: 325–379.

ARX, J.A. VON & GAMS, W., 1967 — Über Pleurage verruculosa und die zugehörige Cladorrhinum-Konidienform. — Nova Hedwigia 13: 199–208.

BARRON, G. & SZIJARTO, 1982 - Structure and biology of

- a new hyphomycete parasitic on rotifers. Canad. J. Bot. 60: 1212-1915.
- BARRON, G. & SZIJARTO, 1984 Fungal parasites of bdelloid rotifers: A new *Phialophora*. — Mycologia 76: 1107–1110.
- BELL, A. & MAHONEY, D. P., 1995 Coprophilous fungi from New Zealand. I. *Podospora* species with swollen agglutinated perithecial hairs. — Mycologia 87: 375– 396.
- BEYMA THOE KINGMA, F. H. VAN, 1943 Beschreibung der im Centraalbureau voor Schimmelcultures vorhandenen Arten der Gattungen *Phialophora* Thaxter und *Margarinomyces* Laxa, nebst Schlüssel zu ihrer Bestimmung. — Antonie van Leeuwenhoek 9: 51–76.
- BORELLI, D., 1980 Causal agents of chromoblastomycosis (chromomycetes). Proc. Fifth Int. Conf. on Mycoses. — PanAmer. Health Org. Scient. Publ. No. 396: 334-335.
- BRYAN, G. T., DANIELS, M. J. & OSBOURN, A. E., 1995 Comparison of fungi within the *Gaeuman-nomyces-Phialophora* complex by analysis of ribosomal DNA sequences. — Appl. Environ. Microbiol. 61: 681–689.
- CHECA, J., BARRASA, J.M., MORENO, G., FORT, F. & GUARRO, J., 1988 — The genus Coniochaeta (Sacc.) Cooke (Coniochaetaceae, Ascomycotina) in Spain. — Cryptog. Mycol. 109: 1–34.
- CHEN, W., GRAY, L. E. & GRAU, C. R., 1996 Molecular differentiation of fungi associated with brown stem rot and detection of *Phialophora gregata* in resistant and susceptible soybean cultivars. — Phytopathology 86: 1140-1148.
- CIFERRI, R., 1958 Osservazioni intorno a tre fungilli dell'Olivo. — Sydowia 11: 286–289.
- COLE, G. T. & KENDRICK, W. B., 1973 Taxonomic studies of *Phialophora*. — Mycologia 65: 661–688.
- CONSTANTINESCU, O., HOLM, K. & HOLM, L., 1995 Teleomorph—anamorph connections in Ascomycetes: the anamorphs of three species of *Chaetosphaeria*. — Mycol. Res. 99: 585–592.
- CROUS, P. W. & GAMS, W., 2000 Phaeomoniella chlamydospora gen. et comb. nov., the causal organism of Petri grapevine decline. — Mycol. Mediterr. 38 (in press).
- CROUS, P. W., GAMS, W., WINGFIELD, M. J. & VAN WYK, P. S., 1996 — *Phaeoacremonium* gen. nov. associated with wilt and decline diseases of woody hosts and human infections. — Mycologia 88: 786–796.
- CUNNINGHAM, P.C., 1981 Isolation and culture. In: ASHER, M.J.C. & SHIPTON, P.J. (eds.): Biology and control of take-all. pp. 103–123. — Academic Press, London.
- DEACON, J. W., 1973 Phialophora radicicola and Gaeumannomyces graminis on roots of the grasses and cereals. — Trans. Brit. Mycol. Soc. 61: 471–485.
- DEACON, J. W., 1974 Further studies on *Phialophora radicicola* and *Gaeumannomyces graminis* on roots and stem bases of grasses and cereals. Trans. Brit.

