## BIONECTRIACEAE Samuels & Rossman, fam. nov.

Type: Bionectria Speg., Bol. Acad. Nac. Ci. 23: 563. 1919.

[= Seliniaceae Arx & E. Müll., Acta Bot. Neerl. 4: 121. 1955, nom. inval., Art. 36.1]

Ascomata perithecialia, raro cleistothecialia, candida, flava, aurantiaca, spadicea vel fusca, nec KOH nec acido lactico mutantia, centrum typi *Nectriae*.

Ascomata perithecial, rarely cleistothecial, white, yellow, orange to tan or brown, not changing color in KOH or lactic acid, centrum of *Nectria*-type.

The Bionectriaceae are characterized by having uniloculate perithecial, rarely cleistothecial, ascomata that are pallid, ranging in color from white, yellow, orange to tan or brown, do not change color in KOH or lactic acid, and are generally superficial, lacking a stroma, or are immersed in the substratum. The *Bionectriaceae* include 26 genera. Five of the six cleistothecial hypocrealean genera, namely *Battarrina*, *Emericellopsis*, *Heleococcum*, *Mycoarachis*, and *Roumegueriella*, are placed in the *Bionectriaceae*. These cleistothecial genera lack the centrum characteristics that place them definitively in the *Hypocreales* but the ascomatal wall structure and anamorph suggest the *Bionectriaceae*. Except for *Battarrina* which is as yet unstudied, molecular studies have confirmed the affinities of these genera with the *Bionectriaceae* (Rehner & Samuels, 1994, 1995; Ogawa *et al.*, 1997; Spatafora & Blackwell, 1993).

## KEY TO THE GENERA OF THE BIONECTRIACEAE

1.	Ascomata cleistothecial, globose, without an organized hymenium; asci generally globose, clavate in <i>Heleococcum</i>
1.	Ascomata perithecial, subglobose to pyriform, with organized hymenium; asci elongate
<b>2</b> (I).	Ascomata dark olive-green to black or hyaline, but appearing brownish due to pale
2.	Ascomata hyaline to pale gray, bright yellow or reddish brown
3 (2). 3.	Ascospores broadly ellipsoid, 1-septate, smooth-walled
4 (2).	Ascospores ellipsoid, 1-septate, smooth to ornamented with wings; anamorph Acremo- nium-like or unknown
4.	Ascospores globose, non-septate, ornamented with sharp, pointed spines
5 (4).	Ascomata on sporocarps of <i>Tuber</i> ; ascospores 4-6 µm diam, with sparse echinulations; anamorph unknown
5.	Ascomata on dung, compost, damp paper and various kinds of organic detritus; ascospores more than 6 µm diam, densely echinulate; anamorph, where known, Gliocladium-like
<b>6</b> (1).	Ascomata immersed in the substratum or in a stroma that may itself be immersed in the
6.	Ascomata superficial or immersed in a hyphal subiculum or thin stroma
7 (6).	Ascomata immersed in a stroma that may itself be immersed in the substratum; ascospores non- to one-septate; on dung, corticolous, or herbicolous, not lichenicolous or fungicolous
7.	Ascomata immersed in substratum, non-stromatic, usually solitary; ascospores non- to multiseptate or muriform; corticolous, herbicolous, fungicolous or lichenicolous, not on dung

8 (7).	Ascospores non-septate, more than 40 µm long, hyaline, smooth, thick-walled; on dung
8.	Ascospores 1-septate, less than 20 µm long; herbicolous or corticolous
9 (8). 9.	Ascomata in a stroma immersed in bambusoid grasses or in living or dead wood; ascospores hyaline or yellow-brown, coarsely striate with age
10 (7). 10.	On algae or wood in marine habitats; ascomata without or each with a long neck 11 Terrestrial or in freshwater habitats; ascomata without a long neck 12
11 (10). 11.	Ascomata each with a long neck, immersed in algae or wood in marine habitats; ascospores long-fusiform, non-septate, smooth-walled
12 (10). 12.	Ascomata immersed in herbaceous tissue, bark or wood, rarely fungicolous; anamorphs Acremonium or Kutilakesa
13 (6). 13.	Ascospores with long, attenuated ends; on lichens or algae
14 (13). 14.	Ascomata with straight, solitary hairs; ascospores 1- to multiseptate Trichonectria Ascomata without hairs or, if present, hairs fasciculate or flexuous; ascospores generally 1-septate
15 (14). 15.	Ascomata usually superficial on a thin subiculum; anamorph Gliocladium; on Aphyllophorales
16 (15). 16.	Ascomata immersed in a stroma or superficial with white to tan or green hyphae covering the ascomatal wall
17 (16). 17.	Ascomata immersed, loosely united in a thin, pseudoparenchymatous stroma; ascomata less than 160 µm diam, walls less than 10 µm thick; ascospores narrowly cylindrical, smooth-walled
<b>18</b> (17).	Ascomata immersed in an effused hyphal stroma; ascospores generally striate, less often smooth or tuberculate; anamorphs Acremonium-like; usually on monocotyledonous plant debris  Ascomata immersed in an effused hyphal stroma or superficial with tan hyphae covering the ascomatal wall; ascospores spinulose or verrucose; anamorphs synnematous
	(Stilbella) or Acremonium-like; corticolous, sometimes on other ascomycetes

19 (16). 19.	Ascomata white to pale yellow, small, generally < 200 µm diam, wall thick, smooth or covered with flexuous hairs; ascospores smooth or spinulose, rarely striate; anamorphs, where known, Acremonium-like, Gliocladium-like, or Septofusidium; fungicolous, on myxomycetes, pyrenomycetes and dematiaceous hyphomycetes, rarely on Aphyllophorales, or on liverworts and mosses
<b>20</b> (19). <b>20.</b>	Ascomata on liverworts or mosses
<b>21</b> (20).	Ascomata on Asterina, Meliola, Schnifferula, growing on superficial, black hyphae
21.	covering living leaves
<b>22</b> (19).	Ascomata globose to subglobose, occasionally doliiform, becoming cupulate upon drying; ascomatal wall of globose, thin-walled cells; ascospores often striate: anamorphs Acremonium-like
22.	Ascomata globose to subglobose or ovoidal, generally not cupulate upon drying; ascomatal wall of thick-walled cells; ascospores smooth, spinulose or striate; anamorphs Acremonium-like, Clonostachys or Didymostilbe
<b>23</b> (22).	Ascomata of three regions, with orange oil droplets in the middle region of the wall  Ochronectria
23.	Ascomata of two regions, without orange oil droplets Hydropisphaera
24 (22). 24.	Ascomata with a flattened apex, often with solitary or fasciculate hairs forming an apical fringe; ascospores striate or spinulose; anamorphs <i>Acremonium</i> -like or unknown 25 Ascomata without distinct hairs, smooth to warted or with short, hyphal hairs; ascospores smooth, spinulose or rarely striate; anamorphs <i>Clonostachys</i> or <i>Didymostilbe</i> 26
<b>25</b> (24).	Ascomata yellow-brown to dark brown, globose to subglobose, with solitary stiff or hyphal hairs, not forming a distinct fringe; ascospores striate or spinulose; anamorphs
25.	Acremonium; herbicolous or corticolous
<b>26</b> (24).	golden brown, smooth or faintly striate; anamorph Fusarium
26.	Ascomata white, bright yellow, orange, tan to brown, smooth, scurfy or with large, white warts; ascospores hyaline, spinulose or striate; anamorph Clonostachys or Didymostilbe
<b>27</b> (26).	Ascomata bright- to dark yellow, with walls over 50 µm thick; ascospores broadly reniform to broadly fusiform, with rounded ends, more than 30 µm long; anamorph
27.	synnematous, Didymostilbe

## THE GENERA OF THE BIONECTRIACEAE

BATTARRINA (Sacc.) Clem., in Clem. & Shear, Genera of Fungi p. 279. 1931.

≡ Hypocrea subgenus Battarrina Sacc., Syll. Fung. 2: 533.
1883.

Type: Battarrina inclusa (Berk. & Broome) Clem. (≡ Hypocrea inclusa Berk. & Broome).

Ascomata immersed in host tissue, forming in discolored areas, non-ostiolate, hyaline to pale yellow, globose, about 250 µm diam, walls thin, membranous. Asci evanescent, scattered, without an organized hymenium. Ascospores globose to angular, non-septate, hyaline, sparsely echinulate. Anamorph unknown. On fruiting bodies of *Tuber*.

Notes.— Battarrina was established as a subgenus of Hypocrea for species with globose ascospores. Although three species, H. inclusa, H. cervina Berk. & M.A. Curtis, and H. cerebriformis Berk., were included in the subgenus, the taxon was raised to generic rank with only B. inclusa and remains a unispecific genus. Benny & Kimbrough (1980) included Battarrina in the Hypocreales based on the literature.

Battarrina inclusa (Berk. & Broome) Clem., in Clem. & Shear, Genera of Fungi p. 279. 1931.

≡ Hypocrea inclusa Berk. & Broome, Ann. Mag. Nat. Hist., Ser. 3, 7: 461. 1861.

Ascomata about 250  $\mu$ m diam, cleistothecial, hyaline to pale yellow, membranous, thin-walled, wall structure of parallel hyphae up to 4  $\mu$ m diam. Asci cylindrical, 30–33  $\times$  5–6  $\mu$ m, clavate when young, evanescent at maturity. 8-spored, ascospores uniseriate. Ascospores globose to slightly angular, 4–6  $\mu$ m diam, hyaline, thin-walled with sparse, slender echinulations about 1  $\mu$ m tall.

Habitat.— Parasitic in ascomata of *Tuber puberulum* Berk. & Broome.

DISTRIBUTION. - England.

HOLOTYPE.— ENGLAND. Bristol, Leigh Wood, Sept. 1859, C.E. Broome (K).

ADDITIONAL SPECIMENS EXAMINED.— ENGLAND. N. Bristol, Nov. 1869, C.E. Broome 2335 (K); Hanham near Bristol, Nov. 1871, C.E. Broome, Rabenhorst, Fungi Europ. 1610 (BPI, K, UPS).

ILLUSTRATIONS.— Berkeley & Broome (I.c., 1861, Fig. 23);
Hawker (1955, Fig. 2); Petch (1938, Fig. 2).

Notes.— The type specimen of Battarrina inclusa was

examined and found to be in poor condition apparently having been stored in liquid preservation and subsequently dried. The two sections of the host ascomata of Tuber puberulum contain indistinct ascomata of B. inclusa, appearing on the cut surface as glazed, pale-yellow areas. These areas are packed with asci and ascospores. The specimens of Fungi Europaei 1610 were also in poor condition. Petch (1938) included B. inclusa in his account of the British Hypocreales, based on the type description. Hawker (1955) discussed the species based on a fresh collection from Great Britain that was not located. Due to the poor condition of the type specimen and lack of additional specimens, many characteristics of B. inclusa remain obscure. The description given here includes details from Hawker (1955) and Petch (1938).

BIONECTRIA Speg., Bol. Acad. Nac. Ci. 23: 563. 1919.

Type: B. tonduzii Speg.

Ascomata solitary to gregarious, superficial to slightly immersed in an erumpent stroma, often occurring on other fungi. Ascomata white, yellow, pale orange, tan or brown, KOH-, wall smooth to warted or with scales, thin-walled hairs or flexuous setae, subglobose or globose to ovoid, when dried not collapsing or collapsing irregularly, ostiolate. Asci narrowly clavate to clavate. Ascospores 1-septate, rarely multiseptate, hyaline, smooth to spinulose or slightly warted. Anamorph Clonostachys. On leaves and decaying woody substrata.

Notes.— At present Bionectria includes species formerly placed in the Nectria ochroleuca-group (Samuels, 1976a; Rossman, 1983; Schroers & Samuels, 1997), but it may also include species placed in the N. ralfsii-group (Samuels, 1976a), the N. muscivoragroup (Rossman, 1983; Samuels, 1988), and species of Nectria having Sesquicillium anamorphs (Samuels, 1989a). Apart from having pallid perithecia, these species are similar in gross morphology and wall anatomy of the perithecium (Schroers & Samuels, 1997; Schroers et al., 1999). Most species occur on woody substrata and are readily grown on agar. Anamorphs of species of Bionectria are useful in distinguishing species (Schroers et al., 1999) and are classified primarily in Clonostachys (syn. Dendrodochium) or Myrothecium-like, the groups differing from each other by conidial color, viz. salmon in Clonostachys, and dark

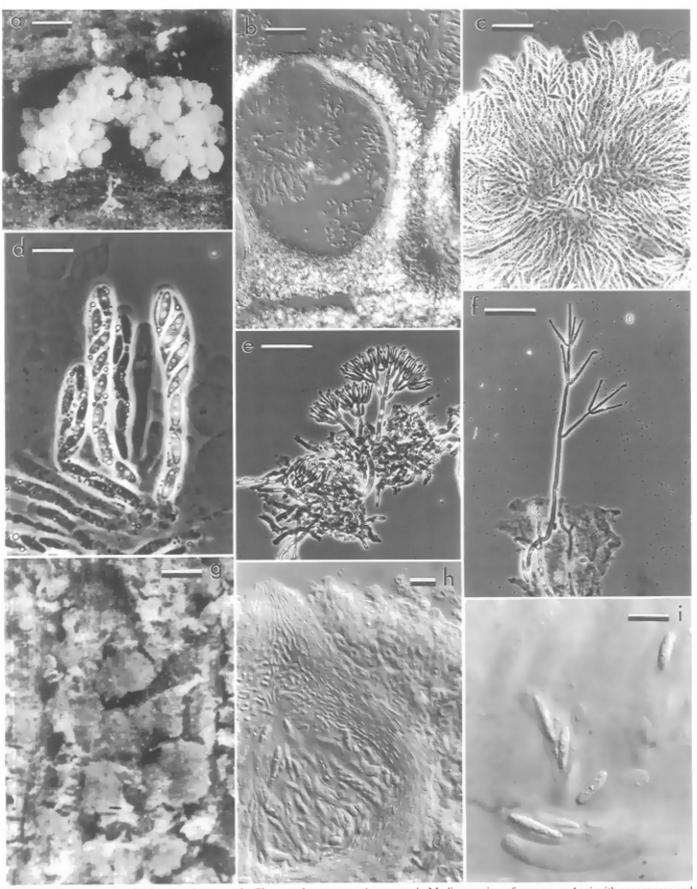


Plate 1. a–f. *Bionectria ochroleuca* and anamorph, *Clonostachys rosea*. a. Ascomata. b. Median section of ascoma. c. Asci with ascospores and inflated cells of apical paraphyses. d. Asci with ascospores. e. Penicillate conidiophores of anamorph. f. Verticillate conidiophore of anamorph. g–i. *Clibanites paradoxa*. g. Stroma with ascomata. h. Median section of ascoma. i. Ascospores. a–d. BPI 749158. e–f. Anamorph grown from BPI 737784. g–i. Holotype of *Clibanites paradoxa* – H. Scale bars: a = 500 μm; b, c, f = 50 μm; c = 25 μm; d, h, i = 10 μm; g = 250 μm.

green in *Myrothecium*-like anamorphs. *Bionectria* ochroleuca, particularly as its anamorph *Clonostachys* rosea, is one of the most commonly encountered soil fungi and is often cited in the biological control literature as a destructive mycoparasite (Schroers et al., 1999).

Bionectria tonduzii Speg., Bol. Acad. Nac. Ci. 23: 563. 1919 [as 'tonduzi'].

≡ Nectria tonduzii (Speg.) Samuels, Mem. New York Bot. Garden 48: 22. 1988 [as 'tonduzi'].

Mycelium tan, spreading. Ascomata seated in mycelium, solitary to caespitose in groups of up to 15, partially immersed in an erumpent, granular stroma, globose, 250-280 µm diam, non-papillate, wall warted, pale orange, KOH-, warts lighter, ascomata collapsing by lateral pinching. Surface of ascomatal wall of nearly circular cells in outline, 12-25 µm diam, walls slightly thickened, 2-4 µm. Ascomatal wall about 50 µm thick, of two regions: outer region about 25 µm thick, of cells circular to slightly elongate, 12-25 µm diam, forming warts up to 50 μm high; inner region about 25 μm thick, of cells flattened to fusiform, 10-15 × 2-3.5 μm. Ascomatal apex of narrow, about 1 µm wide hyphal elements arising from the inner ascomatal wall. Asci clavate,  $(62-)64-81(-85) \times (11-)12.5 \times 15 \mu m$ , apex broad, simple, 8-spored, ascospores entirely or partially biseriate. Ascospores fusiform, (16-)19.5-24.5(-27) × (5-)5.5-6.5(-7) μm, 1-septate, hyaline, smooth to spinulose.

Anamorph.— None known.

Habitat and distribution.— Known only from the type collection.

HOLOTYPE.— COSTA RICA. San José: on leaves of *Byttneria* carthaginensis, possibly parasitizing ascomata of *Puiggarina* costaricensis Syd. (LPS).

ILLUSTRATIONS.— Samuels (1988, Figs. 5 a, b, as N. 'ton-duzi'); Schroers & Samuels (1997, Fig. 6).

Notes.— Bionectria was established for species that are like Nectria but occur on living plant parts. The type species of Bionectria, B. tonduzii, occurs on living leaves of Byttneria carthaginensis Jacq. (Sterculiaceae), possibly parasitizing ascomata of Puiggarina costaricensis Syd. Samuels (1988) redescribed and illustrated this species based on an examination of the type specimen. Bionectria tonduzii has never been cultured or linked to an anamorph.

The four species of *Bionectria* listed below in addition to the type were recently treated by Schroers & Samuels (1997) including color illustrations and references to recent descriptions.

**Bionectria apocyni** (Peck) Schroers & Samuels, Z. Mykol. 63: 153. 1997.

≡ Nectria apocyni Peck, Bull. Buffalo Soc. Nat. Sci. 1: 71.
1873.

Bionectria aureofulva (Cooke & Ellis) Schroers & Samuels, Z. Mykol. 63: 153. 1997.

■ Nectria aureofulva Cooke & Ellis, Grevillea 7: 8. 1878.

Bionectria byssicola (Berk. & Broome) Schroers & Samuels, Z. Mykol. 63: 152. 1997.

≡ Nectria byssicola Berk. & Broome, J. Linn. Soc. Bot.
14: 116. 1873.

Bionectria ochroleuca (Schwein.) Schroers & Samuels, Z. Mykol. 63: 151. 1997. — Plate 1, a-f.

≡ Sphaeria ochroleuca Schwein., Trans. Amer. Philos. Soc., N.S. 4: 204. 1834.

≡ Nectria ochroleuca (Schwein.) Berk., Grevillea 4: 16.
1875.

Anamorph: Clonostachys rosea (Link: Fr.) Schroers et al., Mycologia 91: 369. 1999.

A detailed account of *B. ochroleuca* and its *Clonostachys rosea* anamorph is given by Schroers *et al.* (1999).

Specimens examined.— MEXICO. Veracruz: Laguna Verde, on dead leaves of *Yucca* sp., 2 Oct 1994, G. Bills, G.J.S. 94-122 (BPI 737784); PUERTO RICO. Caribbean National Forest: Luquillo Mts., Bisley Experimental Watershed, on dead bark of *Mangifera indica*, elev. 300 m, 21 Feb 1996, G.J. Samuels, H.J. Schroers (H.J.S. 82), D.J. Lodge, det. H.J. Schroers (BPI 749158).

## KEY TO THE SPECIES OF BIONECTRIA

1.	Ascospores more than 15 µm long, warted
1.	Ascospores generally less than 15 $\mu m$ long, smooth, warted or spinulose
2.	On living leaves, possibly associated with stromatic fungi; ascomata with warts up to 50 µm
	high; ascospores 19.5–24.5 × 5.5–6.5 μm
2.	On decaying bark or wood; ascomata smooth; ascospores 16-33 × 4.5-9.5 µm
	B. apocyni
3.	Ascomata orange with conspicuous white warts; ascospores ellipsoid, (10–)11–14(–16) × 4–5(–6) µm, smooth or spinulose
3.	Ascomata orange to brown, smooth to slightly scaly or covered with a thin layer of hyphae
4.	Ascomata smooth; ascospores 8.5–15 × 2.5–5 μm, spinulose or warted B. aureofulva
	Ascomata slightly scaly or covered with a thin layer of hyphae; ascospores 7.5-14.5 ×
	2.5-4.5 µm, slightly spinulose

## BRYONECTRIA Döbbeler, Nova Hedwigia 66: 334. 1998.

Type: B. hylocomii (Döbbeler) Döbbeler (≡ Nectria hylocomii Döbbeler, Mitt. Bot. Staatssamml. München 14: 78. 1978).

Ascomata superficial, with hyphae penetrating the host cells, solitary or rarely aggregated, non-stromatic, globose to obpyriform, 80–200 µm diam, hyaline to white, not changing color in KOH or lactic acid or rarely reacting. Smooth or with short setae. Ascomatal wall of thick-walled cells. Asci ellipsoid to cylindric, with or without an apical ring. Ascospores ellipsoid, 1- or 2-septate, hyaline, often with a guttule in each cell. Anamorph unknown. Parasitic on liverworts and mosses.

Notes.— Bryonectria was described to accommodate six species of hypocrealean fungi that occur on foliose liverworts and mosses.

## CLIBANITES P. Karst., Bidrag Kännedom Finlands Natur Folk 19: 14, 1871.

≡ Peziza sect. Clibanites P. Karst., Monogr. Peziz, Fenn. p.
155. 1869. — Type: C. paradoxa (P. Karst.) P. Karst. (≡ Peziza paradoxa P. Karst.).

Stroma of intertwined hyphae in the middle and at the base, with highly compacted hyphae near the surface, ascomata immersed in a stroma, loosely united in groups up to ten. Ascomata globose, dark yellow, non-papillate, apex not differentiated, ostiolar canal periphysate, not collapsed upon drying. Ascomatal wall ca 10 µm thick, of several layers of small, flattened cells. Asci subcylindrical, apex broad, blunt, with a ring, as-

cospores biseriate. Ascospores narrowly cylindrical, equally 2-celled, not constricted, hyaline, smooth. Anamorph unknown. On well-rotted wood of *Quercus*. Notes.— *Clibanites* is a unispecific genus originally described by Karsten as a discomycete. An examination of the type specimen reveals that, based on the small, thin-walled, pallid ascomata and non-disarticulating ascospores, *C. paradoxa* is similar to *Nectriopsis* in the *Bionectriaceae*. It differs from *Nectriopsis* in having relatively thick-walled ascomata loosely united in a common stroma and in the non-fungicolous habit.

Clibanites paradoxa P. Karst., Bidrag Kännedom Finlands Natur Folk 19: 14. 1871. — Plate 1, g-i, Plate 2, a.

≡ Peziza paradoxa P. Karst., Monogr. Peziz. Fenn. p. 155.
1869.

Stroma superficial on decorticated wood, evident as pallid scurf, dissected and squamose (possibly as an artifact of drying), entire stromal aggregate easy to remove; ascomata loosely united into groups of up to 10, immersed in a stroma, adjacent ascomata evident as slightly tuberculate, ostiolate areas, ascomata joined by a subiculum of smooth-walled, 2-3 μm wide, branched, septate hyphae with few free ends, thinwalled, hyaline in transmitted light. Stroma 25-30 μm thick, surface consisting of highly compacted, ca 3 μm wide hyphae; internally hyphae more loosely disposed. Ostioles visible as viscid dots against the dull background of the ascomatal wall. Ascomata globose, ca 100-160 μm diam, pale yellow, KOH-, non-papillate, not collapsed on drying, ostiolar canal periphysate. Ascomatal wall ca 10 µm thick, of one region of small,

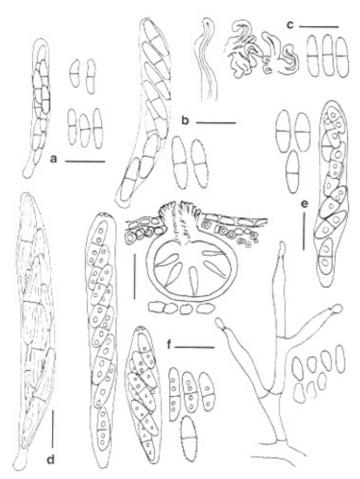


Plate 2. a. Clibanites paradoxa, asci and ascospores. b. Hydropisphaera rufofusca, ascus and ascospores. c. Ijuhya aquifolii, ascomata hairs and ascospores. d. Ijuhya chilensis, ascus. e. Lasionectria mantuana, asci and ascospores. f. Nectriella minuta, median section of ascoma, asci, ascospores, conidiophores and conidia. a. Holotype – H. b. Holotype of Nectriella rufofusca – PAD. c. Lectotype of Peziza aquifolii – BPI 1113199. d. Holotype of Lepidonectria chilensis – LPS. e. Holotype – PAD. f. Holotype – NY. Scale bars: a–f = 10 μm, except upper figure in f = 100 μm.

flattened cells. Asci subcylindrical,  $25\text{--}30 \times 4\text{--}5 \mu m$ , sessile, apex broad, blunt, with a ring, ascospores biseriate. Ascospores narrowly cylindrical,  $6\text{--}10 \times 1.5\text{--}2 \mu m$ , equally 2-celled, not constricted, hyaline, smooth. Habitat and distribution.— Known only from the type specimen.

HOLOTYPE.— FINLAND. Runsala: 'prope oppid. supra lignum *Quercus* vetustum', 26 May 1861, P. Karsten No. 3365 (H).

DIMEROSPORIELLA Speg., Revista Mus. La Plata 15: 10. 1908.

Type: D. paulistana Speg.

= Epinectria Syd. & P. Syd., Ann. Mycol. 15: 215. 1917. — Type: E. meliolae Syd. & P. Syd.

Mycelium white, cottony, often bearing conidia, hyphae septate, branching. Ascomata scattered, superficial on white mycelium or directly on black mycelium of the host fungus, usually easily removed from substratum, subglobose, globose to obovoid, often collapsing by lateral pinching, 100–245(–270) μm diam, pale yellow, KOH–, non-papillate, smooth or with short, flexuous hairs up to 25 μm long. Ascomatal wall thin, often about 10 μm thick, with wall of non-descript, small cells, often forming a textura epidermoidea. Asci clavate, usually less than 70 μm long, often with an apical ring, 8-spored. Ascospores ellipsoid, 1–3-septate, hyaline, smooth, spinulose or striate. Anamorph, where known, Acremonium-like. On black, thick-walled hyphae of Asterina, Meliola. Schiffnerula or related species on living leaves in tropical regions.

Notes.— Dimerosporiella is herein recognized for species that have previously been placed in the Nectria leucorrhodina-group (Samuels, 1976a; Rossman, 1983) or treated within Nectriopsis (Samuels, 1988). Spegazzini placed Dimerosporiella near Dimerosporium in the Englerulaceae differentiated by the presence of an ostiole. Petrak & Sydow (1934) examined the rather sparse type specimen of D. paulistana, presented a detailed description, and concluded that this species belonged in Nectria. Dimerosporiella paulistana represents a species additional to those previously placed in the N. leucorrhodina-group and/or Nectriopsis. The unispecific genus Epinectria was established for a species considered to be close to Hyalocrea but having elongate, one-septate ascospores. Several parts of the type specimen of E. meliolae were examined and the fungus was determined to be a synonym of Dimerosporiella pipericola. Seven species are included in Dimerosporiella differentiated primarily by ascomatal wall surface features and characteristics of the ascospores.

Dimerosporiella paulistana Speg., Revista Mus. La Plata 15: 10. 1908. — Plate 3, a-d.

Ascomata superficial, on black mycelium of *Schiff-nerula* and on the surrounding leaf tissue, obovoidal, minute, 117  $\mu$ m high  $\times$  80  $\mu$ m diam, pale yellow, ostiolate, thin-walled. Ascomatal wall ca 10  $\mu$ m thick, unpigmented, of *textura epidermoidea*. Asci clavate, apex thickened, with a ring, spent asci with open tops following ascospore discharge, ascospores biseriate. Ascospores ellipsoid, 12–14  $\times$  4–4.5  $\mu$ m, 1-septate, hyaline, smooth.

HABITAT. - Known only from type specimen.

HOLOTYPE.— BRAZIL. São Paulo: Ipiranga Moça, on wilting leaves of *Buddleja* sp., Sep 1905, A. Uster, No. 143, det. C. Spegazzini, No. 402 (LPS).

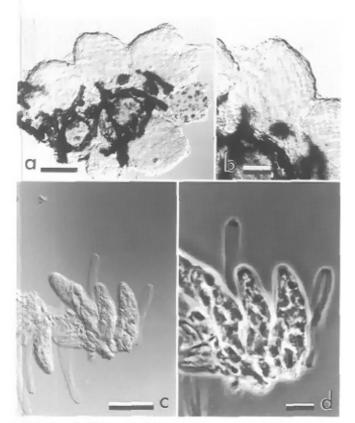


Plate 3. a-d. Dimerosporiella paulistana. a-b. Translucent, thinwalled ascomata on black, thick-walled host hyphae. c, d. Asci with ascospores and overmature asci. a-d. Holotype – LPS. Scale bars: a = 50 μm; b, c = 25 μm; d = 10 μm

Additional species of Dimerosporiella:

# Dimerosporiella cephalosporii (Hansford) Rossman & Samuels, comb. nov.

- ≡ Calonectria cephalosporii Hansford, Mycol. Pap. 15: 117, 1946.
- ≡ Nectriopsis cephalosporii (Hansford) Samuels, Mem. New York Bot. Gard. 48: 38. 1988.

This species was described and illustrated in Gams (1971, anamorph only) and Samuels (1988).

# Dimerosporiella guarapiensis (Speg.) Rossman & Samuels, comb. nov.

- Calonectria guarapiensis Speg., Anales Soc. Ci. Argent. 19: 41. 1885.
  - Nectria microleuca Rossman, Mycotaxon 8: 515. 1979.
- ≡ Nectriopsis guarapiensis (Speg.) Samuels, Mem. New York Bot. Gard. 48: 42. 1988.
- Nectria bakeri Rehm, Ann. Mycol. 6: 319. 1908.[= Nectria perpusilla Sacc., Ann. Mycol. 11: 515. 1913, non (Mont.) Mont. 1856].

This species was described and illustrated in Samuels (1976a, as *Nectria bakeri*, 1988).

Dimerosporiella leucorrhodina (Mont.) Rossman & Samuels, comb. nov. — Plate 4, a.

- ≡ Peziza leucorrhodina Mont., in Sagra, Hist. Phys. Cuba, Bot. Pl. Cell. p. 360. 1842.
- ≡ Calonectria leucorrhodina (Mont.) Speg., Anal. Soc. Ci.
  Argent. 19: 40. 1885.
- ≡ Scutula leucorrhodina (Mont.) Speg., Anal. Soc. Ci. Argent, 26: 58, 1888.
- ≡ Belonidium lecuorrhodinum (Mont.) Sacc., Syll. Fung. 8: 501, 1889.
- ≡ Trichobelonium leucorrhodina (Mont.) Seaver, North Amer. Cup Fungi (Inoperculates), p. 161. 1951.
- Nectriopsis leucorrhodina (Mont.) Samuels, Mem. New York Bot. Gard. 48: 42. 1988.
- = Nectria byssiseda Rehm, in Winter, Rabenhorstii Fungi Europ. Exs., Ed. Nova, Ser. 2, Cent. 22, no. 4152. 1898.
- rop. Exs., Ed. Nova, Ser. 2, Cent. 22, no. 4152, 1898. = Calonectria tubaroënsis Rehm, Hedwigia 37: 195. 1898.
- = Pseudomeliola collapsa Earle, Bull. New York Bot. Gard. 3: 309. 1905.
- = Calonectria limpida Syd. & P. Syd., Leafl. Philipp. Bot. 5: 1545, 1912.
- = Pseudomeliola pipericola F. Stevens, Bot. Gaz. 65: 230. 1918.
- = Nectria puberula Speg. var. microspora Bat. & Nascim., in Batista et al., Inst. Micol. Recife Publ. 33: 5. 1956.
- = Calonectria ukolayii Thaung, Trans. Brit. Mycol. Soc. 67: 435. 1976.

This species was described and illustrated in Samuels (1976a, 1988).

Specimen Illustrated.— UGANDA. Entebbe Rd., on *Meliola* on living leaves of *Trichilia buchamani*, Aug 1944, C.G. Hansford, as *Calonectria cephalosporii* (BPI 631957).

# Dimerosporiella oidioides (Speg.) Rossman & Samuels, comb. nov.

- ≡ Nectria oidioides Speg., Bol. Acad. Nac. Ci. 11: 524.
  1889
- ≡ Nectriopsis oidioides (Speg.) Samuels, Mem. New York
  Bot. Gard. 48: 42. 1988.

This species was described and illustrated in Samuels (1976a, 1988).

# Dimerosporiella pipericola (Henn.) Rossman & Samuels, comb. nov.

- Nectria pipericola Henn., Hedwigia 43: 244. 1904
- ≡ Nectriopsis pipericola (Henn.) Samuels, Mem. New York Bot. Gard. 48: 42. 1988.
- = Epinectria meliolae Syd. & P. Syd., Ann. Mycol. 15: 215. 1917.

This species was described and illustrated in Samuels (1976a, 1988) and one synonym is added here.

Type.— PHILIPPINES: Luzon, prov. Sorsogon, parasitic on mycelium of *Meliola* on leaves of a grass, July-Aug 1915, M. Ramos (Bureau of Science 23722). (Specimen with a typed label at FH is herein designated the **lectotype** of *Epinectria meliolae*; isolectotype at FH; two isolectotypes at BPI).

## KEY TO THE SPECIES OF DIMEROSPORIELLA

	Ascospores generally less than 12 µm long, smooth or finely spinulose
1.	Ascospores generally more than 12 μm long, smooth or striate
2.	Ascospores $8-10.5 \times 2.5-3~\mu m$ , smooth to finely spinulose; ascomata glabrous
2.	Ascospores $9-11 \times 3-4~\mu m$ , smooth; with modified, hair-like cells arising from around the ascomatal apex
3.	Ascospores (14-)17-22(-27) × 3-4 μm, striate
3.	Ascospores smaller, smooth-walled, spinulose or striate
	Ascospores striate, $(11-)12.5-16.5\times 3-4~\mu m$ ; ascomatal hairs lacking; anamorph not present
4.	Ascospores smooth; ascomatal hairs present or lacking; ascomata often associated with an Acremonium-like anamorph having thick-walled conidiophores
5.	Ascospores $(8.5-)11.5-15.5(-18) \times 2.5-4$ µm; ascal apex simple; with hyphal hairs arising from around the ascomatal apex
5.	Ascal apex with a small ring; ascomatal lacking hairs
6.	Ascospores (13.5–)14–16(–17) × 2–3.5(–4) μm
6.	Ascospores 12–14 × 4–4.5 μm

Dimerosporiella sensitiva (Rehm) Rossman & Samuels, comb, nov.

- Nectria sensitiva Rehm. Hedwigia 39: 222. 1900.
- Nectriopsis sensitiva (Rehm) Samuels, Mem. New York
  Bot. Gard. 48: 40. 1988.

This species was described and illustrated in Samuels (1988).

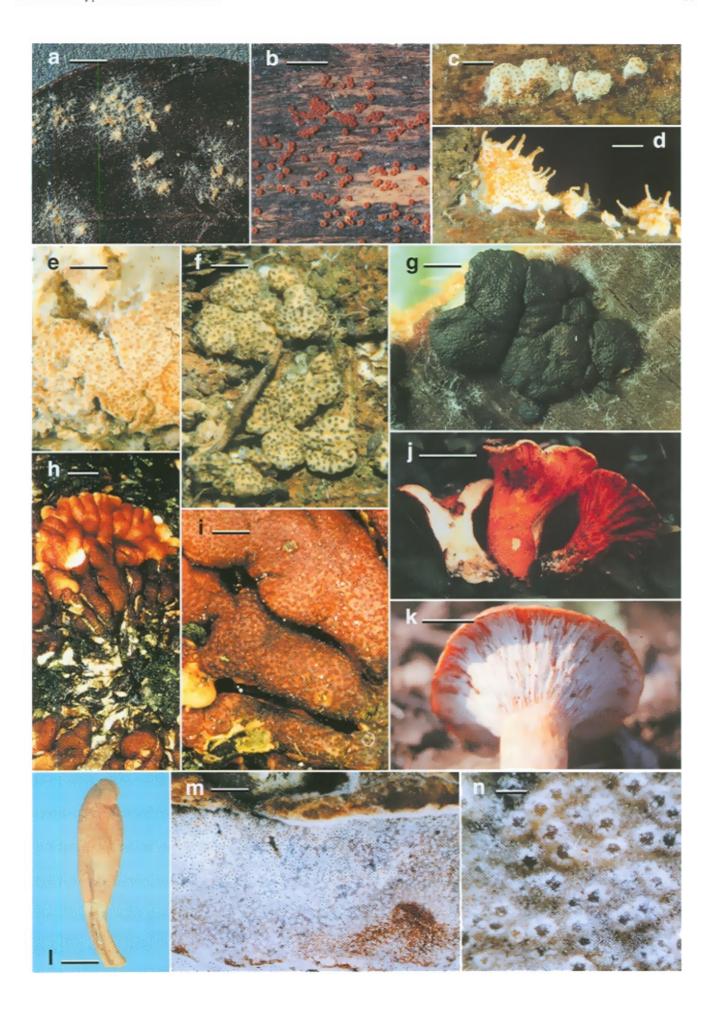
EMERICELLOPSIS J.F.H. Beyma, Antonie van Leeuwenhock Ned. Tijdschr. Hyg. 6: 263, 1940.

Type: E. terricola J.F.H. Beyma.

Ascomata globose, hyaline but appearing brown due to the ascospores, wall of hyaline, flattened cells, non-ostiolate. Asci globose, hyaline, 8-spored. Ascospores ellipsoid, pale brown, 'initially smooth, wide, gelatinous layer collapsing to form 3-6 longitudinal wings at maturity' (Domsch et al., 1980, vol. 1, p. 272). Anamorph Acremonium. Isolated from soil and numerous organic substrata.

Notes.— Emericellopsis was described as a member of the Eurotiaceae and assumed to bear a relationship with the teleomorph of Emericella nidulans based on the distinctively ornamented ascospores. Recent molecular studies have confirmed accounts that place this genus in the Hypocreales. Based on both 18S and 28S sequence data, Emericellopsis grouped within the Hypocreales (Glenn et al., 1996: Ogawa et al., 1997), allied with Mycoarachis in a subclade of the Bionectriaceae along with several anamorph genera. A nomenclatural account of the genus Emericellopsis with descriptions of six accepted species is provided by Gams (1971); since then three more species have been published.

Plate 4. a. Dimerosporiella leucorrhodina. b. Ochronectria calami. c. Protocreopsis fusigera. d. Stilbocrea macrostoma. c. Arachnocrea stipata. f. Hypocrea aureoviridis f. macrospora. g. Hypocrea pseudokoningii. h-i. Hypocreopsis lichenoides. j. Hypomyces lactifluorum. k. Hypomyces lateritius. l. Podostroma alutaceum. m-n. Protocrea farinosa. a. BPI 631957. b. Holotype of Calonectria oödes – K. c. EC 682. d. BPI 744508; e. Fuckel 2358, BPI. f. G.J.S. 96-189, BPI 744424, g. PDD 23871. h, i. Photograph by J.-F. Magni, A8907. j. BPI slide 2030, photograph by K.H. McKnight. k. Photograph by S. Stein. Wayne, Maine, 1963. l. Holotype of Podostroma leucopus – H. m. n. Photograph by J.-F. Magni, specimen A94131. Scale bars: a = 2 mm. b = 2.7 mm. c = 1.8 mm, d = 1.6 mm, e = 3 mm. f = 5 mm, g = 2.5 mm. h = 4 mm, i = 1 mm, j = 50 mm. k = 25 mm, l = 4 mm, m = 2.5 mm, n = 500 μm.



Emericellopsis terricola J.F.H. Beyma, Antonie van Leeuwenhoek Ned. Tijdschr. Hyg. 6: 263. 1940. Anamorph: Acremonium.

Ascomata 30–125(–300) μm diam, non-ostiolate, wall 6–15 μm thick. Asci 14–16 μm long. Ascospores ellipsoid, pale brown, 4.5–6.5 × 2.5–4 μm, surrounded by 4–6 longitudinal, subhyaline wings, finely spinulose. Anamorph Acremonium, with phialides 30–45 μm long, tapering from 1.5–2.5 μm at the base to 1–1.5 μm at the apex. Conidia narrowly ellipsoid, 5.5–8.5 × 2–2.5 μm, about the same length as but narrower than the ascospores, hyaline. Description modified from Domsch et al. (1980).

Habitat.— Isolated from forest- and cultivated soils, fresh and estuarine water, sputum, slime fluxes, bean and potato rhizosphere, mycorrhizae, bee provisions, and air.

DISTRIBUTION, - Worldwide.

EX-TYPE CULTURE.— NETHERLANDS. Isolated from soil, F.H. van Beyma, CBS 120.40, not examined. ILLUSTRATIONS.— Domsch *et al.* (1980, Fig. 113); Gams (1971, Fig. 9 d, e).

HALONECTRIA E.B.G. Jones, Trans. Brit. Mycol. Soc. 48: 287. 1965.

Type: H. milfordensis E.B.G. Jones.

Ascomata partly or totally immersed in the substratum, solitary or gregarious, ascomata orange, globose, each with an elongate neck emerging from the substratum, fleshy. Asci clavate, deliquescing at maturity, 8spored. Ascospores fusiform, non-septate, thin-walled, hyaline, smooth. Anamorph unknown. On intertidal wood.

Notes.— Jones (1965) described this unispecific genus as being similar to Nectria but differentiated by the immersed perithecia with long necks. Kohlmeyer & Kohlmeyer (1968, 1979) provided a description and illustrations of H. milfordensis noting its occurrence on intertidal wood from northern regions of both the Atlantic and Pacific Oceans. They considered the genus to be a member of the Hypocreaceae similar to Trailia stating, however, that 'Halonectria has many characters in common with members of the family Halosphaeriaceae von Arx & E. Müll.', from which it was excluded due to the lack of appendaged ascospores. In a recent classification of filamentous marine ascomycetes, Kohlmeyer (1986) retained Halonectria as one of the four marine hypocrealean genera. The immersed ascomata with long necks and the elongate, aseptate ascospores of H. milfordensis are unlike most hypocrealean fungi. However, the wall surface anatomy and negative reaction in KOH indicate that it could be hypocrealean and, at present, is best included in the Bionectriaceae.

Halonectria milfordensis E.B.G. Jones, Trans. Brit. Mycol. Soc. 48: 287, 1965.

Ascomata immersed, solitary, scattered, orange, becoming dark orange with age, KOH–, yellow in lactic acid, globose to subglobose, 130–250  $\mu m$  tall  $\times$  105–180  $\mu m$  diam, each with an elongate, orange neck 108–252  $\mu m \times 30$ –54  $\mu m$ ; in immature ascomata, necks filled with elongate, hyaline, thin-walled cells. Cells of wall surface forming a textura angularis, thin-walled. Asci clavate, deliquescing at maturity, 21.5–28.5  $\times$  4–6.5  $\mu m$ , 8-spored. Ascospores fusiform, slightly curved, 16.5–29  $\times$  2–3.5  $\mu m$ , non-septate, hyaline, smooth-walled. Associated anamorph with pycnidia partly immersed or superficial, solitary or gregarious, reddish brown, obpyriform or cylindrical, coriaceous, 140–170  $\times$  45–55  $\mu m$ ; conidia filiform, non-septate, hyaline.

Habitat.— On intertidal wood.

DISTRIBUTION.— Temperate regions of the Atlantic and Pacific Oceans.

HOLOTYPE.— GREAT BRITAIN. South Wales: Pembrokeshire, Dale Fort Field Centre, on blocks of Scots pine, 19 Apr 1961, E.B.G. Jones (IMI 86722). This specimen consists of thin slices of wood without bark. A few ascomata lying near the surface were examined but the specimen is in poor condition.

Illustrations.— Jones (1965, Fig. 1), Kohlmeyer & Kohlmeyer (1968, Figs. 1–8).

Notes.— This description is based primarily on the original publication and Kohlmeyer & Kohlmeyer (1979).

HELEOCOCCUM C.A. Jørg., Bot. Tidsskr. 37: 417.

Type: H. aurantiacum C.A. Jørg.

Ascomata superficial, white, pale pink, pale orange, pale brown to greyish or bright yellow, KOH-, globose, surface of loosely interwoven hyphae, wall pseudoparenchymatous, non-ostiolate, disintegrating at maturity. Asci subglobose, globose to broadly clavate or cylindrical, irregularly arranged. Ascospores ovoid to ellipsoid, I-septate, slightly constricted or not, hyaline to pale yellow, smooth, slightly roughened, irregularly striate, or having irregular wing-like ridges, with or without an irregular gelatinous sheath. Anamorph, where known, Acremonium- or Trichothecium-like. Isolated from soil or water submerged in seawater.

Notes.- Heleococcum is a cleistothecial fungus that has been placed in the Eurotiales as well as the Hypocreales. Although characteristics of Heleococcum such as the cleistothecial ascomata, lack of a nectrioid centrum, and deliquescent asci are not typical, the fleshy, bright-colored ascomata, one-septate, hyaline ascospores, and Acremonium- or Trichothecium-like anamorphs suggest that Heleococcum does belong in the Hypocreales. Tubaki (1967) presented a thorough review of the genus, speculating on the relationship of Heleococcum with the Eurotiales or Hypocreales and suggested that the genus represents a link between these two orders. Using 28S rDNA sequence data, Rehner & Samuels (1995) demonstrated that Heleococcum japonense is hypocrealean and grouped with Roumegueriella rufula, another cleistothecial member of the Bionectriaceae.

Heleococcum aurantiacum C.A. Jørg., Bot. Tidsskr. 37: 417. 1922. — Plate 5, a-b.

Ascomata solitary, scattered, superficial, pale orange, KOH-, globose, 210-275  $\mu m$  diam, non-ostiolate, ascomatal wall pseudoparenchymatous, breaking down to release the asci. Asci subglobose to globose, 37.5-45  $\times$  30  $\mu m$ , irregularly arranged, 8-spored. Ascospores ellipsoid, 23.5-27  $\times$  9-10.5  $\mu m$ , 1-septate, hyaline to pale yellow, walls 1.5  $\mu m$  thick, smooth with irregular gelatinous sheath on outer wall, loosening in KOH.

HABITAT.— On moist soil associated with algae or mushroom compost.

DISTRIBUTION. - England and Denmark.

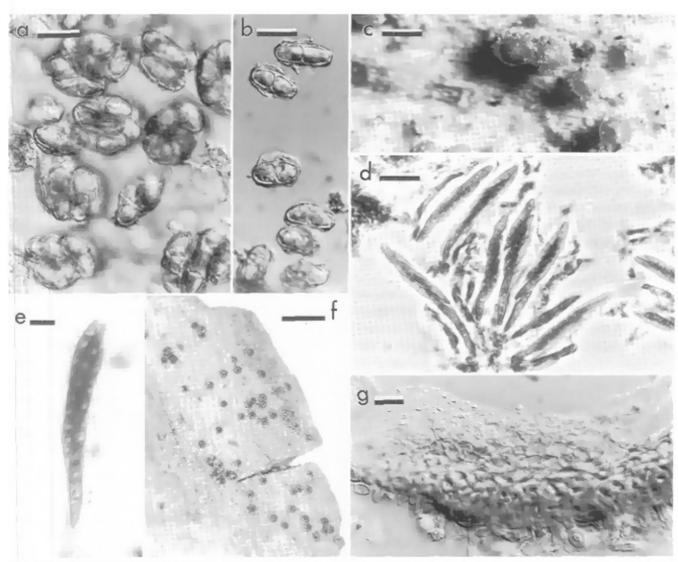


Plate 5. a-b. Heleococcum aurantiacum. a. Asci with ascospores, b. Ascospores, c-e. Ijuhya chilensis. c. Ascomata. d-e. Asci with striate ascospores, f, g, Lasionectria mantuana. f. Ascomata on natural substratum. g. Median section of ascomatal wall. a, b. Holotype – C. c-e. Holotype of Lepidonectria chilensis – LPS. f, g. Holotype – PAD. Scale bars: a, b = 20 μm; c = 250 μm; d = 25 μm; e = 10 μm; f = 1 mm; g = 100 μm.

## KEY TO THE SPECIES OF HELEOCOCCUM modified from Udagawa et al. (1995)

- Ascospores less than 22 µm long, hyaline; anamorph Acremonium-like or Trichothecium-like; isolated from soil or wood immersed in sea water; known from Indonesia, Japan, or the Philippines
- Ascospores 18–21 × 10–13 μm, smooth or slightly roughened; anamorph Trichotheciumlike; isolated from wood immersed in sea water; known from Japan H. japonense Tubaki
- Ascospores less than 18 μm long; anamorph Acremonium-like; isolated from soil; known from Indonesia or the Philippines

HOLOTYPE.— DENMARK. Botanical Garden of the University of Copenhagen, in the moor, on moist soil, Autumn 1921, L. Kolderup Rosenvinge (C; NY, slides of holotype). Culture CBS 201.35

ILLUSTRATIONS.— Dennis (1978, Pl. 44J); Jørgensen (1922, Figs. 1-2); Tubaki (1967, Pl. 2E-F).

Notes.— The holotype specimen and slides of *Heleo-coccum aurantiacum* were examined, on which the few remaining ascomata were broken, apparently disintegrating, revealing globose asci and loose ascospores in the centrum. The above description is based on this fragmentary type specimen, the original description, and the description and illustrations based on the type specimen by Tubaki (1967).

A second species. Heleococcum japonense, was described by Tubaki (1967) that occurs on submerged wood in a marine habitat and produces a Trichothecium-like anamorph (culture CBS 397.67). Two additional species of Heleococcum both having Acremoniumlike anamorphs have been described recently by Udagawa et al. (1995) who included a key to the four species of Heleococcum.

## HYDROPISPHAERA Dumort.. Comment. bot. p. 90. 1822.

Type: *H. peziza* (Tode : Fr.) Dumort. (≡ *Sphaeria peziza* Tode : Fr.).

≡ Nectria subgenus Hyphonectria Sacc., Sylf. Fung. 2:
501. 1883.

≡ Hyphonectria (Sacc.) Petch, J. Bot. 75: 220. 1937. — Lectotype, designated by Samuels (1976a): Nectria peziza (Tode: Fr.) Fr. (≡ Sphaeria peziza Tode: Fr.), recognized as Hydropisphaera peziza (Tode: Fr.) Dumort,

≡ Neuronectria Munk, Dansk Bot. Ark. 17 (1): 56. 1957.

— Type: N. peziza (Tode: Fr.) Munk (≡ Sphaeria peziza Tode: Fr.), recognized as H. peziza.

= Neohenningsia Koord., Verh. Kon. Ned. Akad. Wetensch. Afd. Natuurk., Sect. 2, 13: 164, 1907. — Type: N. stellatula Koord. (≡ Nectria stellatula (Koord.) Höhn., a synonym of H. rufofusca.

= Perrotiella Naumov, Bull. Soc. Oural. Amis Sci. Nat. p. 26. 1916. — Type: P. uralensis Naumov, a synonym of H. peziza.

Ascomata superficial, non-stromatic, pale yellow, orange or umber, KOH –, globose to subglobose or doliiform, usually collapsed and deeply cupulate, smooth or
with fasciculate hairs. Ascomatal wall generally over 25

µm thick, of two regions; outer region of thin-walled,
globose cells. Asci clavate. Ascospores ellipsoid, 1- to
multiseptate, hyaline, generally finely to coarsely striate, rarely smooth or spinulose. Anamorph, where
known, Acremonium-like. On dead herbaceous or
woody monocotyledonous or dicotyledonous substrata.