- Mycol. Soc. 63: 307-327.
- DUPONT, J., LALOUI, W. & ROQUEBERT, M.-F., 1998 Partial ribosomal DNA sequences show an important divergence between *Phaeoacremonium* species isolated from *Vitis vinifera*. — Mycol. Res. 102: 631–637.
- EMDEN, J. H. VAN, 1975 Three new fungi from Surinam soil. — Acta Bot. Neerl. 24: 193-197.
- GAMS, W. & BOEKHOUT, T., 1985 Pigment localization in dematiaceous hyphomycetes and the segregation of *Pseudogliomastix* gen. nov. from *Acremonium*. — Proc. Indian Acad. Sci. (Pl. Sci.) 94: 273–280.
- GAMS, W. & HOLUBOVÁ-JECHOVÁ, V., 1976 Chloridium and some other dematiaceous hyphomycetes growing on decaying wood. — Stud. Mycol. 13: 1–99.
- GAMS, W. & MCGINNIS, M. R., 1983 Phialemonium, a new anamorph genus intermediate between Phialophora and Acremonium. — Mycologia 75: 977–987.
- GERRITS VAN DEN ENDE, A.H.G. & DE HOOG, G.S., 1999
 Variability and molecular diagnostics of the neurotropic species Cladophialophora bantiana. Stud. Mycol. 43: 151–162.
- GREENLEAF, M.A. & KORF, R.P., 1980 Mollisia in Macaronesia: an exercise in frustration. — Mycotaxon 10: 459-472.
- GROVES, J.W. & WILSON, D.E., 1967 The nomenclatural status of Coryne. Taxon 16: 35–41.
- HAASE, G., SONNTAG, L., MELZER-KRICK, B. & DE HOOG, G.S., 1999 — Phylogenetic inference by SSU-gene analysis of members of the *Herpotrichiellaceae* with special reference to human pathogenic species. — Stud. Mycol. 43: 80-97.
- HARNEY, S.K., ROGERS, S.O. & WANG, C.J.K., 1997 Molecular characterization of dematiaceous root endophytes. — Mycol. Res. 101: 1397–1404.
- HAWKSWORTH, D.L. & UDAGAWA, S., 1977 Contributions to a monograph of *Microthecium*. Trans. Mycol. Soc. Japan 18: 143–154.
- HELFER, W., 1991 Pilze auf Pilzfruchtkörpern. Untersuchungen zur Ökologie, Systematik und Chemie. — Libri Bot. 1: 1–157 pp.
- HOOG, G.S. DE, 1977 The black yeasts and allied hyphomycetes: *Rhinocladiella* and allied genera. — Stud. Mycol. 1–140.
- HOOG, G.S. DE, WEENINK, X.O. & GERRITS VAN DEN ENDE, A.H.G., 1999 — Taxonomy of the *Phialophora* verrucosa complex with the description of two new species. — Stud. Mycol. 43: 107–122.
- HORIE, Y., UDAGAWA, S. & CANNON, P.F., 1986 Four new Japanese species of the *Ceratostomataceae* (Ascomycetes). — Mycotaxon 25: 229–245.
- IWATSU, T., UDAGAWA, W. & TOYAZAKI, N., 1988 A new species of *Phialophora*. — Mycotaxon 32: 439-445.
- JONES, E. B. G., HYDE, K. D., READ, S. J., MOSS, S. T. & ALIAS, S. A., 1996 — Tirisporella gen. nov., an ascomycete from the mangrove palm Nypa fruticans. — Can. J. Bot. 74: 1487-1495.
- KENDRICK, W.B., 1961 The Leptographium complex.

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- Phialocephala gen. nov. Canad. J. Bot. 39: 1079–1085.
- KORF, R.P., PEARSON, R.C., ZHUANG, W.-Y., DUBOS, B., 1986 — Pseudopezicula (Helotiales, Peziculoideae), a new discomycete genus for pathogens causing an angular leaf scorch disease of grapes ("Rotbrenner"). — Mycotaxon 26: 457–471.
- LANDSCHOOT, P.J. & JACKSON, N., 1989 Magnaporthe poae sp. nov., a hyphopodiate fungus with a Phialophora anamorph from grass roots in the United States. — Mycol. Res. 93: 59-62.
- LE GAL, M. & MANGENOT, F., 1960 Contribution à l'étude des Mollisioïdées, III. — Rev. Mycol. 25: 135– 214
- LE GAL, M. & MANGENOT, F., 1961 Contribution à l'étude des Mollisioïdées IV. — Rev. Mycol. 26: 263– 331.
- LUNDQVIST, N., MAHONEY, D.P., BELL, A. & LORENZO, L.E., 1999 — Podospora austrohemisphaerica, a new heterothallic ascomycete from dung. — Mycologia 91: 405–415.
- MASCLAUX, F., GUÉHO, E., DE HOOG, G.S. & CHRISTEN, R., 1995 — Phylogenetic relationships of humanpathogenic Cladosporium (Xylohypha) species inferred from partial LS rRNA sequences. — J. Med. Vet. Mycol. 33: 327–338.
- MATSUSHIMA, T., 1983 Matsushima Mycological Memoirs No. 3. Published by the author, Kobe.
- McGinnis, M.R., 1977 Wangiella, a new genus to accommodate Hormiscium dermatitidis. — Mycotaxon 5: 341–352.
- MCKEMY, J. M., ROGERS, S. O. & WANG, C. J., 1999 Emendation of the genus Wangiella and a new combination, W. heteromorpha. — Mycologia 91: 200–205.
- MOUCHACCA, J. & GAMS, W., 1993 The hyphomycete genus Cladorrhinum and its teleomorph connections. — Mycotaxon 48: 415–440.
- NAGAI, K., SAKAI, T., RANTIATMODJO, R. M., SUZUKI, K., GAMS, W. & OKADA, G., 1995 — Studies on the distribution of alkalophilic and alikali-tolerant soil fungi I. — Mycoscience 36: 247–256.
- NAKAGIRI, A., 1993 A new marine ascomycete in Spathulosporales, Hispidocarpomyces galaxauricola gen. et sp. nov. (Hispidocarpomycetaceae fam. nov.), inhabiting a red alga, Galaxaura falcata. — Mycologia 85: 638-652.
- NAKAGIRI, A. & ITO, T., 1997 Retrostium amphiroae gen. et sp. nov. inhabiting a marine red alga, Amphiroa zonata. — Mycologia 89: 484–493.
- PETRINI, L.E., PETRINI, O. & CANDOUSSAU, F., 1984 Anamorphs of Ascomycetes: The *Phialophora*-like state of *Eosphaeria uliginosa* (syn. *Herminia dichroospora*). — Trans. Brit. Mycol. Soc. 82: 554–556.
- RAMALEY, A. W., 1997 Barrina, a new genus with polysporous asci. — Mycologia 89: 962–966.
- RASCHLE, P., 1977 Taxonomische Untersuchungen an Ascomyceten aus der Familie der Hyaloscyphaceae Nannfeldt. — Sydowia 29: 170–236.

- RÉBLOVÁ, M., 1999 Studies in Chaetosphaeria I. The genera Chaetosphaerella and Tengiomyces gen. nov. of the Helminthosphaeriaceae. — Mycotaxon 70: 387–420.
- RÉBLOVÁ, M., BARR, M. E. & SAMUELS, G.J., 1999 Chaetosphaeriaceae, a new family for Chaetosphaeria and its relatives. — Sydowia 51: 49-70.
- RÉBLOVÁ, M. & WINKA, K., 2000 Phylogeny of Chaetosphaeria and its anamorphs based on morphological and molecular data. — Mycologia (in press).
- REHNER, S.A. & SAMUELS, G.J., 1995 Molecular systematics of the *Hypocreales*; a teleomorph gene phylogeny and the status of their anamorphs. Canad. J. Bot. 73 (Suppl.): S816-S823.
- ROGERS, S.O., MCKEMY, J.M. & WANG, C.J.K., 1999 Molecular assessment of *Exophiala* and related hyphomycetes. — Stud. Mycol. 43: 122–133.
- ROLL-HANSEN, F. & ROLL-HANSEN, H., 1979 Ascocoryne sarcoides and Ascocoryne cylichnium. Description and comparison. — Norw. J. Bot. 26: 193–206.
- ROMERO, A.I., CARMARÁN, C.C. & LORENZO, L.E., 1999
 A new species of *Coniochaeta* with a key to the species known in Argentina. Mycol. Res. 103: 689-695.
- SAMUELS, G. J. & BUCHANAN, D. E., 1983 Ascomycetes of New Zealand 5. Mycocalicium schefflerae sp. nov., its ascal ultrastructure and Phialophora anamorph. — New Zealand J. Bot. 21: 163–170.
- SAMUELS, G.J. & RODRIGUES, K.F., 1989 Batistia annulipes and its anamorph, Acrostroma annellosynnema. — Mycologia 81: 52–56.
- SCHOL-SCHWARZ, M. B., 1970 Revision of the genus Phialophora (Moniliales). — Persoonia 6: 59–94.
- SCOTT, D.B. & DEACON, J.W., 1983 Magnaporthe rhizophila sp. nov., a dark mycelial fungus with a Phialophora conidial state, from cereal roots in South Africa. — Trans. Brit. Mycol. Soc. 81: 77–81. 1983.
- SEIFERT, K.A., 1987 Stromatographium and Acrostroma gen. nov.: two tropical hyphomycete genera with distinctive synnema anatomies. — Canad. J. Bot. 65: 2196–2201.
- SEIFERT, K.A., 1989 Coryne trichophora, comb. nov., and the implications of its conidiomatal anatomy. — Stud. Mycol. 31: 157-164.
- SIVASITHAMPARAM, K., 1975 Phialophora and Phialophora-like fungi occurring in the root region of wheat. — Aust. J. Bot. 23: 193–212.
- UDAGAWA, S. & MUROI, T., 1981 Notes on some Japanese Ascomycetes VI. — Trans. Mycol. Soc. Japan 22: 11–26.
- UIJTHOF, J.M.J., 1996 Relationships within the black yeast genus *Exophiala* based on ITS1 sequences. — Mycol. Res. 100: 1265–1271.
- UIJTHOF, J.M.J., FIGGE, M.J. & DE HOOG, G.S., 1997 Molecular and physiological investigations of Exophiala species described from fish. — Syst. Appl. Microbiol. 20: 585-594.
- UNTEREINER, W. A., 1995 Fruiting studies in species of