Notes.— Within the Nectria-like fungi of the Hypocreales, Hydropisphaera is unique in ascomatal wall structure in which the wall is relatively thick, generally over 25 μm, and up to 100 μm thick in H. pachyderma, and is composed of large, thin-walled, globose cells often over 15 µm diam. This ascomatal wall structure results in a characteristic deeply cupulate collapse of the ascomata upon drying (Booth, 1959; Rossman, 1983; Samuels, 1976b). The ascospores of Hydropisphaera are one- to multiseptate, often finely to coarsely striate although also spinulose or smooth. The anamorphs have been placed in Acremonium or, if the conidia are septate, in Cephalosporiopsis, always having simple conidiophores and relatively long, tapering, phialidic conidiogenous cells. Species of Hydropisphaera often occur as saprobes on decaying monocotyledonous plants and ferns, although there are many exceptions including *H. boothii* (on *Oenanthe (Apiaceae*), England), *H. erubescens* (various dicotyledonous herbaceous substrata in temperate regions), *H. gigantea* (on *Conium* and other herbaceous stems. Argentina and Ecuador), *H. haematites* (on unidentified woody plant, Africa), *H. peziza* (on decaying wood, polypores, bark, soil, and other organic substrata, cosmopolitan) and *H. pachyderma* (on bark of unidentified twig, Colombia). In his molecular study of *Fusarium* and related fungi, O'Donnell (1993) included *H. peziza* (as *N. peziza*) and found that this species grouped most closely with other pallid *Nectria*-like species now placed in the *Bionectriaceae*.

At the time the generic name *Hydropisphaera* was proposed, almost all pyrenomycetes were included in the one family *Sphaeriaceae* and the name had been ignored until recently. In proposing *Nectria* for conservation against *Hydropisphaera* and *Ephedrosphaera* Dumort., Cannon & Hawksworth (1983) unearthed *Hydropisphaera* as the oldest name available for the *Nectria peziza*-group when recognized at the generic level.

Saccardo (1883) established Nectria subgenus Hyphonectria Sacc. in which he included nine species. When Petch (1937) raised the subgenus to generic rank, he made reference to Saccardo's subgeneric taxon and included four additional species (Hyphonectria violacea, H. berkeleyana, H. aureonitens, and H. solani). Neither Saccardo (1883) nor Petch (1937) designated a type species. Samuels (1976a) discussed the problem of typification of Nectria subgenus Hyphonectria and the genus Hyphonectria and lectotypified this taxon with Nectria peziza as one of the original nine species included in Nectria subgenus Hyphonectria by Saccardo (1883). Samuels (1976a) considered the reference by Petch to the nine species originally included in Nectria subgenus Hyphonectria to constitute inclusion in Hyphonectria at the generic rank even though Petch did not specifically transfer any of the species to the genus.

Neohenningsia was initially placed in the Aspergilleae, Eurotiaceae, despite the presence of an ostiole. Although Koorders (1907) suggested a relationship with Charonectria Sacc. and Baculospora Zukal in the Hypocreales, he differentiated his genus from these in stating that Neohenningsia had superficial ascomata with fasciculate appendages, thin-walled asci, and septate ascospores. Von Höhnel (1909a) placed the type species of Neohenningsia in Nectria. Samuels (1976b) reviewed the history of Neohenningsia and unsuccessfully sought the type specimen of N. stellulata at BO and FH; it does also not exist at B. Based on the original description, Neohenningsia stellulata is herein neotypified and this species is regarded as a synonym of H. rufofusca.

Perrotiella was described as a discomycete in the

Pezizaceae. Nannfeldt (1932) examined authentic material of the type species and confirmed that *P. uralensis* is a synonym of *Nectria peziza*, thus *Perrotiella* is a synonym of *Hydropisphaera*.

Neuronectria was described for species of Nectria having striate ascospores. This character is not unique to any particular group of nectrioid fungi as evidenced by Samuels (1988). He included thirty species of Nectria having pallid ascomata and hyaline, striate ascospores in seven different groups of Nectria sensu lato, and Rossman (1989) documented three of the 28 species in the Nectria sensu stricto having striate ascospores. Many genera of nectrioid fungi include species having striate ascospores.

Hydropisphaera peziza (Tode: Fr.) Dumort., Comment. bot. p. 90. 1822.

- ≡ Sphaeria peziza Tode: Fr., Tode, Fungi Mecklenb, Sel.,
  Fasc, 2: 46, 1791: Fries, Syst. Mycol. 2: 452, 1823.
- ≡ Nectria peziza (Tode: Fr.) Fr., Summa Veg. Scand. p.
  388. 1849.
- ≡ Dialonectria peziza (Tode : Fr.) Cooke, Grevillea 12:
  110, 1884.
- ≡ Cucurbitaria peziza (Tode : Fr.) O. Kuntze, Rev. Gen.
  Pl. 3: 461, 1898.
- Neuronectria peziza (Tode: Fr.) Munk, Dansk Bot. Ark.
  17 (1): 58. 1957.
- = Nectria fimicola Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 179. 1869 [1870].
- ≡ Byssonectria fimicola (Fuckel) Cooke, Grevillea 12: 109, 1884.
- ≡ Cucurbitaria fimicola (Tode: Fr.) O. Kuntze, Rev. Gen.
  Pl. 3: 461, 1898.
- = Nectria pezicula Speg., Michelia 1: 232. 1878.
- ≡ Byssonectria pezicula (Speg.) Cooke, Grevillea 12: 109.
  1884
- = Nectria epigaea Cooke, Grevillea 8: 10, 1879.
- ≡ Byssonectria epigaea (Cooke) Cooke, Grevillea 12:
  109, 1994.
- = Nectria rimicola Cooke, Grevillea 11: 108. 1883.
- ≡ Cucurbitaria rimicola (Cooke) O. Kuntze, Rev. Gen. Pl.
  3: 461, 1898.
- = Nectria umbellulariae Plowr. & Harkn., Bull. Calif. Acad. Sci. 1: 26, 1884.
- ≡ Cucubitaria umbellulariae (Plowr. & Harkn.) O. Kuntze, Rev. Gen. Pl. 3: 462. 1898.
- = Nectria perforata Ellis & Holw., in Arthur, Geol. Nat. His. Surv. Minnesota Bull. 3: 33. 1887.
- ≡ Cucurbitaria perforata (Ellis & Holw.) O. Kuntze, Rev. Gen. Pl. 3: 461, 1898.
- = Nectria consanguinea Rehm, Hedwigia 26: 92. 1887.
- = Nectria importata Rehm, Hedwigia 27: 171, 1888.
- ≡ Cucurbitaria importata (Rehm) O. Kuntze, Rev. Gen. Pl. 3: 461, 1898.
- = Nectria henningsii Rehm, Hedwigia 28: 352, 1889.
- = Nectria betulina Rehm, Ann. Mycol. 3: 519, 1905 [1906].
- = Nectria sphagnicola Kirschst., Verh. Ver. Prov. Brandenburg 48: 59. 1906.
- = Nectria fallax Rick, Ann. Mycol. 4: 309. 1906.
- = Perrotiella uralensis Naumov, Bull. Soc. Oural. Amis Sci. Nat. p. 26. 1916.

Anamorph: Acremonium sp.

Mycelium not visible or white, surrounding the ascomatal base, subtending hyphae unbranched, thinwalled, ca 4 µm wide. Ascomata superficial or basally immersed, solitary to gregarious, subglobose, globose to urniform or almost discoidal, becoming cupulate when dry, 370-420  $\mu$ m high × (250-)370-430(-550) across the flat tops, yellow to orange, smooth or slightly furfuraceous, papilla lacking or short and acute, of cylindrical, septate, unbranched hyphae with rounded tips, ca 3 µm wide; periphyses 20-30 µm long. Ascomatal wall 30-50(-70) µm thick, of two regions: outer region 15-30(-50) µm thick, cells broadly ellipsoid to globose, 10-15 µm diam, thin-walled; inner region ca 15 µm thick, cells flattened and compacted; cells in surface view spherical, 15-25 µm diam, thin-walled. If present, hairs short, orange, consisting of septate, unbranched, thin-walled hyphae, occasionally forming triangular fascicles. Asci clavate, (49-)60-75(-100) × (5-)8-10(-14) μm, apices flat, 8-spored, ascospores biseriate above, uniseriate below. Ascospores broadly ellipsoid,  $(9-)11-14(-17) \times (3-)5-7 \mu m$ , equally 2celled, not constricted or slightly constricted at the septum, hyaline, conspicuously longitudinally striate.

Habitat.— On well-rotted wood, bark, dung, decaying cloth, and basidiocarps of polypores.

Distribution.— Cosmospolitan, especially common in temperate regions.

LECTOTYPE, designated here.— SWEDEN. On rotten wood, Sclerom. Suec. 24 no. 235. 1822 (BPI, in Sbarbaro collections in bound centuries 1–3). The specimens of Tode were destroyed, thus none of the specimens of *S. peziza* examined by Tode are still in existence, Fries (1823) mentioned several specimens following the description of *S. peziza*, including Sclerom. Suec. 24 no. 235. Booth (1959) stated that the specimen of this number at K was in poor condition. The specimen at BPI is in excellent condition.

ILLUSTRATIONS.— Booth (1959, Fig. 32, as Nectria peziza); Dennis (1978, Pl. 32C, as N. peziza); Dingley (1951b, Fig. 1, as N. peziza); Ellis & Ellis (1985, Fig. 135); Hanlin (1963a, Figs. 1–47, as Neuronectria peziza); Malençon (1979, Fig. 1A, as Nectria peziza); Samuels, (1976b, Figs. 16A, 17A–E, as N. peziza).

Notes.— Hanlin (1963a) studied the centrum development of *H. peziza* as *Neuronectria peziza*. Gams (1971) and Samuels (1976b) described the anamorph in culture.

SEVENTEEN ADDITIONAL SPECIES are included in *Hydropisphaera* as follows:

**Hydropisphaera arenula** (Berk. & Broome) Rossman & Samuels, *comb*, *nov*.

- ≡ Sphaeria arenula Berk. & Broome, Ann. Mag. Nat. Hist., Ser. 2, 9: 320, 1852.
- Nectria arenula (Berk. & Broome) Berk.. Out. Brit.
  Fung. p. 394. 1860.
- Dialonectria arenula (Berk. & Broome) Cooke, Grevillea 12: 110, 1884.
- ≡ Cucurbitaria arenula (Berk. & Broome) O. Kuntze, Rev. Gen. Pl. 3: 460. 1898.
- = Calonectria transiens Rehm, Hedwigia 39: 225. 1900.

This species was described and illustrated in Booth (1959), Samuels (1978), and Schmid & Schmid (1991), each as Nectria arenula.

Hydropisphaera arenuloides (Samuels) Rossman & Samuels, comb. nov.

≡ Nectria arenuloides Samuels, New Zealand J. Bot. 14:
254. 1976.

This species was described and illustrated in Samuels (1976b).

Hydropisphaera boothii (D. Hawksw.) Rossman & Samuels, comb. nov.

≡ Nectria boothii D. Hawksw., Trans. Brit. Mycol. Soc. 74: 572, 1980.

This species was described and illustrated in Hawksworth & Minter (1980) as N. boothii.

Hydropisphaera cyatheae (Dingley) Rossman & Samuels, comb. nov.

≡ Nectria cyatheae Dingley, Trans. Roy. Soc. New Zealand 83: 652. 1956 (as 'cyathea').

This species was described and illustrated in Dingley (1956) and Samuels (1976b) as *N. cyatheae*. Culture CBS 575.76.

Hydropisphaera dolichospora (Penz. & Sacc.) Rossman & Samuels, comb. nov.

≡ Nectria dolichospora Penz. & Sacc., Malpighia 11: 513.
1897.

This species was described and illustrated in Samuels (1976a) and Samuels et al. (1990) as N. dolichospora.

Hydropisphaera erubescens (Desm.) Rossman & Samuels, comb. nov.

- ≡ Sphaeria erubescens Desm., Ann. Sci. Nat., Bot., Sér. 3,
  6: 72. 1846.
- ≡ Calonectria erubescens (Desm.) Sacc., Michelia 1: 309.
  1878.

≡ Amphinectria erubescens (Desm.) Sacc. ex Speg., Bol.
Acad. Ci. (Cordoba) 26: 347. 1927.

= Calonectria umbelliferarum Seaver, Mem. New York Bot. Gard. 6: 507. 1916.

= Calonectria venezuelensis Syd. & P. Syd., Ann. Mycol. 33: 88, 1935.

= Calonectria crescentiae Seaver & Waterston, Mycologia 32: 404, 1940.

This species was described and illustrated in Rossman (1983) and Samuels (1978), as *N. erubescens*. Cultures CBS 333.76–335.76.

Hydropisphaera gigantea (Speg.) Rossman & Samuels, comb. nov.

≡ Lasionectria gigantea Speg., Anal. Mus. Nac. Buenos Aires 3, 1: 77, 1902.

≡ Nectria gigantea (Speg.) Sacc. & D. Sacc., Syll. Fung.
 17: 792. 1905.

This species was described in Samuels (1976a) as N. gigantea, based on the type specimen from Argentina.

A specimen additional to the type has been collected, examined, and is reported here: ECUADOR, Prov. Zamora: ca 21 km from Zamora, on the Zamora-Yanzoza Road, elev. ca 1,000 m, on herbaceous stem, 31 July 1975, K.P. Dumont (Dumont–EC 1779), S.E. Carpenter & P. Buritica (NY).

Hydropisphaera haematites (Syd. & P. Syd.) Rossman & Samuels, comb. nov.

Nectria haematites Syd. & P. Syd., in Mildbraed. Wiss. Ergebn. Deutsch. Zent. Afrika Exped. 2, Bot. prior to 99, 1914.

This species was described and illustrated in Samuels (1976a) as N. haematites.

Hydropisphaera hypoxantha (Penz. & Sacc.) Rossman & Samuels, comb. nov.

≡ Nectria hypoxantha Penz. & Sacc., Malpighia 11: 513.
1897.

This species was described and illustrated in Samuels et al. (1990) as N. hypoxantha.

Hydropisphaera leucotricha (Penz. & Sacc.) Rossman & Samuels, comb. nov.

■ Nectria leucotricha Penz. & Sacc., Malpighia 11: 512. 1897.

This species was described and illustrated in Samuels et al. (1990) as N. leucotricha.

Hydropisphaera macrarenula (Samuels) Rossman & Samuels, comb. nov.

This species was described and illustrated in Samuels et al. (1990) as N. macrarenula.

Hydropisphaera multiloculata (Samuels) Rossman & Samuels, comb. nov.

≡ Nectria multiloculata Samuels, New Zealand J. Bot. 16:
78, 1978.

This species was described and illustrated in Rossman (1983) and Samuels (1978) as *N. multiloculata*. Cultures CBS 339.77–341.77.

Hydropisphaera multiseptata (Samuels) Rossman & Samuels, comb. nov.

■ Nectria multiseptata Samuels, New Zealand J. Bot. 16:
 77. 1978.

This species was described and illustrated in Rossman (1983) and Samuels (1978) as *N. multiseptata*. Cultures CBS 336.77–338.77.

**Hydropisphaera nymaniana** (Henn.) Rossman & Samuels, *comb. nov.* 

■ Nectria nymaniana Henn., Monsunia 1: 161. 1899.

This species was described and illustrated in Samuels (1976a) and Samuels et al. (1990) as N. nymaniana.

Hydropisphaera pachyderma (Rossman) Rossman & Samuels, comb. nov.

≡ Nectria pachyderma Rossman, Mycol. Pap. 150: 75.
1983.

This species was described and illustrated in Rossman (1983) as N. pachyderma.

Hydropisphaera rufofusca (Penz. & Sacc.) Rossman et al., Mycologia 85: 702. 1993. — Plate 2, b.

≡ Nectriella rufofusca Penz. & Sacc., Malpighia 11: 507.
 1897. — Holotype: INDONESIA. Java: Tjibodas, in caulibus emortuis Elettariae, 6 Feb 1897, No. 436 p.p. (PAD).
 = Neohenningsia stellatula Koord., Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Tweede Reeks, Sect. 2, 13: 164.
 1907. Neotype here selected: BRAZIL. Parà: in foliis Monsterae sp., Dec. 1907, Baker, (FH, also holotype of N. brasiliensis).

≡ Nectria stellatula (Koord.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 118: 819. 1909. 

= Neohenningsia brasiliensis Henn., Hedwigia 48: 102. 1909 (1908)

≡ Nectria brasiliensis (Henn.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 118: 1186, 1909.

≡ Pseudonectria brasiliensis (Henn.) Weese, Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturw. Kl. Abt. 1, 125: 518. 1916.

This species was described and illustrated in Samuels et al. (1990) as Nectria brasiliensis.

Hydropisphaera suffulta (Berk. & M.A. Curtis) Rossman & Samuels, comb. nov.

- Nectria suffulta Berk. & M.A. Curtis, J. Linn. Soc., Bot.
  10: 378, 1868.
- = Nectria musae Pat., J. Bot, (Morot) 11: 369. 1897.
- = Nectria pezizelloides, Rehm, Hedwigia 37: 192. 1898.

- = Nectria calamicola Henn. & E. Nyman, in Warburg, Monsunia 1: 161. 1900 [1899].
- = Nectria ornata Massee & E.S. Salmon, Ann. Bot. (London) 16: 75. 1902.
- Nectria setosa Ferd. & Winge, Bot. Tidsskr. 29: 11. 1908.
   Neohenningsia confluens Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 114. 1920.

This species was described and illustrated in Samuels (1976a) and Samuels *et al.* (1990) as *N. suffulta*. Culture CBS 122.87.

## KEY TO THE SPECIES OF HYDROPISPHAERA

1.	Ascospores 1-septate; ascomata without hairs
2 (1). 2.	Ascospores 3-septate, smooth-walled to faintly striate
<b>3</b> (2).	Ascospores $18-26\times4-5~\mu m$ , fusiform; ascomata orange to brown, glabrous to slightly scurfy
<b>4</b> (2). <b>4.</b>	Ascospores 50~70 $\times$ 6–7 $\mu$ m, 15–20-septate, striate; ascomata orange to brown, smooth
5 (1). 5.	Ascomata without hairs, smooth, slightly scaly or warted
6 (5). 6.	As cospores more than 23 $\mu$ m long
7 (6). 7.	Ascomata yellow-orange to orange-red, with a flattened apex; ascospores $23-27 \times 5-6 \mu m$ , striate
8 (6). 8.	Ascospores $1114\times56~\mu\text{m}$ , striate or coarsely striate
9 (8). 9.	Ascospores striate; ascomata yellow to orange, often with yellow subtending hyphae, entirely superficial
10 (8). 10,	Ascospores $14-16\times3.5-4~\mu m$ , ellipsoid-fusiform to fusiform, smooth or striate; ascomata orange, becoming brown

11 (5).	Ascospores averaging more than 25 µm long
11.	Ascospores averaging less than 25 µm long
<b>12</b> (11).	Ascomata dark red with red hairs; ascospores 27-30 × 7-8 μm, spinulose to spinulose stripts
12.	Ascomata dark orange to brown with concolorous hairs; ascospores smooth-walled 13
13 (12). 13.	Ascospores $48-55\times6-7~\mu m$ ; ascomata dark orange with orange hairs $$ $$ $$ $$ $$ $$ $$ $$ $$ $$
<b>14</b> (11).	Ascomata with white or orange, fasciculate hairs; ascospores averaging more than 17 µm long
14.	Ascomata with white, fasciculate hairs; ascospores averaging less than 17 µm long
<b>15</b> (14).	Ascomata yellow to orange or nearly brown with white hairs; ascospores $16-22 \times 4-5$ $\mu m$ , striate or spinulose
15.	Ascomata orange with orange hairs; ascospores $17-23 \times 5-7 \mu m$ , striate H. cyatheae
16 (14). 16.	Ascospores striate, $12-17\times4-5~\mu m$ ; ascomata pale yellow to yellow H. suffulta Ascospores smooth or spinulose, not striate; ascomata orange to dark orange 17
<b>17</b> (16).	Ascospores 12.5–17.5 × 3.5–4 μm, spinulose; warm temperate and tropical
17.	Ascospores 12–15 × 4–5 μm, smooth; known only from England H. boothii

IJUHYA Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25: 30. 1899.

Type: I. vitrea Starbäck, a synonym of I. peristomialis.

= Peristomialis (W. Phillips) Boud., Hist. Classif. Discom. Europe p. 116, 1907.

■ Mollisia subgenus Peristomialis W. Phillips, Man. Brit.

Discome p. 201, 1887.

Discom. p. 201. 1887.

≡ Cyathicula subgenus Peristomialis (W. Phillips) Sacc.,
Syll. Fung. 8: 306. 1889. — Type: P. berkeleyi Boud., a
nomenclatural synonym of I. peristomialis.

= Lepidonectria Speg., Revista Fac. Agron. Univ. Nac. La Plata 6: 97. 1910. — Type: L. chilensis Speg., recognized as I. chilensis.

Ascomata solitary or in small groups, superficial, nonstromatic, white to pale yellow, KOH-, globose to subglobose, usually with a discoidal apex; disk formed of intertwined hyphae that often develop into triangular fasciculate hairs forming an apical crown, rarely apex discoidal without hairs or with short, sinuous hairs. Ascomatal wall usually less than 20 µm thick, of one region of thick-walled, relatively small cells, often forming textura epidermoidea in surface view. Asci clavate, 8spored. Ascospores clavate or fusiform to long fusiform, one- to multiseptate or muriform, hyaline, smooth to striate. Anamorph, where known, Acremonium-like. On decaying herbaceous debris or wood, also on black stroma, hyphae, and ascomata of pyrenomycetous fungi. Notes.— Ijuhya was originally placed in the Gymnoascaceae; later it was considered a member of the Sphaeriaceae (Müller & von Arx, 1973) as well as the Hypocreaceae (Rogerson, 1970). Samuels (1976b) examined the type specimen and determined Ijuhya vitrea to be a synonym of Nectria peristomialis.

Phillips (1887) described Mollisia subgenus Peristomialis for one species having triangular hairs on the ascomata and fusiform, multiseptate ascospores. Mollisia
peristomialis was the only species included in the subgenus, thus, when raised to generic rank, the taxon is automatically typified by that species. When raising the
subgenus to generic rank, Boudier (1907) proposed a
new epithet for the type species in order to avoid a tautonym. He included six species in Peristomialis.
Samuels (1976b) examined the type specimen and regarded the type species as Nectria peristomialis, thus he
considered Peristomialis to be a synonym of Nectria.
Although listed as a synonym of Peristomialis by
Samuels (1976b), Ijuhva has priority over Peristomialis.

Spegazzini (1910) described one species in the genus Lepidonectria. Based on the presence of 'squamules' on the ascomata of L. chilensis, Spegazzini may have intended his species to be in Nectria subgenus Lepidonectria Sacc.; however, no reference is made to that taxon. Spegazzini was the first to use the name Lepidonectria at the generic level which constitutes the valid publica-

tion of a new genus with *L. chilensis* as the implicit type species. None of the species in *Nectria* subgenus *Lepidonectria* were ever placed in the genus *Lepidonectria*. The type specimen of *Lepidonectria* chilensis was examined and the species is herein placed in *Ijuhya*.

Samuels (1988) recognized *Ijuhya* (as *Peristomialis*) as a genus distinct from other pallid members of *Nectria*-like fungi, differentiated primarily by the fasciculate hairs around the ascomatal apex. In addition to the fasciculate hairs, the ascomatal wall anatomy is distinct, composed of thick-walled, relatively small cells. Within *Ijuhya*, ten species are recognized, distinguished primarily on characteristics of the ascospores that vary from one-septate to multiseptate or muriform and are generally coarsely striate, although some species have smooth or spinulose ascospores.

## Ijuhya peristomialis (Berk. & Broome) Rossman & Samuels, comb. nov.

≡ Peziza peristomialis Berk. & Broome, Ann. Mag. Nat. Hist., Ser. 3, 18: 126, 1866.

≡ Mollisia peristomialis (Berk. & Broome) W. Phillips, Man. Brit. Discom. p. 201, 1887.

≡ Cyathicula peristomialis (Berk. & Broome) Sacc., Syll.
Fung. 8: 308, 1889.

≡ Actiniopsis peristomialis (Berk. & Broome) Petch, Trans. Brit. Mycol. Soc. 21: 282. 1938.

≡ Nectria peristomialis (Berk. & Broome) Samuels, New Zealand J. Bot. 14: 251, 1976.

≡ Peristomialis berkeleyi Boud., Hist. Classif. Discom. Europe p. 116. 1907.

= Helotium ciliatum P. Crouan & H. Crouan, Fl. Finistère p.

≡ Cyathicula ciliata (P. Crouan & H. Crouan) Sacc., Syll.
Fung. 8: 306. 1889.

≡ Peristomialis ciliata (P. Crouan & H. Crouan) Boud., Hist. Classif. Discom. Europe p. 116. 1907.

= Ijuhya vitrea Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25: 30. 1899.

= Ijuhya vitrea Starbäck var. javanica Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 121: 380. 1912.

= Nectria tympaniformis Petrak, Sydowia 4: 514. 1950.

Anamorph: Acremonium sp.

Ascomata solitary or in small groups, superficial, without a stroma or on a sparse, hyphal subiculum. Ascomata hyaline to pale yellow, translucent, becoming darker and opaque when dry, KOH-, globose to subglobose with flattened apex, apex often slightly sunken; ascomata not collapsing when dry, 230–375 μm high × 260–350 μm diam, without papilla; each ascoma with an apical disk 150–200 μm diam with a ring of long, fasciculate, hyaline hairs; fascicles of hairs 145–300 × 30–90 μm, tapering to a narrowly rounded tip, agglutinated hyphae hyaline, septate, 2.5–4 μm wide with a 0.5–1 μm thick wall. Ascomatal wall 35–60 μm thick, of three regions: outer region 10–20

μm thick, of loose, prosenchymatous hyphae 3–4 μm wide, with up to 1 μm thick walls; middle region 15–25 μm thick, of elongate cells,  $7-12 \times 5-7$  μm, with hyaline, up to 1 μm thick walls, thicker in the corners, middle region up to 75 μm thick in the upper part, expanding into a flattened apical disk; inner region 10–15 μm thick, of hyaline, thin-walled, elongate cells. Asci  $50-100 \times 12-15$  μm, clavate, simple. 8-spored, ascospores irregularly biseriate to pluriseriate. Ascospores fusiform with rounded ends, often curved,  $(24-)30-60(-110) \times 4-7(-8)$  μm, (1-)5-7(-11)-septate, hyaline, coarsely striate, 3–4 striae per half spore.

Anamorph.— Conidiophores arising from the colony surface, solitary, erect, straight, unbranched, cylindrical, tapering toward the apex, septate at the base, occasionally with one or two additional septa, smooth,  $20-50~\mu\text{m} \times 2-3.5~\mu\text{m}$  at the base. Conidiogenous cells monophialidic, terminal cell of erect conidiophore, cylindrical,  $22-33 \times 1.5-1.8~\mu\text{m}$  at the unflared apex. Conidia ellipsoid to cylindrical with rounded ends,  $5-8 \times 1.7-2.5~\mu\text{m}$ , aseptate, hyaline, smooth, held in a solitary, hyaline drop of liquid.

Habitat.— On monocotyledonous and dicotyledonous wood and herbaceous debris.

DISTRIBUTION.— Brazil, Colombia, England, France, India, Java, New Zealand, Panama and Venezuela (Rossman, 1983; Samuels, 1976b).

Types.— ENGLAND. Penzance, on *Ilex* sp., no. 248 (K. holotype of *P. peristomialis*); BRAZIL. Rio Grande do Sul, on culms of bamboo, 7 Apr 1893, Malme 340 (S, holotype of *Ijuhya vitrea*); INDONESIA. Java, Tjibodas, on bamboo, 1907–8, Höhnel 5792 (FH, holotype of *I. vitrea* var. *javanica*). Culture: CBS 569.76.

ILLUSTRATIONS.— Dennis (1978, Pl. 31K, as A. peristomialis); Rossman (1983, Fig. 40, Pl. 13A-B, as N. peristomialis); Samuels (1976b, Fig. 9, as N. peristomialis).

Notes.— Samuels (1976b) and Rossman (1983) provided descriptions of *I. peristomialis* (as *N. peristomialis*) and its *Acremonium* anamorph.

Ijuhya chilensis (Speg.) Rossman & Samuels, comb. nov. — Plate 2, d, Plate 5, c–e.

≡ Lepidonectria chilensis Speg., Revista Fac. Agron.
Univ. Nac. La Plata 6: 97, 1910.

Ascomata gregarious, superficial, without visible basal mycelium, difficult to remove from the substratum. Ascomata globose, 275 μm diam, non-papillate, dull orange, KOH~, not collapsed, with a fringe of thickwalled, fasciculate hairs around the apex, hairs up to 100 μm long. Ascomatal surface of tightly intertwined hyphae, wall of several regions of intertwined, thick-

walled hyphae. Asci clavate to fusiform,  $65-96 \times 9-11.2 \mu m$ , widest in the middle, apex simple, 6-8-spored, ascospores pluriseriate. Ascospores narrowly fusiform,  $(19-)21-28 \times 3.5-4.5 \mu m$ , 1-septate, slightly or not constricted at septum, hyaline, coarsely striate with striations extending over the length of the spore, few in number.

Habitat and distribution.— Known only from the type specimen.

HOLOTYPE.— CHILE. Valdivia, on decaying shoots of Lobelia lupa, Jan 1909. C. Spegazzini (LPS-1696).

Notes.— Based on Spegazzini's illustrations and the type specimen, *Lepidonectria chilensis* belongs in *Ijuhya* and is similar to *Ijuhya paraparilis* except that ascomata of *I. paraparilis* have a flattened apical disk and occur on black mycelium on bamboo. The ascomata of *I. chilensis* are associated with an effete, black fungus.

EIGHT ADDITIONAL SPECIES are included in *Ijuhya* as follows:

Ijuhya aquifolii (Cooke & Ellis) Rossman & Samuels, comb. nov. — Plate 2, c.

≡ Peziza aquifolii Cooke & Ellis, Grevillea 6: 91. 1878 (as 'aquifoliae'). — Lectotype, designated in Rossman et al. (1993): UNITED STATES. New Jersey: Newfield, on dead leaves of Ilex, associated with dematiaceous hyphae, 21 May 1877, J.B. Ellis 2559 (BPI 1113199).

≡ Pseudonectria aquifolii (Cooke & Ellis) Dennis, Persoonia 3: 35, 1964.

≡ Peristomialis aquifolii (Cooke & Ellis) Rossman et al..

Mycologia 85: 696. 1993 (as 'aquifoliae').

This species was described and illustrated in Rossman et al. (1993) as N. aquifolii.

Ijuhya bambusina (Syd. & P. Syd.) Rossman & Samuels, comb. nov.

≡ Pseudonectria bambusina Syd. & P. Syd., Ann. Mycol.
15: 214, 1917.

≡ Peristomialis bambusina (Syd. & P. Syd.) Rossman et al., Mycologia 85: 699. 1993.

This species was described and illustrated in Rossman et al. (1993) as N. corynespora.

Ijuhya corynespora (Samuels) Rossman & Samuels, comb. nov. ≡ Nectria corynespora Samuels, New Zealand J. Bot. 16:
78. 1978.

≡ Peristomialis corynespora (Samuels) Samuels, Mem. New York Bot, Gard. 48: 18. 1988.

This species was described and illustrated in Samuels (1978) as N. bambusina.

Ijuhya dentifera (Samuels) Rossman & Samuels, comb. nov.

■ Nectria dentifera Samuels, New Zealand J. Bot. 14: 253. 1976.

≡ Peristomialis dentifera (Samuels) Samuels, Mem. New York Bot. Gard. 48: 18. 1988.

This species was described and illustrated in Samuels (1976b) as N. dentifera. Culture CBS 574.76.

Ijuhya dictyospora (Rossman) Rossman & Samuels, comb. nov.

≡ Nectria dictyospora Rossman, Mycol. Pap. 150: 69.
1983.

≡ Peristomialis dictyospora (Rossman) Samuels, Mem. New York Bot, Gard. 48: 20. 1988.

This species was described and illustrated in Rossman (1983) as *N. dictyospora* and Samuels (1988) as *P. dictyospora*.

Ijuhya leucocarpa (Samuels) Rossman & Samuels,

■ Peristomialis leucocarpa Samuels, Mem. New York Bot. Gard. 48: 16. 1988.

This species was described and illustrated in Samuels (1988) as *P. leucocarpa*.

Ijuhya paraparilis (Samuels) Rossman & Samuels, comb. nov.

≡ Peristomialis paraparilis Samuels, Mem. New York Bot. Gard. 48: 16. 1988.

This species was described and illustrated in Samuels (1988) as *P. paraparilis*.

Ijuhya parilis (Syd.) Rossman & Samuels, comb. nov.

= Nectria parilis Syd., Ann. Mycol. 28: 121. 1930.

≡ Peristomialis parilis (Syd.) Samuels, Mem. New York Bot. Gard. 48: 15, 1988.

This species was described and illustrated in Samuels (1988) as P. parilis.

## KEY TO THE SPECIES OF LJUHYA

1. 1.	Ascospores 5- or more-septate or muriform 2 Ascospores 1–3-septate
2 (1). 2.	Ascospores muriform, with 7–13 transverse septa and one, irregular, longitudinal septum, $48–97\times10–16~\mu m$ , smooth
3 (2). 3.	Ascospores 5–7-septate, rarely 1–3- or up to 11-septate, $(24-)30-60(-110)\times 4-7(-8)$ $\mu$ m, fusiform
4 (1). 4.	Ascospores fusiform, more than 10 $\mu m$ long
5 (4). 5.	Ascospores $14.5-20 \times (2.5-)3-5(-5.5)$ µm, spinulose
6 (5). 6.	Ascomata with a flattened apical disk; ascospores (19.5–)21.5–24.5(–25.5) $\times$ 4–5 $\mu$ m
7 (6). 7.	Ascospores (1–)5–7(–11)-septate, (24–)30–60(–110) $\times$ 4–7(–8) $\mu$ m long; ascomata white to pale yellow
8 (4). 8.	Ascospores finely striate, $7.5-9 \times 2.5-3$ µm; ascomata umber, with an acute apex, finely scurfy due to free ends of hyphae
9 (8). 9,	Ascomata white, 150 $\mu$ m diam or less; ascospores ellipsoid to oblong, (7.5–)8.5–10(–11) $\times$ 2–2.5(–3) $\mu$ m, smooth-walled
10 (9). 10.	Ascomata sienna, with short, sinuous hairs extending from the wall; ascospores oblong to narrowly ellipsoid, $7-9 \times 2-2.5 \mu m$ , smooth-walled

KALLICHROMA Kohlm. & Volkm.-Kohlm., Mycol. Res. 97: 759. 1993.

Type: K. tethys (Kohlm. & E. Kohlm.) Kohlm. & Volkm.-Kohlm. (≡ Hydronectria tethys Kohlm & E. Kohlm.).

Ascomata depressed subglobose to ellipsoid, at first immersed, erumpent, ostiolate, periphyses surrounded by a gelatinous matrix, non-papillate, indistinctly clypeate, fleshy-leathery, yellow-orange, gregarious or frequently confluent; clypeoid tissue extending from peridium around the ostiole and often connecting with adjacent ascomata. Ascomatal wall of three regions: outer region of polygonal cells with large lumina; middle region of thickwalled cells with small lumina; inner region of elongate, flattened, thick-walled cells, forming a textura angularis. Apical paraphyses present, septate, simple, apically attached, merging with the periphyses. Asci subcylindrical to clavate, thin-walled at maturity except for the narrow, thick-walled persistent apex, without apical apparatus, maturing successively on the ascogenous tissue at the bottom of the centrum, 8-spored, ascospores biseriate. Ascospores ellipsoid, equally 1-septate, hyaline, longitudinally striate by thin ribs or smooth, with or without at early dissolving mucilaginous sheath. Anamorph un

known. On driftwood, mangrove roots, and submerged wood and branches in marine habitats. Description modified from Kohlmeyer & Volkmann-Kohlmeyer (1993). Notes.— Kohlmeyer & Volkmann-Kohlmeyer (1993) established this genus for two taxa previously described in Hydronectria. They provided a thorough account of the genus including the type, K. tethys, and one additional species, K. glabrum (Kohlm.) Kohlm. & Volkm.-Kohlm. (≡ Hydronectria tethys var. glabra Kohlm.). Kallichroma tethys occurs in warm temperate and tropical regions of the Atlantic, Indian and Pacific Oceans, while K. glabrum has been less commonly collected, known only from the Indian Ocean and the Pacific Ocean on the east coast of Australia.

LASIONECTRIA (Sacc.) Cooke, Grevillea 12: 111. 1884.

≡ Nectria subgenus Lasionectria Sacc., Syll. Fung. 2: 505.

Lectotype, designated by Clements & Shear (1931): L. mantuana (Sacc.) Cooke (≡ Nectria mantuana Sacc.).

Ascomata non-stromatic, superficial, orange to dark red-orange or dark brown, slightly darker in KOH but not KOH+, subglobose to globose, collapsed slightly cupulate when dry or not, often with fasciculate and/or solitary hairs. Ascomatal wall generally more than 20 μm thick, of two regions: outer region of thick-walled, pigmented cells; inner region of elongate, thin-walled, hyaline cells. Asci clavate. Ascospores broadly ellipsoid, 1-septate, hyaline, generally smooth-walled. Anamorph, where known, Acremonium. On dead woody and herbaceous substrata including basidiocarps.

Notes.— Saccardo (1883) established Nectria subgenus Lasionectria for nine species of Nectria having hairs on the ascomata. Cooke (1884) listed nineteen species in Lasionectria including many of the original nine species; he divided the genus into two subgenera based on ascospore septation. The presence of hairs on the ascomata is not a characteristic that reflects relatedness, thus most species previously placed in Lasionectria are excluded from the genus. Clements & Shear (1931) considered Dasyphthora Clem., Epinectria Syd. & P. Syd., and Neohenningsia Koord, to be later synonyms of Lasionectria. Based on an examination of their type specimens, none of these genera are synonyms of Lasionectria and they are discussed elsewhere. The type specimen of Lasionectria mantuana was examined. This species is similar to species previously placed in the Nectria sylvana-group (Samuels, 1976b) for which Lasionectria provides a generic name. The genus Lasionectria is distinct among genera in the Bionectriaceae in that the ascomatal wall is composed

of thick-walled cells each with a small lumen. Three species are included in Lasionectria as described below.

Lasionectria mantuana (Sacc.) Cooke, Grevillea 12: 111. 1884. - Plate 2. e; Plate 5. f. g.

■ Nectria mantuana Sacc., Michelia 1: 52, 1877.

Ascomata superficial with bases slightly immersed on decorticated wood fibers, solitary or in groups of 2-3; ascomata subglobose, collapsed cupulate when dry, 150 μm high × 250 μm diam, dark red-orange, becoming slightly darker in KOH but not blood-red or purple, without papilla. Ascomatal wall appearing roughened due to both short, fasciculate and solitary hairs. Ascomatal wall 18-35 µm thick, of two regions: outer region 12-15 µm thick, of thick-walled cells forming a textura angularis, walls pigmented in the upper portion of the ascomata, hyaline towards the base, about 1.5 μm thick, each with a small lumen; inner region 8-21 µm thick, cells elongate, forming a thin-walled textura angularis to textura prismatica. Short, solitary hairs extending from the outer region, hairs 12-18 µm long × 3-3.5 µm wide, 0-1-septate, slightly flexuous to wavy, hyaline, tapering slightly from the base to the broadly rounded apex. Around the apex beset with sparse, fasciculate hairs, 30-36 µm long × 15-22 µm wide at the base, individual hyphae 3-4.5 µm wide with rounded apices. Asci clavate, 42 × 6 µm, apex simple, 8-spored, ascospores uniseriate. Ascospores broadly ellipsoid, 8.5-9.5 × 3-3.5 μm, 1-septate, slightly constricted, hyaline, smooth (although one striate spore was seen).

HABITAT AND DISTRIBUTION.— Known only from the type specimen.

HOLOTYPE. TTALY. Mantova, Migliaretto, on decorticated poplar wood, Feb 1873, A. Magnaguti-Rondinini (PAD).

ILLUSTRATIONS, - Weese (1916, Figs. 4 A-C).

Lasionectria sylvana (Mouton) Rossman & Samuels,

= Nectria sylvana Mouton, Bull. Soc. Roy. Bot. Belgique 39: 49. 1900. = Calonectria fimbriata Seaver & Waterston, Mycologia 32:

Anamorph: Acremonium sp.

404, 1940,

Ascomata superficial on sparse subtending hyphae, hyphae septate, branched, smooth, ca 3 µm diam, walls ca 1 μm thick. Ascomata urniform, ca 150 μm high × (185-)210-250 μm across the flat top, superficial, solitary or in groups of 2-4, orange, becoming slightly collabent, when dry; papilla lacking or very short and acute; ostiolate, apex flattened, of cylindrical, septate, unbranched hyphae, tips of the hyphae rounded, ca 2 μm wide. Ascomatal wall 15-25 μm thick, cells in longitudinal section indistinct, walls ca 1  $\mu$ m thick; outer wall cells producing ascomatal hairs. Hairs orange, up to 100  $\mu$ m long, ca 3  $\mu$ m wide, walls ca 1  $\mu$ m thick, septate, unbranched, straight, smooth, solitary or bound in fascicles and forming triangular projections. Asci clavate to fusiform,  $55-75\times6-9$   $\mu$ m, apex simple, 8-spored, ascospores biseriate. Ascospores fusiform–ellipsoid,  $(9-)11-15(-17)\times(2.5-)3-4(-5)$   $\mu$ m, equally 2-celled, not constricted or slightly constricted at the septum, hyaline, smooth-walled.

Anamorph.— Conidiophores arising from the agar surface and aerial hyphae, monophialidic, unbranched or rarely verticillately branched, straight or undulate below the tip, smooth,  $(40-)45-55(-78) \times (2-)2.5-3$  (-4)  $\mu$ m wide at the base. Phialides terminal,  $(17-)37-50(-54) \mu$ m long,  $1-2 \mu$ m wide at the unflared tip. Conidia ellipsoid to nearly cylindrical, rarely elongating,  $(3-)5-7(-11) \times (1.5-)2-3 \mu$ m, unicellular, without a visible basal abscission scar, hyaline, arising in basipetal succession, held in a solitary, hyaline drop of liquid.

Habitat.— On decaying herbaceous stems.

DISTRIBUTION.— Belgium, Bermuda, Germany, New Zealand, and United States (New York).

Types.— BELGIUM. Near Liège, on stems of Angelica sylvestris, no. 21 (BR, holotype of N. sylvana); BERMUDA. On dead stems of Foeniculum vulgare (NY, holotype of Calonectria fimbriata).

ADDITIONAL SPECIMENS EXAMINED.— GERMANY. Kirnitschthal: near Schandow, on stems of Calamagrostis arundinacea, 6 June 1898, Krieger, Fungi Saxonici no. 1421, as Nectria graminicola Berk. & Broome (NY). UNITED STATES. New York: Fort Hunter, Erie Canal State Park, Yankee Hill Lock, on Equisetum arvense, 29 July 1972, Haines 2191 (NY). — NEW ZEALAND. Westland Prov., ex rachis of Cyathea smithii, G.J.S. 74-75 = CBS 566.76.

ILLUSTRATIONS.— Samuels (1976a, Fig. 14; 1976b, Fig. 18, both as N. sylvana); Seaver & Waterston (1940, Fig. 3 lower, as C. fimbriata).

Notes.— The Acremonium anamorph known in culture (CBS 566.76) is present on the type specimen and on the Haines collection. Neohenningsia stellulata Koord. and N. brasiliensis Henn., previously regarded as synonyms of L. sylvana (as N. sylvana) by Samuels (1976 a, b), are here considered synonyms of Hydropisphaera rufofusca.

Lasionectria vulpina (Cooke) Rossman & Samuels, comb. nov.

- ≡ Nectria vulpina (Cooke) Ellis, North Amer. Fungi no.
  774, 1882.
- ≡ Peziza vulpina Cooke, Hedwigia 14: 82, 1875.
- ≡ Dialonectria vulpina (Cooke) Cooke, Grevillea 12: 83.
  1884.
- ≡ Nectriella vulpina (Cooke) Sacc., Syll. Fung. 9: 941.
  1891.
- = Nectria incrustans Weese, Z. Gärungsphysiol. 1: 144. 1912.

Anamorph: Acremonium sp.

Mycelium white, flat, scanty, hyphae septate, branched, 3-4 µm wide, thin-walled. Ascomata urniform, (155-)190-220(-230) µm high × (200-)240-270(-375) µm across the flattened apices, solitary or in groups of a few, orange, becoming slightly cupulate when dry; papilla lacking or very short; ostiolar area formed by cylindrical, septate unbranched hyphae with rounded tips, ca 2 μm wide; periphyses 15-20 μm long. Ascomatal wall when dry shining, 15-25(-40) μm thick; cells ellipsoid, 5-7  $\times$  3-4  $\mu$ m, with ca 1  $\mu$ m thick walls, cells becoming progressively more flattened toward the interior. Hairs orange, 10-50 µm long, 3-5 µm wide, with ca 1 µm thick walls, septate, unbranched, straight, solitary, or in triangular fascicles. Asci broadly cylindrical to clavate, 37–58 × 6–8(–10) μm, 8-spored; with an apical ring, ascospores obliquely uniseriate, becoming irregularly arranged. Ascospores fusiform-ellipsoid,  $(7-)8-11(-13) \times 3-4 \mu m$ , equally 2-celled, slightly constricted at the septum, hyaline, stri-

Anamorph.— Conidiophores arising from surface of the agar and the aerial mycelium; monophialidic, unbranched, aseptate, straight, smooth, 49–57 μm long, 2.5–3 μm wide at the base. Conidiogenous cells phialidic, 44–52 μm long, 1–1.5 μm wide at the unflared apex. Conidia ellipsoid to cylindrical, 5–7 × 1.5–2 μm, unicellular, hyaline, lacking a visible basal abscission scar, arising in basipetal succession, held in a solitary pink to salmon or hyaline drop of liquid.

HABITAT.— On decorticated, deciduous wood and on basidiocarps of Stereum subpileatum.

### KEY TO THE SPECIES OF LASIONECTRIA

- 2. Ascospores striate, 8-11 × 3-4 μm; ascomatal hairs 10-50 μm long ...... L. vulpina
- Ascospores smooth, 8.5–9.5 × 3–3.5 μm; ascomatal hairs 12–36 μm long ... L. mantuana

Distribution.— Common in temperate regions.

TYPES.— UNITED STATES. New Jersey: Newfield, on fallen apple wood, 6 Dec 1874 (NY, lectotype of *P. vulpina*, designated in Samuels, 1976a). GERMANY. Mark Brandenburg: Triglitz in der Prignitz, on *Alnus glutinosa*, 6 Oct 1908, Jaap, Höhnel 5519 (FH-Höhnel), lectotype of *Nectria incrustans*, designated in Samuels, 1976a); same location, on *Betula*, 1 Oct 1909, Jaap, Höhnel 5519 (FH, paratype of *N. incrustans*). Culture: CBS 565.76.

ILLUSTRATIONS.— Samuels (1976a, Figs. 9f, 15; 1976b, Figs. 6, 29, both as N. vulpina).

MYCOARACHIS Malloch & Cain, Canad. J. Bot. 48: 1820, 1970.

Type: M. inversa Malloch & Cain.

Ascomata subglobose to globose, dark olive-green to black, non-ostiolate. Asci subglobose to globose, irregularly disposed, 8-spored. Ascospores one-septate, hyaline, smooth-walled. Conidia borne in clusters at the tips of simple phialides, one-celled, hyaline, smooth. Anamorph Acremonium. Isolated from herbivore dung.

Notes.— The unispecific genus *Mycoarachis* was described in the cleistothecial family *Pseudeurotiaceae* and distinguished by dark olive-green to black ascomata, hyaline, one-septate ascospores, and an *Acremonium* anamorph. Later Benny & Kimbrough (1980) suggested that the fungus belonged in the *Hypocreales* because of the two-celled ascospores and *Acremonium*-like anamorph. Rehner & Samuels (1994, 1995) included the type culture in their molecular study and confirmed that *M. inversa* is a cleistothecial member of the *Hypocreales* related to *Bionectria*. Ogawa *et al.* (1997) reported the affinity of another cleistothecial genus, *Emericellopsis*, with *Mycoarachis* in a subclade of the *Bionectriaceae*.

Mycoarachis inversa Malloch & Cain, Canad. J. Bot. 48: 1822, 1970.

Anamorph: Acremonium sp.

Mycelium hyaline, hyphae septate, branched, 1–3 μm wide, occasionally developing very thick walls near the septa and swelling up to 7 μm wide. Ascomatal initials at first simple coils with filaments about 1–3 μm wide, later becoming compact and contorted as a result of abundant proliferation. Ascomata subglobose to globose, 50–200 μm diam, metallic in reflected light when dry, dark green to black, non-ostiolate; ascomatal wall 11–30 μm thick, consisting of a pale outer region and a darker inner region; outer region of hyaline to pale brown cells, 3–22 μm diam; inner region of dark brown, flattened, cells, 4–20 × 2–10.5 μm. Asci subglobose to globose, 5.5–11 μm diam, evanescent, 8-

spored. Ascospores cylindrical to broadly ellipsoid,  $5-5.5 \times 3-3.5 \mu m$ , with a constricted, single median septum dividing the spore into two equal globose cells, hyaline, smooth.

Anamorph.— Conidiophores arising from submerged mycelium, simple or sparingly branched, septate, tapering from the base to the apex, ending in a phialide,  $14-35 \times 2-3.5$  µm. Conidia ellipsoid, fusoid, ovoid, cylindrical or allantoid,  $3-10 \times 1.5-3.5$  µm, non-septate, hyaline, smooth-walled, borne in moist clusters at the tip of the conidiophores. Description modified from Malloch & Cain (1970).

Habitat.— Isolated from cow-, elephant- and unidentified herbivore dung.

DISTRIBUTION.— Tanzania, Uganda, and United States (Nebraska), as reported by Malloch & Cain (1970).

HOLOTYPE.— UGANDA. Mweya Lodge: Queen Elizabeth National Park, isolated from elephant dung, 27 July 1966, Cain, Griffin & Krug (TRTC 66.2166f, not examined). Culture CBS 517.70.

Illustrations.— Malloch & Cain (1970, Figs. 20–25).

MYCOCITRUS A. Möller, Bot. Mitt. Tropen 9: 297. 1901.

Type: M. aurantium A. Möller.

= Shiraiella Hara, Bot. Mag. (Tokyo) 28: 274. 1914. — Type: S. phyllostachydis (Syd. & P. Syd.) Hara (≡ Ustilaginoidea phyllostachydis Syd. & P. Syd.), recognized as Mycocitrus phyllostachydis (Syd. & P. Syd.) Doi).

Stroma well-developed, buff to rufous, clasping and surrounding the substratum. Ascomata immersed, with apices barely visible, densely gregarious, forming a single layer. Asci cylindrical, ascal apex simple. Ascospores ellipsoid, 1-septate, hyaline, spinulose. Anamorph Acremonium. On living stems of bamboo.

Notes.— Mycocitrus is characterized by ascomata partially to fully immersed in the upper region of large, fleshy stromata that clasp and surround bamboo stems. Möller (1901) illustrated an Acremonium-like anamorph. Although the type specimen of M. aurantium probably no longer exists, an excellent illustration serves to characterize this species. The unispecific genus Shiraiella was established for Ustilaginoidea phyllostachydis. Doi (1967a) was unable to locate the type specimen of U. phyllostachydis at B suggesting that it was destroyed. He designated a non-type specimen examined by Hara (TNS 209286) as the neotype and transferred U. phyllostachydis to Mycocitrus. Mycocitrus is recognized with these two species.