- Capronia (Herpotrichiellaceae). Antonie van Leeuwenhoek 68: 3-17.
- UNTEREINER, W. A. & NAVEAU, F. A., 1999 Molecular systematics of the Herpotrichiellaceae with an assessment of the phylogenetic positions of Exophiala dermatitidis and Phialophora americana. — Mycologia 91: 67–83.
- UNTEREINER, W. A., STRAUS, N.A. & MALLOCH, D., 1995
 A molecular-morphotaxonomic approach to the systematics of the *Herpotrichiellaceae* and allied black yeasts. Mycol. Res. 99: 897–913.
- WALKER, J., 1980 Gaeumannomyces, Linocarpon, Ophiobolus and several other genera of scolecospored Ascomycetes and Phialophora conidial states, with a note on hyphopodia. — Mycotaxon 11: 1–129.
- WALKER, J., 1981 Taxonomy of take-all fungi and related genera and species. — In: ASHER, M.J.C. & SHIPTON, P.J. (eds.): Biology and control of take-all. pp. 15–84. — Academic Press, London.
- WANG, C.J.K. & WILCOX, H.E., 1985 New species of ectendomycorrhizal and pseudomycorrhizal fungi: Phialophora finlandica, Chloridium paucisporum, and Phialocephala fortinii. — Mycologia 77: 951–958.
- WARD, E. & BATEMAN, G. L., 1999 Comparison of Gaeumannomyces- and Phialophora-like fungal pathogens from maize and other plants using DNA methods. — New Phytol. 141: 323–331.
- WINGFIELD, M.J., SCHALK VAN WYK, P. & WINGFIELD, B.D., 1987 — Reclassification of *Phialocephala* based on conidial development. — Trans. Brit. Mycol. Soc. 89: 509–520.
- WONG, P.T.W. & WALKER, J., 1975 Germinating phialidic conidia of *Gaeumannomyces graminis* and *Phia-lophora*-like fungi from Gramineae. — Trans. Brit. Mycol. Soc. 65: 41–47.
- YAMAMOTO, H., KOBAYASHI, K. & OGOSHI, A., 1992 Genetic relatedness between *Phialophora gregata* f. sp. adzukicola and *Phialophora gregata* f. sp. sojae. — Trans. Mycol. Soc. Japan 33: 461–465.
- YAN, Z. H., ROGERS, S. O. & WANG, C. J. K., 1995 Assessment of *Phialophora* species based on ribosomal DNA internal transcribed spacers and morphology. — Mycologia 87: 72–83.