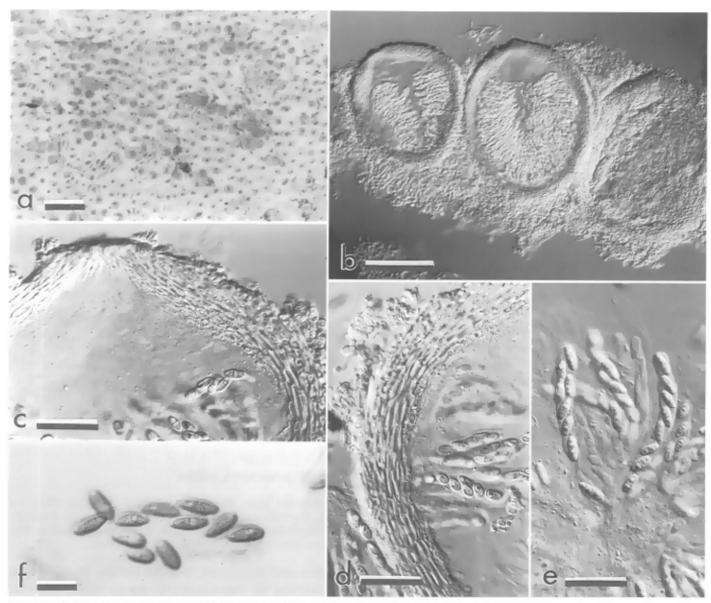


Plate 6. a-f. Mycocitrus aurantium. a. Surface view of stroma with ostioles of immersed ascomata. b. Median section of ascomata and stroma. c. Median section of ascomatal apex. d. Median section showing ascomatal wall. e. Asci with ascospores and remnants of apical paraphyses. f. Asci with ascospores. a, c-f. BPI Lloyd 37616, b. Rick 117 – BPI. Scale bars:  $a = 500 \mu m$ ;  $b = 100 \mu m$ ;  $c-e = 25 \mu m$ ;  $f = 10 \mu m$ .

Mycocitrus aurantium A. Möller, Bot. Mitt. Tropen 9: 297. 1901. — Plate 6, a–f.

Anamorph: Acremonium-like.

Stroma very large, 7–10 cm diam, globose, surface buff to rufous, clasping and surrounding the substratum, internal tissue hyphal, white. Ascomata partially to fully immersed at surface, apices barely visible, densely gregarious, formed in a single layer, yellow to orange, KOH–; ascomatal apex not sharply differentiated, ostiolar opening formed of narrowly clavate cells, continuous with periphyses. Ascomatal wall ca 20 µm thick, forming a single region of cells of intertwined, hyaline, thick-walled hyphae, cell lumina ellipsoid to fusiform, 2.5–10 µm long × 2–2.5 µm wide, with outer cells be-

coming conspicuously hyphal. Asci  $45-60 \times 4.5-8.5$   $\mu$ m, ascal apex simple, eight-spored, irregularly biseriate. Ascospores ellipsoid,  $(6-)7.5-10(-11) \times 3.5-4.5$   $\mu$ m, 1-septate, hyaline, spinulose.

Habitat.— On living stems of bamboo. Distribution.— Brazil.

Types.—BRAZIL. Blumenau: on living stems of bamboo, A. Möller (iconotype: Möller, 1901, Tafel II, Figs. 38a-f, Tafel IV, Fig. 45); São Leopoldo: on living *Arundinaria*, 1905, Rick, Fungi austro-americani 117 (BPI, in bound set, **epitype** designated herein; BPI 631727, FH-General, FH-Höhnel, FH-Patouillard, isoepitypes).

ADDITIONAL SPECIMEN EXAMINED.— BRAZIL. Rio Grande Do Sul: P. Pio Buck, 14317, Lloyd 37616 (BPI 744967).

ILLUSTRATION. - Müller & von Arx (1962, Fig. 257).

## KEY TO THE SPECIES OF MYCOCITRUS

- Stromata 5–9 mm long × 4–5 mm diam, elongate, tuberculate; ascospores 6.5–9 × 3.5–5.5 Mm, hyaline, smooth to slightly roughened; on *Phyllostachys* in Japan. *M. phyllostachydis*

Notes.— The type specimen of Mycocitrus aurantium, deposited either at B or HBG, was apparently destroyed. The detailed illustration accompanying the type description is considered the iconotype, with the Rick exsiccata herein designated as the epitype. In these and other specimens, the ascomata develop only at the surface of a very large stroma. In the illustrations by Möller (1901) and Müller & von Arx (1962), the perithecia are shown to be immersed at the surface and below it in several layers. However, in all the specimens examined, the ascomata were found to be irregularly arranged but developing only at the surface of the stroma. If the extensive stroma were sectioned obliquely, the ascomata might appear to be distributed as shown in these illustrations.

Mycocitrus phyllostachydis (Syd. & P. Syd.) Doi, Bull Natl. Sci. Mus. 10: 31. 1967.

≡ Ustilaginoidea phyllostachydis Syd. & P. Syd., Mem. Herb. Boiss. 4: 5. 1900.

≡ Hypocreopsis phyllostachydis (Syd. & P. Syd.) Miyake & Hara, Bot. Mag. (Tokyo) 24: 333. 1910.

Shiraiella phyllostachydis (Syd. & P. Syd.) Hara, Bot. Mag. (Tokyo) 28: 402. 1914.

Anamorph: Acremonium-like.

Doi (1967a) provided a good description and illustrations of M. phyllostachydis including the anamorph.

NECTRIELLA Nitschke, in Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 175. 1869 [1870].

Lectotype, designated by Seaver (1909a): N. fuckelii Nitschke. Ascomata immersed to partially erumpent, scattered or in groups, non-stromatic, obpyriform, less often subglobose, 100-500 µm diam, white, pale yellow, pale red or pale brown, not changing color in KOH or lactic acid or rarely reacting weakly, ostiolate, collapsing vertically by lateral pinching or not collapsing. Ascomatal wall 10-30(-40) μm thick, often of two intergrading regions: outer region of thick-walled, angular to rounded cells; inner region of thin-walled, elongate cells. Ascomatal apex of parallel rows of vertically elongate cells, continuous with the inner wall region; cells increasingly narrow, merging with periphyses at the interior, somewhat expanded or clavate at the exterior. Gelatinized remains of apical paraphyses sometimes seen. Asci clavate, apex usually with a ring, 8-spored, ascospores usually biseriate in the middle, uniseriate above and below, occasionally entirely uniseriate. Ascospores navicular, ellipsoid or fusiform, 1-septate, rarely non-septate, hyaline, smooth, faintly spinulose, verrucose or longitudinally striate. Anamorph, where known, Acremonium-like or Kutilakesa. On dead woody and herbaceous substrata.

Notes.— Nectriella was established for species of Nectria that are immersed in the substratum. Lowen (1991) recircumscribed the genus based on a study of the type species; she provided descriptions and illustrations of over twenty species in Nectriella. Nectriella is distinguished from other hypocrealean genera by the nonstromatic, immersed, KOH-, ascomata with walls often composed of two regions. Species of Nectriella generally have small, pale yellow, inconspicuous ascomata and occur on dead wood, herbaceous debris, rarely on fungi or as plant parasites, as in N. pironii (Alfieri & Samuels, 1979). It is expected that many species have yet to be discovered. Because species of Nectriella are immersed in the substratum, they tend to lose characteristics of the ascomata that might be useful in placing them taxonomically, thus species now classified in Nectriella may not be closely related.

Nectriella fuckelii Nitschke, in Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 175. 1869 [1870]. — Plate 7, a–f.

≡ Calonectria fuckelii (Nitschke) Sacc., Michelia 1: 310.
1878.

Ascomata solitary or in groups of up to 10, immersed to partially erumpent with emergent papilla, difficult to remove from substratum, globose to obpyriform, 220-400 µm high × 200-410 µm diam, at first pale pink, then pale yellow, KOH-; papilla rounded to flattened, 88-168 μm high × 150-200 μm diam, of loosely joined hyphal elements, ca 3.5-4.5 µm wide, with rounded ends that extend to same height, scattered, very thick-walled, unbranched elements arising from and extending beyond the margin of the ascomata. Setae clavate, 24-64 μm long × 4-5 μm wide, tapering to 3 μm at the base, sparingly septate, hyaline, brittle, with round apex and 1.5-2 µm thick walls. Ascomatal wall 21-28 μm thick, of two regions: outer region 14–18  $\mu$ m thick of thick-walled, rounded cells, 3–7  $\times$ 2-8 µm, with 2 µm thick walls, fusing and terminating

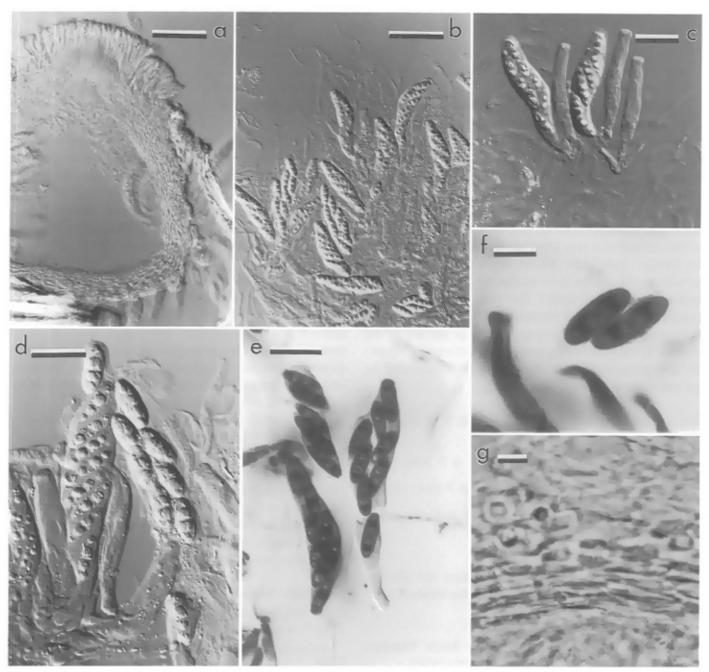


Plate 7. a-f. Nectriella fuckelii. a. Median section of ascoma. b. Asci with deliquescing apical paraphyses. c, d. Asci with ascospores. e. Asci with ascospores stained in cotton blue. g. Nectriella utahensis. Median section of ascomatal wall. a-f. Isolectotype – BPI. g. Holotype – NY. Scale bars: a, b =  $50 \mu m$ ; c =  $25 \mu m$ ; d, e =  $20 \mu m$ ; f, g =  $10 \mu m$ .

at the upper side of the papilla; inner region 8–10  $\mu$ m thick, of elongate cells 3.5–10.5 × 1–5  $\mu$ m. Apical paraphyses visible among mature asci, narrowly filamentous. Asci clavate, 68–95 × 12–19  $\mu$ m, constricted at the base, apex rounded to truncate, with a ring; ascospores biseriate. Ascospores narrowly ellipsoid, (12–)16–19 × (5.5–)6–7(–8)  $\mu$ m, 1-septate, at first hyaline, then slightly yellow, spinulose due to collapse of the narrow sheath.

Habitat and distribution.— Known only from the type collection.

Types.— GERMANY. Hessen (formerly Nassau): Rheingau, on dead but still hard wood of *Populus nigra*, spring, Fuckel, Herb. Boissier No. 915 as *Calonectria fuckelii* (FH, lectotype, designated by Rossman, 1979b; B, BPI, FH – Höhnel, S, isolectotypes).

Twenty-one additional species are included in *Nectriella* based on Lowen (1991), three of which are described here as new species.

Nectriella alpina (G. Winter) Weese, Ann. Mycol. 12: 148. 1914.

■ Nectria alpina G. Winter, Hedwigia 19: 175. 1880.

Anamorph.— None known.

Ascomata immersed, becoming erumpent, scattered or in groups of 5-6, obpyriform, 220-260 µm high × 195-250 μm diam, at first orange-yellow, then yellow with brownish ostiole, KOH-; papilla truncate, 50 μm high × 100-150 µm diam, apex of clavate, diverging, thin-walled hyphae,  $10-17.5 \times 3-5 \mu m$ , 0-2 septate, ends free. Cells on surface consisting of intertwined hyphae. Ascomatal wall 10-12 µm thick, of two regions: outer region of intertwined hyphae with ellipsoid cells  $5-9 \times 1.5-3.5$  µm; inner region of parallel hyphae,  $0.5-1.5 \mu m$  wide. Asci clavate,  $60-89 \times 9-12.5 \mu m$ ; apex rounded, with an inconspicuous apical ring, ascospores biseriate. Ascospores ellipsoid. (12.5-)  $13-17.5(-19) \times 3.5-5(-7) \mu m$ , 1-septate, sometimes slightly constricted, with upper cell often wider than lower cell, hyaline, spinulose.

Habitat.— On basal leaves and stems of Arabis and Saxifraga.

DISTRIBUTION. — Austria, Switzerland.

Lectotype, designated herein: SWITZERLAND. Grisons: Rhaetia, Albula near Hospiz, in wilted and dry leaves of Arabis pumila (Brassicaceae) associated with Pleospora pyrenaica Niessl, May 1880, G. Winter (NY).

ADDITIONAL SPECIMEN EXAMINED.— AUSTRIA. Steiermark: Eisenerzer Alpen, Reiting W von Trofaiach, NE-Abhang des Grieskogels, ca 2050 m, Caricetum firmae, on Saxifraga paniculata, 9 July 1984, J, Hafellner & A. Nograsek (GZU 140–88).

Notes.— The species was recollected and redescribed by Nograsek (1990).

Nectriella balansiae R.H. Arnold, Mycologia 59: 248. 1967.

ANAMORPH: None known.

Ascomata immersed in stromata and occasionally in empty ascomata of *Balansia*, obpyriform, 110–128 μm high × 70–120 μm diam, nearly white, KOH-; papilla 30–66 μm high × 16–30 μm diam. Ascomatal wall 12 μm thick, of one region of elongate cells 6 × 2 μm, with 1 μm thick walls; periphyses prominent. Asci clavate, 31.5–40 × 5.5 μm; apex truncate, with a ring; ascospores biseriate. Ascospores irregularly ellipsoid–fusiform, 9.5–10.5 × 2.5–3 μm, typically 1-septate, one cell often narrower, hyaline, smooth, two prominent guttules per cell.

Habitat.— Fungicolous, on stromata of *Balansia*.

Distribution.— Known only from the type collection.

LECTOTYPE, designated herein.— CENTRAL AFRICAN REPUBLIC ('French Congo'). Kouti Region [near Ndéle], journey II, immersed in the stromata of *Balansia volkensii* on high herbs [grass], 15 Nov 1891, J. Dybowski, Patouillard Herb. 597 (FH, as *Hyalodothis clavis*).

Notes.— Nectriopsis epichloë (Speg.) Samuels also occurs on Balansia and produces similarly sized and shaped asci and ascospores, but differs from Nectriella balansiae by yellow, superficial ascomata and ascospores that lack prominent guttules.

Nectriella bloxamii (Berk. & Broome) Fuckel, Jahrb. Nassauischen Vereins Naturk. 29–30: 21. 1876–1877 [1877].

≡ Nectria bloxamii Berk. & Broome, Ann. Mag. Nat. Hist., Ser. 2, 13: 467, 1854.

= Nectria umbelliferarum P. Crouan & H. Crouan, Fl. Finistère, p. 37. 1867.

= Nectria heraclei P. Crouan & H. Crouan, Fl. Finistère p. 38, 1867

= Nectria fuscidula Rehm, Hedwigia 21: 199. 1882.

= Nectria dacrymycelloides Rehm, Hedwigia 42: 175. 1903. [≡ Nectriella dacrymycelloides (Rehm) Höhn. & Weese, Ann. Mycol. 8: 465. 1910, comb. inval., Art. 33.1].

Ascomata immersed, becoming erumpent, scattered or in groups of up to 8, subglobose, 250–300 μm diam, pale at first, orange-brown, then tan, KOH-; collapsed vertically, nonsetose. Cells on surface of ascomata angular, ca 10 μm diam. Ascomatal wall 20 μm wide, of two regions: outer region 12 μm thick, of thick-walled, angular to rounded cells, 10 μm diam; inner region 8 μm thick, of thin-walled, elongate cells. Asci clavate, 40–70 × 8–9(–12) μm; apex truncate, with a ring; ascospores biseriate. Ascospores ellipsoid, 16–24 × 3–5 μm, 1-septate, often slightly constricted, hyaline, pale orange in mass when fresh, smooth to slightly roughened, often many guttules per cell.

ANAMORPH.— None known.

Habitat.— On dead stems of herbaceous dicotyledonous plants.

DISTRIBUTION.— Europe, New Zealand.

Type specimens.— UNITED KINGDOM. England: Leicestershire, Twycross, on Jerusalem artichoke (Helianthus tuberosus L., Asteraceae), 16 Nov 1855, Rev. A. Bloxam 781 (holotype of N. bloxami K; isotypes: IMI 52290, slides; K, 3 collections; K, BM, PC, all as Sphaeria (Nectria) bloxami); FRANCE. Brittany: Finistère, on dead stems of Oenanthe crocata, 20 June 1864, Crouan & Crouan (lectotype of Nectria umbelliferarum, designated herein, CO); on dead stems of Urtica, 18 June 1864, Crouan & Crouan (paratype of Nectria umbelliferarum, CO); on stem of Umbelliferae (Apiaceae), 30 Jan 1862, Crouan & Crouan (paratype of Nectria umbelliferarum, CO); on stems of Heracleum sphondylium, 27 Mar 1863, Crouan & Crouan (holotype of Nectria heraclei, CO).

Notes.— Nectriella bloxamii can be confused with N. luteola in macroscopic appearance but is distinguished by longer ascospores, differences in substrata (herbaceous stems versus leaf veins and petioles) and the color of the ascomata. Nectriella dacrymycella has orange, translucent ascomata that usually remain under the epidermis of the substratum and have a more thickened

apex due to the globose, thick-walled cells of the outer region of the ascomatal wall.

## Nectriella crouanii Lowen, nom. nov.

≡ Nectria aurea P. Crouan & H. Crouan, Fl. Finistère, p.
37. 1867 [non Nectriella aurea Sacc. & Speg. 1878].

Ascomata immersed, often becoming erumpent, scattered, subglobose, 100–200 μm diam, yellow; ascomatal apex red, becoming darker in KOH, papilla truncate. Ascomatal wall 21–23 μm thick, of two regions at the apex and sides, one at the base: outer region 9–13 μm thick; walls indistinct; inner region 10–12 μm thick, of thin-walled, elongate cells. Asci clavate, 44 × 4–5 μm; apex truncate, with a ring; ascospores biseriate. Ascospores ellipsoid to fusiform, 12 × 3 μm, 1-septate, not constricted, hyaline, smooth-walled, with two guttules per cell.

ANAMORPH. - None known.

Habitat. - Rubus and 'moulin'.

DISTRIBUTION.— France, known only from the two Crouan collections.

HOLOTYPE.— FRANCE. Brittany: Tenfeld, on small branches of Rubus, 26 Feb 1863, Crouan & Crouan (CO).

ADDITIONAL SPECIMEN EXAMINED.— FRANCE. Brittany: Vallon, on white 'moulin', 8 March 1866, Crouan & Crouan (CO).

Notes.— Nectriella crouanii and N. exigua are distinguished from other species of Nectriella by the ascomatal walls that have two regions at the apex and sides and by asci that average less than 45 µm long. Nectriella crouanii differs from N. exigua by the substratum and the ascomata that are red at the apex and yellowish on the sides and base.

#### Nectriella curtisii (Berk.) Lowen, comb. nov.

≡ Nectria curtisii Berk., Grevillea 4: 46. 1875.

= Nectria lacustris Kirschst., Ann. Mycol. 34: 186. 1936.

≡ Nectriella lacustris (Kirschst.) Magnes & Hafellner, Biblioth. Mycol. 139: 105. 1991.

Notes.— Magnes & Hafellner (1991) provided a recent description and illustrations of this species as N. lacustris.

Nectriella dacrymycella (Nyl.) Rehm, Ascomyceten no. 232, 1874.

≡ Sphaeria dacrymycella Nyl., Flora 46: 332, 1863.

Ascomata immersed, scattered or in groups of up to 20, subglobose,  $140-200~\mu m$  high  $\times~180-230~\mu m$  diam, bright orange, remaining covered by host epidermis, KOH–. Cells on the surface of the ascomata angular, 6–10  $\mu m$  diam. Ascomatal wall 10  $\mu m$  thick, up to 20

 $\mu m$  at the apex as expanded outer region of rounded, thick-walled cells; inner region of elongate cells; ostiolar region of thick-walled, angular to rounded cells, outer cells merging with the substratum. Asci clavate,  $50-75 \times 9-10~\mu m$ , apex truncate, with a ring; ascospores biseriate. Ascospores fusiform-ellipsoid,  $13-20 \times 4-5.5~\mu m$ , 1-septate, sometimes slightly constricted, curved, at first hyaline, then pale brown, smooth to slightly roughened, sometimes with 2 guttules per cell.

ANAMORPH .- None known.

Habitat.— On herbaceous stems.

DISTRIBUTION.— Europe.

HOLOTYPE.— FINLAND. Tavastia australis, Lempäälä, on stems of *Urtica*, 9 Sep 1860, P.A. Karsten (H 2236; isotypes, IMI 52313, slide; H 2237).

ADDITIONAL SPECIMENS EXAMINED.— CZECH REPUBLIC. Moravia: Mähr. Weißkirchen, on Urtica dioica, Oct 1926, F. Petrak (C; as Nectria dacrymycella). FRANCE. Brittany: on dead stems of Urtica, 22 Oct 1869, Crouan & Crouan (CO; as Nectria dacrymycella). UNITED KINGDOM. England: Bucks, on stems of Iris pseudacorus (IMI 96574); Isle of Lyigha, E. Bay at Tabet, on Iris pseudacorus. 12 May 1981, R.W.G. Dennis (K; as Nectria arenula); Norfolk: King's Lynn, on stem of Urtica dioica, 26 June 1979, A. Moore (K; as Nectria arenula); Norfolk: Norwich, on stems of Iris pseudacorus (IMI 70195); Wales: Monmouthshire: Gwernesey, on stems of Iris pseudacorus, (IMI 49092); Powys, Forden, on nettle stems, 27 Sep 1934, Vize 37 (K).

Illustration.— Ellis & Ellis (1985, Fig. 1541).

Notes.— Nectriella dacrymycella and Charonectria sceptri both have ascomata that look like orange blisters under the epidermis of herbaceous stems. Nectriella dacrymycella usually has smaller, non-papillate ascomata with a narrower lateral wall, shorter asci and, unlike C. sceptri, does not have true paraphyses. Ascomata of N. dacrymycella are found on Iris pseudacorus and Urtica dioica, whereas those of C. sceptri are known on species of Aconitum, Pedicularis sceptrum-carolinum, and other dead herbaceous plants. Nectriella dacrymycella has ascospores that overlap in size and shape with those of Nectriella bloxamii and N. luteola, but it is distinguished from the latter two species by ascomata that have a wider wall at the ascomatal apex and the bright orange pigment. Nectriella dacrymycella is illustrated and briefly described by Ellis and Ellis (1985).

#### Nectriella dakotensis (Seaver) Lowen, comb. nov.

≡ Hyponectria dakotensis Seaver, Mycologia 1: 20, 1909.

= Nectriella muelleri Samuels, Rogerson, Rossman & J.D. Sm., Canad. J. Bot. 62: 1899. 1984.

This species was described and illustrated by Samuels et al. (1984) as N. muelleri, including the Acremonium-like anamorph.

Nectriella exigua Dennis, Revista Biol. (Lisbon) 12: 22. 1983.

This species was described and illustrated by Dennis (1983).

Nectriella funicola (Berk. & Broome) Petch, Naturalist 970; 281, 1937.

≡ Sphaeria funicola Berk. & Broome, Ann. Mag. Nat. Hist., Ser. 2, 7: 188. 1851.

= Nectria charticola Fuckel, Fungi rhenani no. 990. 1864.

≡ Nectriella charticola (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23~24: 176. 1869 [1870].

= Nectria fibricola Plowr., in Sacc., Michelia 2: 152, 1880.

This species was described and illustrated by Booth (1959) and Ellis & Ellis (1985).

Nectriella galii (Plowr. & Harkn.) Lowen, comb. nov.

≡ Nectria galii Plowr. & Harkn., Bull. Calif. Acad. Sci. 1:
26. 1884.

Ascomata immersed, raising the epidermis of the stem, scattered, subglobose, 160  $\mu$ m high  $\times$  220–240  $\mu$ m diam, pale pink, KOH–; collapsing vertically, setae flexuous, septate, to 200  $\mu$ m long. Ascomatal wall 10  $\mu$ m thick, of two regions: outer region 8  $\mu$ m thick, of thickwalled, angular to rounded cells; inner region 2  $\mu$ m thick, of thin-walled, elongate cells. Asci cylindrical, 40–60  $\times$  4–8  $\mu$ m, apex truncate, simple, thin-walled, deliquescing early; ascospores overlapping, uniseriate. Ascospores ellipsoid, 10–12  $\times$  4–6  $\mu$ m, 1-septate, hyaline, smooth to slightly roughened, one guttule per cell, ascospores occurring in great numbers, filling the centrum.

Anamorph.— None known.

Habitat.— On stems of Galium.

DISTRIBUTION.— Known only from the type collection.

HOLOTYPE.— UNITED STATES. California: on Galium trifolium (Rubiaceae), H.W. Harkness 3070 (K).

Notes.— Nectriella galii has ascomata with an enlarged apical region typical of the genus Nectriella. Nectriella galii is similar to N. bloxamii and N. luteola, two other species that have subglobose ascomata. The ascomata of N. galii could be mistaken for pycnidia because the fragile asci rupture easily in a squash mount liberating great numbers of ascospores into the centrum.

Nectriella guttulata Lowen, Mem. New York Bot. Gard. 49: 244, 1989.

This species is described and illustrated by Lowen (1989).

### Nectriella halonata Lowen, nom. nov.

≡ Charonectria umbelliferarum Höhn., Hedwigia 42: 187. 1903 [non Nectriella umbelliferarum P. Crouan & H. Crouan 1867].

Ascomata immersed, scattered or in groups of 20 or more, obpyriform, 300–400  $\mu$ m high  $\times$  180–420  $\mu$ m diam, pale yellow, KOH–, papilla truncate, 60–170  $\mu$ m high  $\times$  80–140  $\mu$ m diam, collapsing vertically with the papilla retaining its shape; setae forming a circle around the ostiole, clavate, 10–80  $\times$  2–5  $\mu$ m, widening to 6  $\mu$ m at the apex, wall 1  $\mu$ m thick, apex rounded, thin-walled, base sometimes uneven, 0–2-septate, hyaline. Ascomatal wall 14–16  $\mu$ m wide, of one region of thin-walled, elongate cells. Asci clavate, 40–80  $\times$  6–10  $\mu$ m; apex thickened, with a ring; ascospores biseriate. Ascospores ellipsoid–fusiform, often slightly curved, (10–)15–20(–22)  $\times$  4–5.5  $\mu$ m, 1-septate, at first hyaline, then brownish yellow, distinctly verrucose when mature, with 1–2 guttules per cell.

Anamorph.— None known.

ETYMOLOGY.— Halonata, referring to the halo of setae on the papilla.

Habitat.— In herbaceous stems of dicotyledonous plants.

DISTRIBUTION.— Known from temperate regions, Europe, and U.S.A. (Colorado).

HOLOTYPE.— AUSTRIA. Tirol: Ötztal, Tumpener See, on dry stems of umbellifer, 8/99, 27 Aug 1902, Höhnel (FH, FH – Höhnel, slide).

ADDITIONAL SPECIMENS EXAMINED.— FRANCE. Côte-d'Or: Morvan plateau, edge of pond of Ste. Isabelle, on Angelica sylvestris, Aug 1891, F. Fautrey, Roumeguère Fungi Sel. Exs. 6049 (K, NY, as Nectria umbelliferarum). GERMANY. Bavaria: Hochvogel, Bärgündele-Alpe, (Allgäuer Hochalpen), on dry stems of umbellifer, ca 1300 m, 1909, Rehm, Ascomyceten 1867 (FH, K, NY as Charonectria umbelliferarum); Bavaria: München, Kiesgrube near Fürstenried, on dry stems of umbellifer, Oct 1902, Rehm, Rehm Ascomyceten no. 1466 (IMI 104344; K; as Calonectria bloxami). SWEDEN. Uppland: Dalby par., roadside ca 125 m W of Jerusalem, on Carlina vulgaris (Asteraceae), overwintered stems and leaves, 28 June 1988, K. & L. Holm 4941a (NY). UNITED KINGDOM. England: Suffolk: Dunwich Forest, on Angelica sylvestris, 21 Sep 1979, M.B. & P. Ellis (IMI 241564); Yorkshire: Pickering marshes, on Urtica dioica, 21 June 1956, W.G. Bramley (K; as Lasionectria). UNITED STATES. Colorado: location unknown, in herbaceous stems, 1910, Seaver & Bethel (NY; as Nectriella fuckelii).

Notes.— Nectriella halonata is similar to N. dakotensis in having ascomata ornamented with setae encircling the ostioles and collapsing vertically with the papillae retaining their shapes. Nectriella halonata has larger asci and ascospores than N. dakotensis. Although no ascomata remain in the holotype collection, there is a slide in the Höhnel herbarium that serves as the type.

The description herein is based primarily on Rehm, Ascomveeten 1867.

Nectriella jucunda (Durieu & Mont.) Sacc., Michelia 1: 278. 1878.

≡ Sphaeria jucunda Durieu & Mont., in Durieu, Explor. sci. Algérie, Bot. 1: 478, 1849.

■ Nectria jucunda (Durieu & Mont.) Mont., Syll, Gen. Sp. Crypt., p. 225, 1856.

= Hyponectria jucunda (Durieu & Mont.) Weese, in Höhnel & Weese, Ann. Mycol. 8: 466, 1910.

= Nectriella cacti Ellis & Everh., J. Mycol. 8: 66. 1902.

≡ Hyponectria cacti (Ellis & Everh.) Seaver, Mycologia 1: 20. 1909.

Ascomata immersed, scattered or in groups of up to 20, nonstromatic, subglobose, 360  $\mu$ m high  $\times$  250–300  $\mu$ m diam, pale red or orange to yellow, KOH–, apex truncate, ostiolar area sometimes darker. Surface cells of the ascomata epidermoid to angular, 5–10  $\mu$ m in the longest dimension. Asci clavate, 42–70  $\times$  3–4  $\mu$ m, apex truncate, simple; ascospores irregularly uniseriate to biseriate in the middle, uniseriate above and below, filling the upper two thirds of the ascus. Ascospores cylindrical to allantoid, ca 5.5  $\times$  1.5  $\mu$ m, unicellular, hyaline, smooth-walled.

Anamorph.— Leptodermella opuntiae Dodge (circumstantial).

Habitat.— In stems of *Opuntia*.

Distribution.— Algeria, U.S.A. (Alabama), Spain.

Types.— ALGERIA. Hill over Bab-Azoun, in stems of *Opuntia*, 22 Feb 1840, M.C. Durieu de Maisonneuve (lectotype, designated herein of *Sphaeria jucunda*, PC, filed as *Hypocrea jucunda*, two isotypes at PC filed as *Sphaeria jucunda*). UNITED STATES. Alabama: Tuskegee, on *Opuntia ficus-indica*, Carver 584 (holotype of *Nectriella cacti* NY). Additional specimen examined.— SPAIN. Los Retacos, Almeria, on *Opuntia*, June 1997, J. Checa, det. A. Rossman (BPI 744973).

Notes.— Pink pycnidia of Leptodermella opuntiae were present on the type of Nectriella cacti but the anamorph association with N. jucunda is not proven.

Nectriella luteola (Desm.) Weese, Ann. Mycol. 12: 131, 1914.

■ Sphaeria luteola Desm., Pl. Crypt France, Ed. 1, Sér. 1, Fasc. 42: 2078. 1850.

≡ Calonectria luteola (Desm.) Sacc., Michelia 1: 315.
1878.

≡ Charonectria luteola (Desm.) Höhn., Sitzungsber, Kaiserl. Akad. Wiss.. Math.-Naturwiss. Kl., Abt. 1, 115: 1193, 1906.

Ascomata immersed-erumpent, scattered, on veins or petioles of leaves, subglobose, 300 µm diam, orange-

brown, translucent, KOH–, collapsing vertically. Ascomatal wall 20–30  $\mu m$  thick, of two regions: outer region 10–15  $\mu m$  thick, of thick-walled, angular to rounded cells, 6  $\mu m$  diam; inner region of thin-walled, elongate cells. Asci clavate, 44–60  $\times$  8–12  $\mu m$ ; apex truncate, with a ring; ascospores biseriate. Ascospores naviculate–ellipsoid, 11–16  $\times$  3–5(–6)  $\mu m$ , 1-septate, often slightly constricted, slightly roughened, several small guttules per cell.

ANAMORPH. - None known.

Habitat.— In leaf veins and petioles of deciduous trees. Distribution.— Europe.

LECTOTYPE, DESIGNATED HEREIN.— FRANCE. In leaves and petioles of *Populus* and *Fraxinus*, summer, no collector, Pl. Crypt. France 2078 (PC; isolectotypes, BPI; FH – Höhnel; H; K; NY).

ADDITIONAL SPECIMENS EXAMINED.— LUXEMBOURG. On petiole of Fraxinus, Apr 1902, J. Feltgen (FH-A2899; as Charonectria luteola). SWITZERLAND. Valais: Aletschwaldreservat, Reidialz, on fallen leaves of Alnus viridis, 9 Sep 1970, R.W. Dennis (K; as Nectria arenula;). UNITED KINGDOM. England: Devon, Slapton Ley Nature Reserve, on stem of Rubus, 10 Oct 1979, M.C. Clark (IMI 24778; K).

Notes.— Nectriella luteola is similar to N. bloxamii, separated by host and ascomatal color.

#### Nectriella minuta Lowen, sp. nov. — Plate 2, f.

Ascomata obpyriformia,  $160\text{--}200 \times 130\text{--}175~\mu\text{m}$ , immersa, gregaria; apex luteus; setae nullae; parietes  $14\text{--}20~\mu\text{m}$  crassi, bistratosi. Asci clavati,  $40\text{--}70 \times 6\text{--}8~\mu\text{m}$ , annulo praediti. Ascosporae biseriatae, ellipsoideae,  $10\text{--}12 \times 2\text{--}3~\mu\text{m}$ , 1-, demum 3-septatae, hyalinae, laeves, guttulatae. Anamorphosis: Acremonium sp.

Ascomata immersed with only the papilla emerging, or appearing superficial if epidermis erodes, then seated on the mycelium in dense groups. Ascomata obpyriform, smooth,  $160-200~\mu m$  high  $\times$   $130-175~\mu m$  diam, nearly white, KOH-, apex yellow, papilla truncate to acute,  $40-50~\mu m$  diam, smooth, collapsing by lateral pinching. Ascomatal wall  $14(-20)~\mu m$  thick, of two regions: outer region of thick-walled, angular to rounded cells.  $ca~4~\times~4~\mu m$  diam; inner region of thin-walled cells. Asci clavate,  $40-70~\times~6-8(-14)~\mu m$ ; with an apical ring, ascospores biseriate. Ascospores narrowly ellipsoid, one end occasionally tapered,  $(9-)10-12(-14)~\times~2-3(-4)~\mu m$ , 1-septate, becoming 3-septate, slightly constricted, hyaline, smooth to spinulose.

CHARACTERISTICS IN CULTURE.— Conidiophores arising from aerial hyphae and from agar surface, solitary; terminating in a single phialide; phialides (10–)15–38  $\times$  1.5–3  $\mu$ m at the base, narrowing to 1–1.5  $\mu$ m at the apex. Conidia oblong to ellipsoid, 3–5(–12)  $\times$  1–1.5(–2)  $\mu$ m, unicellular, smooth, hyaline; basal ab-

scission scar protuberant and flattened on ellipsoid conidia, not seen on oblong conidia; conidia held in hyaline liquid.

Habitat and distribution.— Known only from the type collection.

Type.— VENEZUELA. Edo. Mérida, La Montaña, El Teleférico above Mérida, on dead culms of bamboo, 30 July 1971, K.P. Dumont VE 3435 & G.J. Samuels, culture C.T.R. 71-347 (NY, holotype).

ETYMOLOGY. - Referring to the small, inconspicuous ascomata.

Notes.— Nectriella minuta is distinguished from other species of Nectriella by the tropical distribution, occurrence on bamboo, nearly white ascomata, and Acremonium-like anamorph. The species was grown in culture from single ascospores.

Nectriella paludosa Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 176. 1869 [1870].

[≡ Nectria paludosa (Fuckel) Sacc., Michelia 1: 289, 1898, non Nectria paludosa H. Crouan & P. Crouan 1876]. [= Nectriella diaphana Fuckel & Nitschke, in Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 176, 1869 [1870], name invalid, Art. 34.1, not accepted by author]

Ascomata often remaining immersed, sometimes erumpent, scattered or in groups of up to 5, often only the papilla showing through the epidermis, obpyriform, 190–290  $\mu$ m high × 230–430  $\mu$ m diam, at first pale pink, then yellow, KOH–, with truncate papilla, collapsing vertically or not collapsing. Setae on papilla, clavate, 8 × 2  $\mu$ m. 2-septate, sparse. Ascomatal wall 20–24  $\mu$ m thick, of two regions: outer region of thickwalled, angular to rounded cells, surface cells ca 8  $\mu$ m diam; inner region of thin-walled, rectangular cells. Asci clavate,  $68-85 \times 7.5$ –10  $\mu$ m, apex truncate, with a ring; ascospores biseriate. Ascospores ellipsoid,  $(12-)14-23 \times 4-5 \mu$ m, 1-septate, often slightly constricted, hyaline, pale pink in mass, echinulate, with 2 guttules per cell.

ANAMORPH.— None known.

Habitat.— In stems of *Typha* and *Iris*. Distribution.— Europe, United States.

Plate 8. a. Nectriella rubricapitula, median section of ascoma and ascus, b. Nectriella utahensis, median section of ascoma, ascus, ascospores, and conidiophore. c. Pronectria robergei, ascomatal wall cells, ascal apex, and ascospores. d. Pronectria echinulata, ascomatal wall cells, ascus, and ascospores. e. Pronectria pertusariicola, median section of ascoma, asci and ascospores. a. Isotype – NY. b. Holotype – NY. c. Lectotype of Cryptodiscus lichenicola – BPI, d. Holotype – IMI, e. Holotype – UPS. Scale bars: a. for ascoma = 50 μm, for ascus = 10 μm, upper b, upper e = 100 μm; lower b, c, d, and lower e = 10 μm.

HOLOTYPE.— GERMANY. Hessen: Near Budenheim, on rotting leaves of *Typha angustifolia*, floating in swamp water, spring, Fuckel & Nitschke, Fungi rhenani 2048, Fasc. 6. 1867 (G; isotypes, Herb. Barb. Boiss., as *N. diaphana* FH, IMI, K).

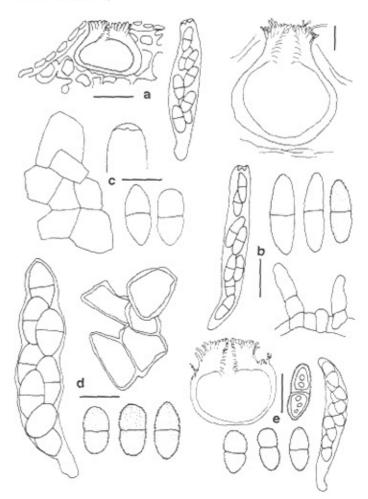
ADDITIONAL SPECIMENS EXAMINED.— CZECH REPUBLIC. Moravia: Mähr.-Weißkirchen, Thein, on Typha latifolia, Sep 1927, F. Petrak (C, IMI); Lipnik, Thrin, on T. latifolia (IMI 9446). GERMANY. On Iris pseudacorus, Aug 1904, Krieger, Fungi Sax. 1769 (FH, K); on T. latifolia, Petrak 456 (K); Petrak 2488 (K); (IMI 202253; as N. dacrymycella). UNITED KINGDOM. England: West Sussex. Loder Valley Nature Reserve, on T. latifolia, 26 Oct 1986, R. Lowen 232 (IMI 348647). UNITED STATES. Maine: Sagadahoc County, Merrymeeting Bay, ca 6 mi N of Woolwich, highway 128, across road from R.P.T. Coffin Wild flower Sanctuary, 44°01′ N, 69°48′ W, on damp, dead stems of T. latifolia, 19 Sep 1987, C.T. Rogerson & R. Lowen 356–87 (NY).

Notes.— The papilla of *Nectriella paludosa* sometimes appears dark due to the presence of algae on the surface of the substratum. In the cultures produced from single ascospores no anamorph formed. Ascomata developed from single ascospores, thus the species is homothallic.

Nectriella pironii Alfieri & Samuels, Mycologia 71: 1181. 1979 [1980].

Anamorph: Kutilakesa pironii Alfieri, Mycotaxon 10: 217. 1979.

This species was described and illustrated by Alfieri & Samuels (1979).



## Nectriella rubricapitula Lowen, sp. nov. - Plate 8, a.

Ascomata obpyriformia,  $80\times100~\mu\text{m}$ , immersa, solitaria vel gregaria; apex ruber; setae nullae; parietes 14  $\mu\text{m}$  crassi, e strato unico compositi. Asci clavati,  $35\text{--}45\times4.5\text{--}5.5~\mu\text{m}$ , annulo praediti. Ascosporae biseriatae, ellipsoideae,  $6\text{--}7.5\times2~\mu\text{m}$ , 1-septatae, hyalinae, laeves, guttulatae. Anamorphosis ignota.

Ascomata obpyriform, 80  $\mu$ m high  $\times$  100  $\mu$ m diam, immersed, scattered or in groups of up to 10, strongly adherent to the substratum, red at ascomatal apex, immersed part of the wall pale yellow, KOH-; papilla truncate, 50  $\mu$ m diam, smooth. Ascomatal wall 14  $\mu$ m thick, of one region, thick-walled, elongate. Asci clavate, (29–)35–45  $\times$  4.5–5.5(–6.5)  $\mu$ m; apex truncate, with a ring, ascospores biseriate. Ascospores ellipsoid, 6–7.5  $\times$  2  $\mu$ m, 1-septate, often slightly constricted, hyaline, smooth, inconspicuously guttulate. Anamorph unknown. Habitat and distribution.— Known only from the type collection.

HOLOTYPE.— BRAZIL. Amazonas: Plateau of Serra Araca, N side of N Mountain, 1250 m, 00° 57′ N, 63° 21′ W, cloud forest, on twig, 17–22 Feb 1984, G.J. Samuels 481a, G.T. Prance & J. Pipoly (INPA; NY, isotype).

Etymology.— Referring to the red apex of the ascomata.

Notes.— Nectriella rubricapitula is distinguished from other species of Nectriella by the adherence of the ascomata to the woody substratum and the red ascomatal apex.

Nectriella sambuci (Höhn.) Weese, Ann. Mycol. 12: 150. 1914.

≡ Charonectria sambuci Höhn., Hedwigia 42: 187, 1903.

Ascomata immersed, sometimes erumpent, scattered or in groups of up to 30, some contiguous, obpyriform, 100-300 μm high × 220 μm diam, at first reddish yellow, then yellow, KOH-; papilla truncate, 60-140 µm high × 120-160 μm diam; sometimes collapsing vertically, setae clavate,  $40 \times 4 \mu m$  (not seen in type). Cells on surface of ascomata angular, 5-10 µm diam. Ascomatal wall 20-30 µm thick, of two regions: outer region of rectangular to ellipsoid cells,  $6-10 \times 2-3 \mu m$ , with 2 µm thick walls; cells of the inner region having 1 μm thick walls. Asci clavate when young, mature asci in type not seen,  $50-68 \times 6-8 \mu m$ , apex truncate, with a ring, deliquescing at maturity; ascospores biseriate. Ascospores ellipsoid, 14-16 × 3-4(-6) µm, typically 1-septate, occasionally 3-septate, often slightly constricted, many curved, at first hyaline, then slightly brown, smooth, often several guttules per cell.

Anamorph.— None known.

Habitat.— On stems of woody plants.

Distribution.— Europe.

HOLOTYPE.— YUGOSLAVIA. Herzegovina, Jablanica, on thin, dry stems of Sambucus nigra, Apr 1903, Höhnel (FH). ADDITIONAL SPECIMENS EXAMINED.— UNITED KINGDOM. England: N. Yorkshire, Levisham, Hagg Wood Marsh NR, BW, C-73-4, on Filipendula ulmaria [Rosaceae]. 1 July 1973, collector unknown (IMI 177564).

Notes.— Nectriella sambuci has papillate ascomata with scattered hairs and a wall with two regions. Like the related species, N. fuckelii and N. paludosa, the ascomata have scattered secretory setae and do not usually collapse when dry.

Nectriella silenes-acaulis Nograsek, Biblioth. Mycol. 133: 69. 1990.

This species was described and illustrated by Nograsek (1990).

Nectriella utahensis Lowen & Rogerson, sp. nov. — Plate 7, g; Plate 9, b.

Ascomata obpyriformia,  $238-500 \times 255-400 \, \mu m$ , immersa, solitaria vel gregaria, luteo-aurantiaca; setae cylindricae,  $7-30 \times 3.5-5 \, \mu m$ , parum septatae; parietes  $18-35 \, \mu m$  crassi, bistratosi. Asci clavati,  $40-60 \times 5.5-12 \, \mu m$ , annulo praediti. Ascosporae biseriatae, ellipsoideo-fusiformes,  $14-20 \times 4-6.5 \, \mu m$ , 1-septatae, hyalinae, demum leviter coloratae, spinulosae, guttulis compluribus. Anamorphosis ignota.

Ascomata obpyriform, 238-500 µm high × 255-400 µm diam, immersed to erumpent, scattered or in groups up to 10, yellow-orange, KOH-; papilla truncate, 62-100 µm high × 123-176 µm diam; collapsing by lateral or vertical pinching of ascomatal base. Setae around the ostiole, 7-30 × 3.5-5 µm, tapering to the rounded apex, septate, walls slightly thickened. Ascomatal wall 18-35 µm thick, of two regions: outer region 7-18  $\mu$ m thick, of ellipsoid cells, 3.5-7  $\times$ 1.5-4.5 µm; inner region 11-18 µm thick, of thinwalled cells  $7-15 \times 0.5-2$  µm. Asci clavate,  $40-60 \times$ 5.5-12 µm, apex truncate, with a ring, ring often pushed to one side in mature asci, ascospores biseriate. Ascospores ellipsoid-fusiform, 14-20 × 4-6.5 μm, 1septate, pale orange in mass, spinulose when mature, with several orange guttules per cell. Anamorph unknown.

Habitat.— On dead leaves of Swertia radiata (Gentianaceae) and possibly larkspur.

DISTRIBUTION.— United States (Colorado, Utah).

HOLOTYPE.— UNITED STATES. Utah: Weber County: north of N. Ogden Divide, Wasatch Crest trail, on decaying leaves of *Swertia radiata*, 19 Aug 1987, C.T. Rogerson (NY). ADDITIONAL SPECIMEN EXAMINED.— UNITED STATES. Colorado: Mesa County, Grand Mesa Nat. Forest, S.E. of Mesa Lake, on stems of larkspur?, 14 July 1930, R.W. Davidson 646 (BPI, as *Nectriella pedicularis*).

ETYMOLOGY.— Named for the location of the type collection, a favorite locale of the collector.

Nectriella verrucosa Urries, An. Jard. Bot. Madrid 1: 67, 1941.

Ascomata immersed, scattered or in groups of up to 20, obpyriform, 313–387  $\mu$ m high × 176–317  $\mu$ m diam, at first reddish, then yellow, KOH-; papilla truncate, 282–310  $\mu$ m high × 132–308  $\mu$ m diam, setae broken, the largest ones  $66 \times 5 \mu$ m, base tapered to 2  $\mu$ m, hyaline, septate, with ca 1  $\mu$ m thick wall. Ascomatal wall 20–35  $\mu$ m thick, of two regions: outer region 12–26  $\mu$ m thick, of thick-walled cells; inner region 7–9  $\mu$ m thick, of thin-walled, parallel, densely packed cells. Asci clavate,  $(70-)80-135 \times 10-20 \mu$ m; apex angular; simple; ascospores biseriate in the middle; base clavate, lower quarter occasionally empty. Ascospores narrowly

ellipsoid, one side often curved, the other side straight,  $(20.5-)22-28(-30) \times (5-)6-8(-9) \mu m$ , 1-septate, septum inconspicuous in mature ascospores, hyaline, verrucose at maturity, often many guttules per cell, tending to disappear at maturity.

Habitat.— On rotting paper.

DISTRIBUTION.— Known only from the type collection

HOLOTYPE.— SPAIN. Near Madrid, on rotting paper, 14 Jan 1940, Urries (MA).

Notes.— Nectriella verrucosa is related to N. funicola but distinguished by the larger, more ornamented ascospores, more elongate, thin ascomatal wall cells and an angular apex of the ascus. The two taxa are represented by only a few collections.

## KEY TO THE SPECIES OF NECTRIELLA

1.	Smooth-walled
2 (1). 2.	Ascospores generally less than 10 $\mu m$ long, non- to 1-septate, smooth-walled 3 Ascospores generally more than 10 $\mu m$ long, 1-septate, smooth or ornamented 6
3 (2). 3.	Ascospores non-septate, cylindrical to allantoid, $5-6 \times 1.5 \mu m$ , hyaline; ascomata pale yellow to orange; on <i>Opuntia</i>
4 (3). 4.	Ascomata yellow with red apical region; ascospores ellipsoid, 6–7.5 × 2 μm; on woody twigs
5 (4). 5.	Ascomata pale pink, non-setose; ascospores ellipsoid, $8-10\times2-3~\mu m$ , on decaying leaves of Ammophila
6 (2). 6.	Ascomata with long, straight setae, or flexuous hyphae or hairs, often in a circle around the ostiole, globose to obpyriform, generally papillate
7 (6). 7.	mata pale yellow to yellow, with clavate, thick-walled hairs up to 20 µm long surrounding the ostiole; ascospores ellipsoid–fusiform, 12.5–26 × 2.5–4.5 µm, finely striate; anamorph <i>Kutilakesa</i>
	where known, Acremonium-like

In empty ascomata of Balansia: ascospores ellipsoid-fusiform, 9.5–10.5 × 2.5–3 µm.

8 (7).	On decorticated wood of <i>Populus</i> ; ascospores narrowly ellipsoid, 16-19 × 4-7 µm
8.	Spinulose
9 (8). 9.	On rotting paper and rope
10 (9). 10.	Ascospores ellipsoid, $16-24\times4-8~\mu\text{m}$ , spinulose to verrucose; on rotting paper and rope, known throughout Europe
11 (9). 11.	On decaying leaves including petioles
12 (11).	On decaying leaves of <i>Swertia</i> in western United States; ascomata yellow-orange with cylindrical, straight setae; ascospores ellipsoid–fusiform, 14–17.5 × 4–6.5 µm, spinulose
12.	On petioles of Gunnera in Chile; ascomata bright orange with cylindrical, recurved setae; ascospores ellipsoid–fusiform, 17–20 × 3.5–5.5 µm, minutely spinulose
13 (11). 13.	On Iris and Typha; ascospores ellipsoid, 14–23 × 4–5 µm, spinulose N. paludosa On dicotyledonous herbaceous stems; ascospores smooth, spinulose, verrucose or finely striate
14 (13).	On Silene; ascospores finely striate, ellipsoid-fusiform, 12.5-16 × 2.5-4.5 µm
14.	On herbaceous stems other than Silene; ascospores smooth or verrucose
15 (14). 15.	Ascomata subglobose, pale pink; ascospores $1012 \times 46~\mu\text{m}$ , ellipsoid, smooth to slightly roughened; on $Galium$
	Ascospores ellipsoid, $14-16 \times 3-4 \mu m$ , smooth-walled
17 (16).	Ascospores ellipsoid, 13–18 × 3.5–5 μm, spinulose; on Arabis or Saxifraga
17.	Ascospores ellipsoid–fusiform, 15–20 × 4–5.5 μm, verrucose; on Apiaceae, Asteraceae or unknown herbaceous stems
18 (6).	Ascomata yellow with a red apical region, subglobose to globose; ascospores ellipsoid to fusiform, 12 × 3 μm, smooth-walled; on <i>Rubus</i>
18.	Ascomata concolorous
19 (18). 19.	On decaying monocotyledonous plant parts, either on <i>Typha</i> and <i>Poaceae</i> ; ascomata pale yellow
20 (19).	On dead leaves of <i>Typha</i> or grasses; ascomata subglobose; ascospores ellipsoid,

- 21 (19). Ascospores 11–16 × 3–5(–6) μm, ellipsoid–naviculate, spinulose; ascomata orange-brown; on decaying leaves including petioles of deciduous trees and stems of Rubus N. Interela
- Ascospores averaging longer than 16 μm, ellipsoid to ellipsoid–fusiform, smooth to spinulose; ascomata bright orange to orange-brown or tan; on various herbaceous stems

NECTRIOPSIS Maire, Ann. Mycol. 9: 323. 1911, nom. cons. prop.

Lectotype, designated by Weese (1913): N. violacea (Fr.) Maire (≡ Sphaeria violacea Fr.).

- = Dasyphihora Clem., Gen. Fungi p. 45, 1909, nom. rej. prop. Type: D. lasioderma (Ellis) Clem. (≡ Nectria lasioderma Ellis), recognized as Nectriopsis lasioderma (Ellis) Samuels.
- = Peloronectriella Doi, Bull. Natl. Sci. Mus. Tokyo 11: 179. 1968. — Type: P. sasae Doi, recognized as Nectriopsis sasae (Doi) Rossman & Samuels.

Ascomata superficial or immersed in substratum, generally not conspicuously stromatic, generally less than 200 µm diam, nearly white to pale yellow or orange, rarely violet or purple, KOH—. Ascomatal wall less than 20 µm thick, usually of a single region of small, thin-walled, non-descript cells; wall cells at surface forming a textura epidermoidea. Anamorph, where known, Acremonium, Gliocladium-like, or Verticillium-like. On free-living fungi, lichens, and myxomycetes, less frequently on herbaceous substrata.

Notes. - Nectriopsis was established with four species of hypocrealean fungi having ascomata in a byssoid stroma and considered intermediate between Nectria and Hypomyces. Samuels (1988) presented a thorough account of the genus including 43 species each of which was described and illustrated. In the present work, the species that occur on Meliola have been removed to the genus Dimerosporiella. Thus, 39 species, including two additional species described below, are recognized in Nectriopsis. Clements (1909) placed Dasyphthora in the Hypocreaceae with only one species, D. lasioderma, that was included in Nectriopsis (Samuels, 1988). Although Dasyphthora provides an earlier name, Nectriopsis has been proposed for conservation (Rossman & Samuels, 1998). The unispecific genus Peloronectriella was described for a species on bamboo having an elongate, tuberculate stroma with Nectria-like ascomata and 1-septate ascospores. The type specimen of Peloronectriella sasae was examined and found to be a Nectriopsis growing on the surface of overmature stromata of Shiraia bambusicola Henn. Thus Peloronectriella sasae belongs in the genus Nectriopsis and Peloronectriella is a synonym of Nectriopsis.

Nectriopsis violacea (Fr.) Maire, Ann. Mycol. 9: 323. 1911.

- ≡ Sphaeria violacea Fr., Summa Veg. Scand. 2(2): 441.
  1823.
- ≡ Nectria violacea (Fr.) Fr., Summa Veg. Scand. 2: 388.
  1849.
- ≡ Hypomyces violaceus (Fr.) Tul., Ann. Sci. Nat. Bot. ser.
  4, 13: 14. 1860.
- ≡ Peckiella violacea (Fr.) Sacc., Syll. Fung. 9: 945, 1899,
   ≡ Hypolyssus violaceus (Fr.) O. Kuntze, Revis. Gen. Plant.
  3 (2): 488, 1898.
- ≡ Byssonectria violacea (Fr.) Seaver, Mycologia 2: 65.
  1910.
- ≡ Hyphonectria violacea (Fr.) Petch, J. Bot. 75: 222.
  1937.

Anamorph.— Acremonium fungicola (Sacc.) Samuels, Mycologia 65: 404, 1973

≡ Diplosporium album var. fungicola Sacc., Syll. Fung. 4:
178. 1886.

Mycelium white, becoming violet immediately surrounding each perithecium, dense, covering the surface of the host aethalia. Ascomata immersed in mycelium, becoming collabent when dry, broadly pyriform, (116–)240–275(–390) μm high × (150–)240–260 (–310) μm diam, or globose, (170–)240–260(–340) μm diam, violet to purple; surface cells thin-walled, angular, 7–10 μm diam; papilla acute, of thick-walled, septate, unbranched hyphae; hyphae extending outwardly as hairs, 10–50 μm long, 5 μm wide at the rounded apices, forming a fringe around the papilla; periphyses ca 15 μm long, 2 μm wide at the base, rounded apices

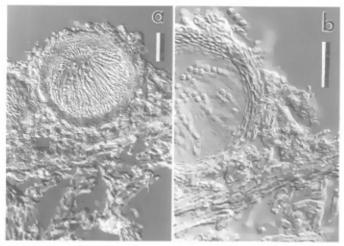


Plate 9. a-b. Nectriopsis queletii. a. Median section of ascoma on natural substratum. b. Close-up of ascomatal wall. a-b. Holotype – H. Scale bars = 25 μm.

1  $\mu m$  wide. Ascomatal walls 15–20  $\mu m$  thick. Asci cylindrical, (40–)50–60(–75)  $\times$  3–5  $\mu m$ , 8-spored, sessile, with an apical ring, ascospores obliquely uniseriate with overlapping ends. Ascospores cylindrical, (5–)7–8  $\times$  2.5–3  $\mu m$ , equally 2-celled, not constricted, hyaline, spinulose.

Anamorph.— Conidiophores arising from both surface and aerial mycelium, white, unbranched phialides or 2–3 phialides arising from tip of one axis. Phialides aseptate or uniseptate, smooth, 30–50  $\mu$ m long, from 2  $\mu$ m at the base tapering to 1  $\mu$ m at the tip. Conidia unicellular, smooth-walled, hyaline, ellipsoid, 6–9.5(–17) × 2–3  $\mu$ m, in solitary, slimy, hyaline heads at apices of phialides.

Habitat.— On the myxomycete Fuligo septica (L.) Wiggers.

DISTRIBUTION.— Known throughout temperate North America and Europe.

HOLOTYPE.— GERMANY. Bernstadt, on Fuligo violacea, 1817 (UPS; herb. E. Fries, as Sphaeria violacea).

ILLUSTRATIONS.— Müller & von Arx (1962, Fig. 250); Munk (1957, Fig. 8, as *Nectria violacea*); Plowright (1882, Pl. 157, Fig. 2 a-e, as *N. violacea*); Samuels (1973b, Figs 1, 2, 7–11, as *N. violacea*); Schmid & Schmid (1990; Fig. 32).

Notes.— Samuels (1971) studied the ontogeny of ascomatal development in *Nectriopsis violacea* and *N. candicans* (Plowr.) Maire, a similar myxomyceticolous species, and demonstrated that both had a *Nectria*-type of centrum development.

Nectriopsis sasae (Doi) Rossman & Samuels, comb.

≡ Peloronectriella sasae Doi, Bull. Natl. Sci. Mus. Tokyo
11: 179, 1968.

Ascomata basally to almost totally immersed in a stroma covering overmature stromata of the Shiraia host: Nectriopsis stroma readily differentiated from that of Shiraia, evident when sectioned; stroma of Nectriopsis 100-500 µm thick, prosenchymatous to pseudoparenchymatous, with hyphal hairs on the surface, in section often of two regions: lower region 0-270 µm thick, of thin-walled cells 2-6 µm diam, forming a prosenchyma; upper region 150-380 µm thick, of thinwalled cells forming a textura prismatica, cells 8-14 μm × 2.5 μm. Hairs on stromatal surface and upper portions of ascomata, 12-40 × 3-6 µm, thin-walled, septate, flexuous, apex rounded. Ascomata globose to broadly pyriform, 200-250 µm high × 175-200 µm diam, collapsing when dry or not, ochraceous to umber, becoming pale ochraceous when dry, KOH-, ascomatal wall of one 20-25 µm thick region, cells thin-walled, 6-10 × 3-4 μm, forming a textura prismatica, toward the apex becoming textura angularis, cells 4-6 µm diam, with walls slightly thickened up to 1.5 µm. Ostiole lined with periphyses. Apical paraphyses seen in immature ascomata, of thin-walled, inflated cells. Asci cylindrical with truncate apex, 77-87 × 5.5-6 µm, apical ring barely visible, ascospores uniseriate. Ascospores ellipsoid, 8.5-10 × 3-4.5 μm, 1-septate, hyaline, spinulose.

HABITAT.— Parasitic on stromata of *Shiraia bambusicola*. Distribution.— Japan, known only from the type specimen.

HOLOTYPE.— JAPAN. Iwate Pref.: Mt. Hayachine, Ohazamacho, 18 July 1967, Y. Doi, D-304 (TNS-F-199660). Culture CBS 333.69.

ILLUSTRATION. - Doi (1968, I.c.), as Peloronectriella sasae.

Notes.—All ascomata on the type specimen were slightly immature, thus ascospores were measured inside the ascus. *Nectriopsis sasae* is placed among the mostly fungicolous species of *Nectriopsis* and appears to be similar to those species of *Nectriopsis* that occur on large clavicipitaceous stromata on grasses, namely *N. epichloë* and *N. macroepichloë*. The anamorph described for *N. sasae* is similar to the *Acremonium*-like anamorphs known for other species of *Nectriopsis*.

One species is described below in addition to those included above and in Samuels (1988).

Nectriopsis queletii (P. Karst.) Samuels, comb. nov. — Plate 9, a-b.

≡ Hyponectria queletii P. Karst., Hedwigia 21: 34. 1882.
≡ Nectriella queletii (P. Karst.) P. Karst., Acta Soc. Fauna Fl. Fenn. 2: 15. 1885.

Anamorph: Acremonium sp.

Ascomata superficial, effused stroma of *Nectriopsis* intermixed with host hyphae, densely gregarious to caespitose with a sparse basal fringe, subglobose, 60–78 × 75–82 μm, yellow to orange, KOH–, becoming cupulate, smooth, apex not differentiated. Cells at surface angular to *textura epidermoidea*. Ascomatal wall 7–10 μm thick, of a single region of compressed cells, cells 3–7 μm diam, walls slightly thickened. Asci cylindrical, 53–75 × 6.5–9.1 μm, apex simple, sessile, 8-spored. Ascospores oblong to subglobose, 3–4.5 × 2–3 μm, 1-septate, hyaline, smooth.

Anamorph.— Conidiophores  $23-36 \times 2.5-4.5 \mu m$  at the base, apex not thickened, not flared, smooth-walled. No conidia seen on the type.

Habitat.— On hymenium of Phlebia albida.

DISTRIBUTION. - Finland and Sweden.

HOLOTYPE.— FINLAND. Near Mustiala, in hymenium of Stereum subcostatum P. Karst. [host identified as Phlebia albida], on fallen stems of Betula in shady places, 10 Oct 1881, ex Herb. Karsten 1367b (H).

ILLUSTRATION,— Eriksson et al. (1981, Fig. 553, as Nectriella queletii).

Notes.— The small, pallid, superficial ascomata and mycoparasitic habit place this species in *Nectriopsis* as defined by Samuels (1988), unlike the immersed ascomata on decaying herbaceous or lignicolous substrata characteristic of *Nectriella*. *Nectriopsis queletii* is similar to *N. oropensoides* in having very small ascospores and occurring on basidiomycetes in temperate regions; however, the ascospores of *N. queletii* are among the smallest in *Nectriopsis*, even smaller than those of *N. oropensoides*. Eriksson *et al.* (1981) noted that *N. queletii* occurs in both Finland and Sweden.

For a comprehensive account and a key to the remaining species of *Nectriopsis*, see Samuels (1988).

# OCHRONECTRIA Rossman & Samuels, gen. nov.

Type: Ochronectria calami (Henn. & E. Nyman) Rossman & Samuels (= Nectria calami Henn. & E. Nyman).

Ascomata superficialia, vulgo aggregata super stromate bene effecto, subglobosa vel globosa vel ellipsoidea, alba vel luteola, KOH-, parietes > 45 μm crassi, cellulae strati exterioris hyalinae, globosae; cellulae strati medii guttulis aurantiis oleaginosis interspersae. Asci 4–8-spori. Ascosporae fusiformes, pluriseptatae, hyalinae, laevigatae vel striatae.

Ascomata superficial, solitary to gregarious on a thin subiculum. Ascomata subglobose to globose, cupulate when dry, pale yellow to yellow-orange, KOH-, ascomatal surface smooth to slightly roughened, walls more than 45 µm thick, of three regions: outermost region of

hyaline, thin-walled, globose cells; middle region of angular to globose, thin-walled cells, with abundant, orange, oily droplets between the cells; inner region of hyaline, thin-walled, elongate cells. Asci narrowly clavate, 4–8-spored. Ascospores fusiform, multiseptate, hyaline, smooth to faintly striate. Anamorph *Acremonium*-like. On dead woody, often monocotyledonous, also dicotyledonous substrata.

Notes.— This unispecific genus is similar to Hydropisphaera recognized for members of the Nectria pezizagroup in which the relatively thick ascomatal wall is composed of large, thin-walled, inflated cells resulting in a cupulate collapse when dry. The characteristic wall structure consists of three regions with orange oil droplets in the middle region. Recent unpublished molecular studies of the Bionectriaceae suggest that the type species, O. calami, is distinct from species of Hydropisphaera. Ochronectria calami is relatively common in tropical regions.

Ochronectria calami (Henn. & E. Nyman) Rossman & Samuels, comb. nov. — Plate 4. b (see page 25).

- ≡ Calonectria calami Henn. & E. Nyman, in Warburg, Monsunia 1: 163, 1899.
- ≡ Nectria calami (Henn. & E. Nyman) Rossman, Myxotaxon 8: 494, 1979.
- = Calonectria blumenaviae Henn., Hedwigia 41: 6. 1902.
- = Calonectria oödes Petch, Ann. Roy. Bot. Gard. (Peradeniva) 7: 135, 1920.
- = Calonectria ignota Chardón, Scientific Survey of Porto Rico and Virgin Islands 8: 41. 1926.
- = Calonectria kampalensis Hansford, Proc. Linn. Soc. Lond. 153: 34. 1941.

Anamorph: Acremonium-like.

Ascomata solitary to gregarious, superficial on a thin subiculum of hyaline, thin-walled, 2-3 µm wide hyphae. Ascomata pale yellow to orange, becoming darker when dry, KOHN-, globose to subglobose, cupulate when dry,  $185-240 \mu m \text{ high} \times 175-260 \mu m \text{ diam, with}$ small, pointed papilla 10-20 µm high, ascomatal surface smooth to slightly roughened. Ascomatal wall 45-60 µm thick, of three regions: outer region of one layer of hyaline, globose, thin-walled cells, 7.5-10 μm diam: middle region 15-30 µm thick, widest near ascomatal apex, of angular to globose, thin-walled cells, 3-10 µm diam, with abundant, orange oily droplets formed between the cells: inner region 10-25 µm thick. thickest near the apex, of hyaline, thin-walled, elongate. 5-10 µm long cells. Asci unitunicate, 47-63 × 8-12 μm, narrowly clavate, without specialized apical discharge mechanism, 8-spored, ascospores obliquely uniseriate. Ascospores 24-38 × 4-5.5 µm, fusiform. sometimes curved or sigmoid, with narrowly rounded

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ends, (3-5-)7-9-septate, hyaline, smooth or faintly striate

Anamorph.— Conidiophores solitary, cylindrical,  $35-100~\mu m$  long,  $3.5-4~\mu m$  wide at the base, straight to slightly sinuous, thin-walled, smooth, developing from aerial fascicles or from the agar surface. Conidiogenous cells monophialidic, integrated, solitary, terminal, cylindrical,  $30-80\times3-3.5~\mu m$  wide at the base, tapering slightly, becoming  $2-2.5~\mu m$  wide at the apex, apex with flaring collarette up to  $2~\mu m$  long. Conidia broadly cylindrical, straight, (0-)1-3-(5-7)-septate, 0-septate  $8-13\times3.5-4~\mu m$ , 1-septate  $8-13\times3.5-4~\mu m$ , 2-septate  $11-13\times4-4.5~\mu m$ , 3-septate  $15-26\times4-5.5~\mu m$ , 5-septate  $22-25\times5-6~\mu m$ , 7-septate,  $21-36\times5-6~\mu m$ , hyaline, smooth. Hyphae hyaline, smooth,  $2.5-4~\mu m$  wide, chlamydospores lacking. Ascomata forming on PDA and V-8 after four weeks.

HABITAT.— On monocotyledonous wood and woody parts such as palm fruits, leaves and leaf sheaves, rarely also on tree ferns and dicotyledonous wood known from Calamus, Cocos, Heliconia, Hoya, Musa, Pipturus and Sabal.

DISTRIBUTION.— Pantropical, known from Bermuda, Brazil, French Guiana, Guadeloupe, Indonesia, Jamaica, Java, Panama, Peru, Puerto Rico, Sri Lanka, Uganda, United States (Hawaii), Venezuela (Rossman, 1983; Samuels et al., 1990).

Type.— JAVA. Hort. Bogor, on leaf sheaths of *Calamus* sp., E. Nyman, 4 Mar 1898, FH-general, lectotype, designated by Rossman, 1979b, isolectotypes FH – Höhnel, GZU. Cultures: CBS 125.87, 445.96, 454.96. Additional specimens examined listed in Rossman (1983) and Samuels *et al.* (1990).

ILLUSTRATIONS.— ROSSMAN (1983, Fig. 33, Pl. 11 C-F, as N. calami); Samuels et al. (1990, Fig. 23 D-F, as N. calami).

SPECIMEN ILLUSTRATED.— SRI LANKA (Ceylon). Peradeniya, on a decaying stem. Jun 1919, Petch 6009 (K – holotype of Calonectria oödes).

#### PARANECTRIA Sacc., Michelia 1: 317. 1878.

Type: P. affinis (Grev.) Sacc. (≡ Sphaeria affinis Grev.). = Ciliomyces Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt, 1, 115: 673. 1906. — Type: C.

oropensis (Ces.) Höhn. (≡ Nectria oropensis Ces.), recognized as Paranectria oropensis (Ces.) D. Hawksw. & Piroz.

Ascomata solitary, superficial on a white, thin, byssoid stroma or stroma lacking. Ascomata hyaline to pale orange or pale pink when fresh, KOH-, broadly pyriform to globose or subglobose, collapsing laterally or not at all when dry, smooth, scurfy or with short, septate hairs, wall relatively thin, less than 30 mm thick, of two regions. Asci cylindrical, 8-spored. Ascospores fusiform to ellipsoid with long, attenuated ends, multiseptate to muriform, hyaline, smooth. Anamorph unknown. On decaying lichens.

Notes.— The genus Paranectria was established for species with Nectria-like ascomata and long-attenuated, 3-septate ascospores, Within the Hypocreales, Paranectria is distinguished by the lichenicolous habit, white to pale vellow, often orange when fresh, KOH-, relatively thin-walled ascomata, and multiseptate to muriform ascospores with thin, attenuated ends. Paranectria belongs to the nectrioid Hypocreales affiliated with Ijuhva and Trichonectria based on similarities in ascomatal morphology and habitat. The type species, P. affinis, has been well-characterized (Rossman, 1983) and two additional species are included in Paranectria. Hawksworth & Pirozynski (1977) clarified the nomenclature of the generic names, Paranectria and Paranectriella. Ciliomyces was introduced by Von Höhnel for a Nectria-like species having muriform ascospores with attenuated ends. The type and only species, Ciliomyces oropensis, is found to be congeneric with Paranectria (Hawksworth & Pirozynski, 1977; Rossman, 1983).

Paranectria affinis (Grev.) Sacc., Michelia 1: 317, 1878.

≡ Sphaeria affinis Grev., Scott. Crypt. Flor. 4: 186. 1826.
 ≡ Nectria affinis (Grev.) Cooke, Grevillea 8: 9. 1879.

Anamorph: Unknown.

#### KEY TO THE SPECIES OF PARANECTRIA

- Ascospores broadly ellipsoid, 30–46 × 13–18 μm; asci 2- or 4-spored; on Peltigera rufescens
   P. superba

Ascomata solitary, superficial, loosely attached to the substratum by a sparse, white subiculum of hyphae, 5-6 µm wide. Ascomata white to pale yellow, KOH-, globose, cupulate when dry, ca 235  $\mu$ m high  $\times$  215  $\mu$ m diam, with a small, pointed papilla, ascomatal surface smooth, slightly roughened, or with loose strands of hyphae. Ascomatal wall 25-30 µm thick, of two intergrading regions: outer region 20-25 µm thick, of angular to elongate cells,  $8-13 \times 4-6 \mu m$ , with up to 1  $\mu m$ thick walls; inner region ca 5 µm thick, of hyaline, thin-walled, elongate cells. Asci clavate, 45-70 × 15-18 μm, simple, 8-spored, pluriseriate. Ascospores narrowly ellipsoid to fusiform, 24–34 (excluding ends)  $\times$  6–8  $\mu$ m, with long, thin, attenuated ends, 8–15  $\mu$ m long × 0.8 µm wide; ascospores 3-septate, hyaline, smooth-walled.

Habitat.— On thalli of lichens, Ephebe lanata and E. pubescens.

DISTRIBUTION. - Great Britain and France.

HOLOTYPE.— GREAT BRITAIN. Scotland: Appin, Carmichael (K. not examined; PC. possible isotype).

Specimen examined.— FRANCE. Fontainebleau, on Ephebe pubescens, 1893, De Notaris (RO).

ILLUSTRATIONS.— Dennis (1978, Pl. 31H); Greville (1826, Figs. 1 a-d, as *Sphaeria affinis*); Petch (1938, Fig. 21); Rossman (1983, Pl. 13E, Fig. 45).

Paranectria oropensis (Ces.) D. Hawksw. & Piroz., Canad. J. Bot. 55: 2555. 1977.

- ≡ Sphaeria oropensis Ces., in Rabenh., Bot. Zeitung 15: 406. 1857.
- ≡ Ciliomyces oropensis (Ces.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 115: 673. 1906.
- = Nectria lichenicola P. Crouan & H. Crouan, Fl. Finistère, p. 256, 1867.
- ≡ Pleonectria lichenicola (P. Crouan & H. Crouan) Sacc., Michelia 1: 325. 1879.
- = Pleonectria appendiculata Vouaux, Bull. Trimestriel Soc. Mycol. France 28: 193. 1912.

This species is described and illustrated in Samuels (1976a, as *Ciliomyces oropensis*) and Hawksworth (1982a). It is known from Austria, France, Ireland, Italy and Scotland on the lichens *Cladonia* sp., *Lecidea enteroleuca*, *Parmeliella atlantica*, and an undetermined leprose thallus.

Paranectria superba D. Hawksw., Notes Roy. Bot. Gard. Edinburgh 40: 390. 1982.

Hawksworth (1982a) described and illustrated this species that is known only from the type collection on thallus of *Peltigera rufescens* in Great Britain.

PEETHAMBARA Subram. & D.J. Bhat, Rev. Mycol. (Paris) 42: 49, 1978.

Type: P. sundara Subram. & D.J. Bhat.

Ascomata scattered, solitary to aggregated in small groups; superficial, on a thin, pseudoparenchymatous stroma. Ascomata bright- or dark yellow, globose to subglobose with a flattened apex, ostiolate. Ascomatal wall very thick, over 50 µm, of two regions: the outer region of very thick-walled, angular cells. Asci cylindrical, clavate, to broadly clavate, simple. Ascospores broadly reniform, 1- to 3-septate, hyaline. Anamorph synnematous, Didymostilbe. On dead woody substrata.

Notes.— The genus Peethambara was established for the teleomorph of Putagraivam sundaram, now Didymostilbe sundara. The type specimen of Peethambara sundara is apparently lost. The description included here is based on the original publication. Peethambara was described as having a Nectria-type centrum with a distinct ascomatal wall of two regions, one of which consists of extremely thick-walled, sclerenchyma-like cells. Seifert (1985) examined the type and additional specimens of the anamorph from Indonesia and Sierra Leone on woody hosts. Despite the lack of a type specimen, Peethambara is included in the Hypocreales based on the ascomatal wall characteristics and distinctive anamorph. Peethambara resembles members of Bionectria in having large, pale yellow to yellow, thickwalled ascomata, large, ascospores, and a synnematous anamorph. Preliminary molecular data suggest that Peethambara belongs in the Bionectriaceae allied with several anamorph genera having synnema and green, often multiseptate conidia (Rossman et al., 1998).

Peethambara spirostriata and P. sundara are similar in their thick-walled ascomata, broadly fusiform ascospores, and synnematous anamorphs producing multiseptate, greenish conidia. In addition, molecular data also suggest a close relationship between these species and the anamorph species, Albosynnema elegans E.F. Morris (Rossman et al., unpubl.).

Peethambara sundara Subram. & D.J. Bhat, Rev. Mycol. (Paris) 42: 49. 1978.

Anamorph: Didymostilbe sundara (Subram. & D.J. Bhat) Seifert, Stud. Mycol. 27: 140. 1985.

≡ Putagraivam sundarum Subram. & D.J. Bhat, Proc. Indian Acad. Sci., Sect. B, 87: 103, 1978.

Ascomata scattered, solitary to aggregated in small groups; superficial, with thin, pseudoparenchymatous stroma, stroma  $148-162 \times 33 \mu m$ , of golden-yellow hyphae. Ascomata golden-yellow, globose to subglo-

56 A. Y. Rossman et al.

bose with a flattened apex,  $380{\text -}440 \times 360{\text -}420~\mu\text{m}$ , smooth, ostiolate. Ascomatal wall  $60{\text -}70~\mu\text{m}$  thick, of two regions: outer region  $38{\text -}46~\mu\text{m}$  thick, of very thick-walled, angular cells,  $7{\text -}16.5 \times 5{\text -}6.5~\mu\text{m}$ ; inner region  $16{\text -}23~\mu\text{m}$  thick, of thin-walled, elongate cells. Periphyses cellular, cylindrical,  $15{\text -}20 \times 1.5{\text -}2.2~\mu\text{m}$ , ends rounded. Apical paraphyses evident in young ascomata, visible as remnants in mature ascomata. Asci cylindrical, clavate to broadly clavate,  $81{\text -}105 \times 21{\text -}28~\mu\text{m}$ , simple,  $4{\text -}8{\text -}\text{spored}$ , ascospores uniseriate above to biseriate below. Ascospores broadly reniform with rounded ends,  $31{\text -}42 \times 14.5{\text -}21~\mu\text{m}$ ,  $1{\text -}\text{septate}$ , hyaline, smooth.

Habitat and distribution.— Known only from the type collection.

HOLOTYPE.— INDIA. Karnataka State: South Kanara district, near Irde, at Darbhe, on dead twigs of *Macaranga indica* Wight. 22 Dec 1976. D.J. Bhat (MUBL 2358 – apparently lost; ex-type culture CBS 646.77). Culture CBS 521.96.

ILLUSTRATIONS.— Seifert (1985, Fig. 47; 1990, Fig. 5F, anamorph); Subramanian & Bhat (1978b, Figs. 1–22, anamorph; 1978c, Figs. 1–2, Pl. 1).

# Peethambara spirostriata (Rossman) Rossman. comb. nov.

≡ Nectria spirostriata Rossman, Mycol. Pap. 150: 61.
1983.

Anamorph: *Didymostilbe echinofibrosa* (E.F. Morris) Rossman, *comb. nov*,

■ Virgatospora echinofibrosa E.F. Morris, Mycologia 59: 538, 1967.

Ascomata superficial, solitary or in groups of up to five, without stroma, yellow to dark yellow, becoming darker when dry, KOH–, globose to subglobose, irregularly cupulate or not collapsing when dry, 305–470  $\mu$ m high  $\times$  360–575  $\mu$ m diam, without papilla, smooth. Ascomatal wall 50–70  $\mu$ m thick, of two regions: outer region 25–35  $\mu$ m thick, of angular to globose cells 10–18  $\mu$ m diam, with hyaline up to 1.5  $\mu$ m thick walls; inner region 25–35  $\mu$ m thick, of small angular to slightly elongate cells, 7.5–12  $\times$  5–7.5  $\mu$ m, with 2–3  $\mu$ m thick walls, the cells with only a small lumen; empty shell of ascomatal wall remaining when overmature. Asci clavate, 87–100  $\times$  17–23  $\mu$ m, simple, number of ascospores in

each ascus variable, often only 4–6, ascospores irregularly biseriate. Ascospores broadly fusiform with narrowly rounded ends, curved, 38–55 × 10–13 μm, 3–(4–5)-septate, with large guttules in each cell, hyaline, spirally striate, with 10–12 striae per half spore. Anamorph.— Synnemata scattered, solitary, 400–1500

μm tall  $\times$  40–70 μm wide at the apex, broadening to 150 μm at the base, stalk dark olivaceous-grey, paler toward the base, cells of stalk with dark walls, elongate, 12–25  $\times$  2–3 μm, head of synnemata globose, 125–200 μm diam, with conidia in a slimy, olivaceous-black mass. Conidiophores unbranched along most of their length, branching penicillately toward their apices. Conidiogenous cells phialidic, determinate, cylindrical to clavate, 10–30  $\times$  3–4 μm. Conidia broadly fusiform with papillate, truncate ends, straight or curved, 3-septate, 38–45  $\times$  10–15 μm, olivaceous-grey, coarsely striate.

Habitat.— On decaying woody substrata.

Distribution.— Tropical, known primarily from the Neotropics, also Gabon and Malaysia.

Types.— PANAMA: Prov. Panama. vicinity of Altos de Pacora, 26–31 km N of Pan American Hwy, on old road to Mandinga, elev. ca 700–730 m, on trunk of Cecropia sp., associated with Virgatospora echinofibrosa, K. P. Dumont et al., 30 June 1975, PA 1553 (holotype of Nectria spirostriata NY): Barro Colorado Island, Pierson Trail, on dead twigs, 3 Aug 1964, E.F. Morris & J.W. Strain 780, (lectotype of Virgatospora echinofibrosa, designated by Rossman, 1983; BPI 449174, isolectotype ILLS).

Additional specimens examined are cited in Rossman (1983).

ILLUSTRATIONS.— Ellis & Ellis (1971, Fig. 401, anamorph only); Rossman (1983, Fig. 32, Pl. 11 A, B).

PRONECTRIA Clem., in Clem. & Shear, Gen. Fungi p. 282. 1931.

Type: P. lichenicola (Ces.) Clem. (≡ Cryptodiscus lichenicola Ces. ≡ Nectria lichenicola (Ces.) Sacc.), a synonym of Pronectria robergei (Mont. & Desm.) Lowen.

Ascomata immersed in the host thallus, scattered or in groups, non-stromatic, subglobose to obpyriform, 100–500 µm diam, pale yellow to orange or red, rarely yellow, KOH– or rarely reacting. Setae rarely present. Cells on the ascomatal surface usually angular. Ascomatal wall 10–40 µm thick, generally of one region, also of two, rarely three, regions. Ascomatal apex of rows

## KEY TO THE SPECIES OF PEETHAMBARA

- Ascospores 1-septate, broadly reniform with rounded ends, 31–42 × 14.5–21 μm, smooth
   P. sundara

of parallel, vertically elongate cells, continuous with the inner region of the ascomatal wall. Asci clavate, usually less than 200 μm long or 15 μm wide, apex truncate, usually with a ring, 2–8-spored, ascospores biseriate in the middle, uniseriate above and below, or rarely uniseriate. Ascospores fusiform, ovoid or ellipsoid, typically not over 25 μm long and 8 μm wide, 1-septate, hyaline, smooth-walled, verruculose or spinulose. Anamorph, where known, Acremonium. On lichenized fungi and algae.

Notes.— Clements (in Clements & Shear, 1931) described and differentiated the genus Pronectria based on the lichenicolous habit, keying it out among hypocrealean fungi having one-septate, hyaline ascospores and non-stromatic, superficial ascomata. Although Pronectria was considered a synonym of Nectriella by Rogerson (1970), Lowen (1991) differentiated Pronectria from other hypocrealean genera by immersed ascomata, occurrence on lichens and algae, and a combination of morphological characters, i.e. ascomatal wall thin, often of one region, presence of ascomatal cells intermingled with those of the host, pale yellow to orange or dark red ascomata, generally KOH- except in P. fissuriprodiens, P. septemseptata, and P. subimperspicua. Another hypocrealean genus that includes lichenicolous species is Xenonectriella in the Nectriaceae. This genus has KOH+ ascomata with thickened wall cells, generally cylindrical asci, and verrucose to tuberculate ascospores that become yellowish brown with age. The species of Pronectria with KOH+ ascomata may belong in Xenonectriella; however, careful observation of ascomatal anatomy is needed to make this determination. Although Pronectria and Nectriella are similar in having KOH-, thin-walled ascomata immersed in the substratum. Nectriella differs in occurring on decaying herbaceous or woody plant material and having an ascomatal wall of two regions. Anamorphs of species of Pronectria have been placed in Acremonium, Diplosporium, and Illosporium. Lowen (1991) provided an account of the genus Pronectria with a key to the accepted species including two new species published here.

Pronectria robergei (Mont. & Desm.) Lowen, Mycotaxon 39: 462, 1990. — Plate 8, c.

■ Nectria robergei Mont. & Desm., Pl. Crypt. France, Ed.
 3, Fasc. 8: 374, 1856.

■ Nectriella robergei (Mont. & Desm.) Weese, in Höhn. & Weese, Ann. Mycol. 8: 467, 1910.

= Cryptodiscus lichenicola Ces., in Rabenh. Herb. Mycol. ed. 2, fasc. 6: 523. 1857.

≡ Nectria lichenicola (Ces.) Sacc., Michelia 1: 289, 1878.

≡ Calonectria lichenicola (Ces.) Rehm, Ascom, Lojk. p.
44, 1882.

 ■ Nectriella lichenicola (Ces.) Fuckel, in Höhn. & Weese, Ann. Mycol. 8: 466, 1910.

≡ Pronectria lichenicola (Ces.) Clem., in Clem. & Shear, Gen. Fungi p. 282. 1931.

= Nectriella carnea Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–23: 176. 1869 [1870].

= Nectria peltigerae W. Phillips & Plowr., Grevillea 4: 123.

Ascomata immersed, scattered or in groups of up to 20, closely adherent and raising the epidermis of the lichen or visible through star-like cracks in the epidermis, obpyriform, 240-340 µm high × 250-320 µm wide, pale red to orange, yellow when dry, KOH-; papilla truncate, 60-130 µm diam; ostiolar area sometimes depressed and hyaline. Ascomatal wall ca 35 µm thick, of two regions: outer region ca 17 µm thick, of thickwalled, globose to ellipsoid cells, 3.5 μm diam or 5 × 3.5 µm; inner region ca 18 µm thick, of thin-walled. elongated cells  $3.5-10.5 \times 2-3 \mu m$ ; periphyses directed downward into the centrum. Asci clavate, 40-70 × 8-9(-14) μm; apex with a ring, 8-spored, ascospores biseriate. Ascospores ovoid to ellipsoid, 8-16 × (3–)4–8 μm, 1-septate, sometimes slightly constricted, hyaline, smooth to slightly roughened.

Habitat.— On *Peltigera canina* and other species of *Peltigera*, often damaging the thallus of the lichen, discoloring areas delimited by a dark line from the inner layers, leaving smooth craters where ascomata have fallen out.

DISTRIBUTION.— Chile, Europe (Belgium, Finland, France, Germany, Italy, Luxembourg, Romania, Scotland, Spain – *fide* Martínez & Hafellner, 1998, Sweden), United States (Montana, New Hampshire, New York).

Type.— FRANCE. Normandy: Lebisey Park, on thallus of *Peltigera canina*, on old elm, Apr 1843, Roberge, Pl. Crypt. France, ed. 3, fasc. 8: 374. 1856 (PC, lectotype of *Nectria robergei*, designated herein: FH, H, IMI, K, PC, isolectotypes). ITALY. Piedmont: Oct.—Nov. 1856. Rabenhorst, Herb. Mycol. ed. 2, fasc. 6: 523. 1857 (BPI. lectotype of *Cryptodiscus lichenicola*, designated herein: IMI, K, S isolectotypes). GERMANY. Freienweinheim: Kiefernwald (pine wood). in thallus of *Peltigera canina*, spring, Kalchbrenner, Fungi Rhenani Exsiccati 1835 (G, holotype of *Nectriella carnea*, FH – Höhnel, IMI, K, S, isotypes). UNITED KINGDOM. Norfolk: Castle Rising [as 'Lynn'], Nov 1875, collector unknown (E, FH – Höhnel, isotypes of *Nectria peltigerae*).

Many additional specimens examined as cited in Lowen (1991).

ILLUSTRATIONS.— Müller & von Arx (1962, Fig. 247, as Nectriella robergei).

ADDITIONAL SPECIES OF PRONECTRIA:

Pronectria anisospora (Lowen) Lowen, Mycotaxon 39: 461, 1990.

≡ Nectriella anisospora Lowen, Mem. New York Bot. Gard 49: 248, 1989. This species was described and illustrated in Lowen (1989).

Pronectria casaresii Etayo, Nova Hedwigia 67: 504. 1998.

Recently described and illustrated from Spain by Etayo (1998), this is one of two species in *Pronectria* having more than one-septate ascospores.

Pronectria dealbans (Müll. Arg.) Etayo & Breuss, Cryptogamie, Bryol. Lichénol. 17: 220. 1996.

≡ Sphaerella dealbans Müll. Arg., Flora 55: 507, 1872.

This species was described and illustrated in Etayo & Breuss (1996).

### Pronectria echinulata Lowen, sp. nov. — Plate 9, d.

Ascomata obpyriformia,  $120-140\times100-130~\mu m$ , immersa, gregaria, aurantiaco-brunnea. Setae nullae. Parietes  $8-12~\mu m$  crassi, unistratosi. Asci clavati,  $52\times2~\mu m$ ; annulo deficientes. Ascosporae biseriatae, ellipsoideae vel ovoideae  $12-14\times5.5-8~\mu m$ , 1-septatae, hyalinae, echinulatae. Anamorphosis ignota.

Ascomata immersed in discolored, raised host thallus, in groups of up to 20, obpyriform, 120–140(–250) μm high × 100–130(–160) μm diam, orange-brown, becoming darker in KOH, not changing color in lactic acid; papilla truncate, non-setose. Surface cells angular, 5–9.5 × 8.5–12 μm. Ascomatal wall 8–12 μm thick, of one region of thin-walled angular cells, ca 7 × 5 μm. Asci clavate, 52 × 12 μm; simple, ascospores biseriate. Ascospores ellipsoid–ovoid, (11–)12–14(–18) × 5.5–8(–10) μm, 1-septate, sometimes slightly constricted, thin-walled, hyaline, spinulose. Anamorph unknown

Habitat.— On Physcia.

DISTRIBUTION.— Austria, Ireland, Spain (Etayo, 1998), U.S.A. (Idaho).

HOLOTYPE.— IRELAND. Gortnaskehy (H10), on *Physcia aipolia* on *Salix*, 30 Aug 1985, M.R.D. Seaward (IMI 105139).

ADDITIONAL SPECIMENS EXAMINED.— AUSTRIA: Steiermark: Hochschwab-Gruppe, Seetal W of Seewiesen, ca 10 km NE of Aflenz, 930 m, on *Physcia ascendens*, on *Fraxinus*, 19 Jan 1985, J. Hafellner 12580 & A. Ochsenhofer (Herb. J. Hafellner); UNITED STATES. Idaho: Lochsu River near Howell, on *Physcia* on stick, 29 June 1989, Katia Rodrigues (NY). Etymology.— Referring to the echinulate ascospores.

Notes.— Pronectria echinulata is distinguished from other species of Pronectria by its thin lateral ascomatal wall of brownish cells, spinulose ascospores with fragile walls that fracture easily with pressure, and ascomata aggregated in a raised, discolored area of the lichen thallus.

Pronectria erythrinella (Nyl.) Lowen, Mycotaxon 39: 461. 1990.

≡ Sphaeria erythrinella Nyl., Not. Sällsk. Fauna Fl. Fenn.
Förh. 4: 125. 1859.

≡ Nectria erythrinella (Nyl.) Tul. & C. Tul., Sel. Fung.
Carpol, 3: 95. 1865.

≡ Nectriella erythrinella (Nyl.) Höhn. & Weese, Ann. Mycol. 8: 466, 1910.

= Nectriella kalchbrenneri Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 177. 1869 [1870].

Anamorph: Illosporium sp.

Ascomata immersed, scattered or in groups of up to six, obpyriform, 280-320 µm high × 240-340 µm diam, at first red to orange, fading to yellow, KOH-; papilla truncate occasionally with hyphal ends free at edges, averaging 80 µm high × 160 µm diam, non-setose. Cells on surface angular to irregularly rectangular, mostly 10 × 5 μm. Ascomatal wall 20 μm thick, of one region, of thin-walled rectangular cells  $5-12 \times 2.5-3.5$ μm, widening to two regions in upper quarter; outer cells thick-walled, subglobose. Asci clavate, 72-90 × 10-14 µm; apex truncate, with a ring; ascospores biseriate. Ascospores ellipsoid-fusiform, (17–)18–20(–30) × (4-)5.5-6(-8) μm, 1-septate, septa often jagged, at first hyaline to pale yellow, then pale orange, verruculose, guttules often present in immature ascospores.

Anamorph.— Sporodochia erumpent, scattered or in groups on the lichen thallus, sometimes contiguous with ascomata, ca 250 μm diam, reddish orange. Conidia germinating from groups of cells, not from single cells.

Habitat.— On thalli of *Peltigera* spp.

Distribution.— In cool temperate regions.

Type specimens.— FINLAND. Nylandia: Helsinki (Helsingfors), Grid 27°E, on *Peltigera* sp., Nov 1858, W. Nylander (holotype of *N. erythrinella*. H; isotype, IMI 211135, slide, as *Sphaeria erythrinella*). CZECH REPUBLIC. Near Spis-Olaszi, in *Peltigera canina*, parasitic and ultimately destructive, June 1860, after heavy rain, Kalchbrenner, Rabenh. Fungi Eur. no. 73b (isotype of *N. kalchbrenneri*, NY, filed as *Illosporium carneum*).

SELECTED SPECIMENS EXAMINED.— CANADA. Alberta: SW of Calgary. Eau Claire camp ground, 1400 m (4600'), on Peltigera rufescens [= P. leptophora], on rock outcrops, lower subalpine region, 19 July 1981, R. Rosentreter 2198 (IMI 269698, Illosporium sp. also present). FINLAND. Pp: Ii: Iin aseman ratapihan N-pää sillanpiel en ratavallin W-rinne, grid 27°E, on Peltigera didactyla, 21 July 1964, J. Suominen (H, as Nectriella robergei, Illosporium sp. present); Myllyperä, on Peltigera, äng, May 1866, Karsten, Fungi Fenn. 475 (K, as S. erythrinella). RUSSIA. Bologoye: prov. Nangorva, 29 Aug, 10 Sep 1897, W. Tranzschel (S). SWEDEN. location unknown, 16 Aug 1974, R. Santesson (UPS, as N. robergei); UNITED STATES. Idaho: Lemhi County, Gilmore Summit,

Lemhi-Birch creek valley T13N, R27E, 2500 m, 20 June 1987, R. Rosentreter 4243, culture as R. Lowen 359a-87 (NY, Herb. Rosentreter); New Hampshire: Coos County, Shelburne, Sep 1891, W.G. Farlow 406 (FH, S, *Illosporium* sp. present).

Notes.— The ascomatal papilla of *Pronectria erythrinella* is barely visible through cracks in the thallus of the host, or it can become emergent surrounded by host tissue. The ascomatal apex is composed of parallel hyphae. Although similar to *P. robergei*, ascomata of *P. erythrinella* are brighter in color than those of *P. robergei* at first, but in the herbarium, where the colors usually fade, these species cannot be distinguished macroscopically. *Pronectria robergei* has smaller ascospores, shorter asci, and ascomatal walls of two regions unlike *P. erythrinella*. *Pronectria robergei* is usually found on *Peltigera* cf. *canina* whereas *P. erythrinella* often occurs on *Peltigera didactyla*.

Pronectria fissuriprodiens Etayo, in Etayo & Diederich, Bull. Soc. Nat. Luxemb. 97: 110. 1996.

This species was described and illustrated in Etayo & Diederich (1996).

Pronectria laminariae (O.E. Erikss.) Lowen, Mycotaxon 39: 461. 1990.

■ Nectriella laminariae O.E. Erikss., Svensk Bot, Tidskr.
58: 233, 1964.

This species was described and illustrated in Eriksson (1964).

Pronectria oligospora Lowen & Rogerson, Mycotaxon 53: 88. 1995.

This species was described and illustrated in Lowen (1995).

Pronectria oligospora var. octospora Etayo, Nova Hedwigia 67: 505. 1998.

This species was described by Etayo (l.c.).

Pronectria pertusariicola Lowen, sp. nov. — Plate 8, e.

Ascomata obpyriformia vel subglobosa, 220-300 × 150-270 μm, immersa, gregaria, aurantiaca. Setae nullae. Parietes

Plate 10, a. Pronectria subimperspicua, median section of ascoma, ascus and ascospores. b. Protocreopsis javanica, ascus and ascospores. c. Stilbocrea impressa, hairs on ascomatal wall, ascus, and ascospore. d. Valsonectria boldoae, asci and ascospores. e. Valsonectria pulchella, ascus and ascospores. a. Holotype – LPS. b. Holotype of P. palmicola – TNS. c. G.J.S. 84-256 – NY. d. Holotype – LPS. e. Holotype – LPS. Scale bars: left a = 100 μm; right a, b–e = 10 μm.

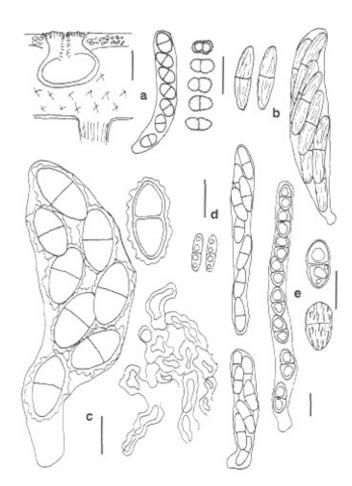
17–20 μm crassi, bistratosi. Asci clavati,  $60–80\times7–9$  μm; annulo indistincto. Ascosporae biseriatae, ovoideae,  $9–12\times4.5–5$  μm, 1-septatae, hyalinae, echinulatae, guttulatae. Anamorphosis ignota.

Ascomata immersed in the host thallus, in groups of 20 to 100, obpyriform to subglobose, 220–300 high  $\times$  150–270  $\mu$ m diam, orange to yellow, KOH–; papilla truncate, 40  $\mu$ m high  $\times$  100–200  $\mu$ m diam, non-setose. Ascomatal wall 17–20  $\mu$ m thick, of two regions: outer region 9–13  $\mu$ m thick, of thick-walled, angular to rounded cells, 2–3.5  $\times$  1–1.5  $\mu$ m; inner region 7–8  $\mu$ m thick, of thin-walled, elongate cells, 2–8.5  $\times$  0.5  $\mu$ m. Asci clavate, 60–80  $\times$  7–8(–12)  $\mu$ m; apex truncate and simple; ascospores irregularly uniseriate to biseriate. Ascospores ovoid, (9–)15–20  $\times$  4.5–5(–6)  $\mu$ m, 1–septate, slightly constricted; hyaline, spinulose, spines sometimes arranged in rows as striations. Anamorph unknown.

Habitat.— On thallus of *Pertusaria* sp. Distribution.— France, Spain (Etayo, 1998), Sweden.

HOLOTYPE.— SWEDEN. Skåne: Brunnby par., Krapperup, by a road, on *Pertusaria pertusa* on *Ulmus*, 19 July 1947, R. Santesson (UPS).

ADDITIONAL SPECIMENS EXAMINED.— FRANCE, Brittany: Finistère, Coatadon, on Pertusaria pertusa (as P. communis), 18



June 1870, Crouan & Crouan (CO, filed under *Nectria*); same locality, [on *Pertusaria* sp.] on bark of elm, 18 Oct 1868, Crouan & Crouan (CO).

ETYMOLOGY.— The specific epithet is based on the host lichen. Pertusaria.

Notes.— The collections of *Pronectria pertusariicola* from Sweden and France differ somewhat in ascospore size. The collection from Sweden is chosen as the holotype because it is in better condition and is more readily available for study than the collections from the Crouan herbarium. *Pronectria pertusariicola* and *P. robergei* are morphologically similar, but *P. pertusariicola* differs in host and ascomatal wall anatomy, having asci with an apical ring, and ascospores that are more conspicuously ornamented than those of *P. robergei*.

Pronectria santessonii (Lowen & D. Hawksw.) Lowen, Mycotaxon 39: 462, 1990.

= Nectriella santessonii Lowen & D. Hawksw., Lichenologist 18: 322. 1986.

This species was described and illustrated in Lowen & Hawksworth (1986).

Pronectria septemseptata Etayo, Nova Hedwigia 67: 507. 1998.

Recently described and illustrated from Spain by Etayo (1998), this species is unique in having generally 7-septate ascospores.

Pronectria subimperspicua (Speg.) Lowen, Mycotaxon 39: 462. 1990. — Plate 10, a.

≡ Nectria subimperspicua Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 290, 1899.

Ascomata immersed, scattered or in groups of up to 30, obpyriform, 120–240 high × 120–240 μm diam, pale orange, outer wall region KOH+ red, changing to yellow in lactic acid; papilla conical to truncate, 60 μm diam, slightly paler than lateral and basal walls. Ascomatal wall 10 μm thick, of two regions: outer region 5 μm wide of thick-walled, round to oval cells; inner region 5 μm wide, of thick-walled, elongate cells. Centrum contents pale orange; orange oily drops emerging from crushed ascomata. Asci clavate, 40–50 × 6.5–7.5 μm, apex rounded, simple; asci in fascicles; ascospores diagonally uniseriate, filling the ascus. Ascospores subglobose, 6.5–8 × 5–6 μm, 1-septate, slightly constricted, at first hyaline, then pale orange, verruculose.

Habitat.— On thallus of *Punctelia*.

Distribution.— Known only from type.

HOLOTYPE. — ARGENTINA. Buenos Aires: La Plata, in park, on the wilting thallus of *Punctelia constantimontium* [as *Ricasolia casarettoana*]. 1 Apr 1890, Spegazzini 1618 (LPS).

Notes.— Pronectria subimperspicua differs from other species of Pronectria in the KOH reaction of the outer ascomatal wall. Pronectria paucispora is also found on Punctelia, but has subglobose ascomata and longer, narrower ascospores that those of P. subimperspicua.

Pronectria tenacis (Vouaux) Lowen, Mycotaxon 39: 462, 1990.

≡ Pharcidia mamillula (Anzi) Vouaux f. tenacis Vouaux, in Bouly de Lesdain, Rech. Lich. Dunkerque p. 273, 1910.

Ascomata immersed in host thallus and apothecia, scattered or in groups of up to six, obpyriform, 240  $\mu$ m high  $\times$  200  $\mu$ m diam, orange, KOH–, papilla conical to truncate, purplish red. Ascomatal wall 17–22  $\mu$ m thick, of one region of thin-walled rounded to angular cells 3  $\mu$ m diam, cells becoming longer and thinner toward the centrum. Centrum contents pale orange; orange oily drops emerging from crushed ascomata. Asci clavate,  $50-70\times8-14~\mu$ m, with an apical ring; ascospores biseriate. Ascospores ellipsoid–ovoid,  $10-16\times5-6~\mu$ m, 1-septate, at first hyaline, then pale orange, verruculose. Anamorph.— None known.

Habitat.— In thallus of Collema tenax on dunes.

Distribution.— United Kingdom and France.

NEOTYPE DESIGNATED HEREIN.— UNITED KINGDOM. England: North Devon: Braunton Barrows NNR, on *Collema tenax* on dunes, 16 Apr 1988, D.L. Hawksworth 5314 (IMI 327003).

Notes.— The type specimen at the Vouaux herbarium was destroyed, thus this taxon is neotypified with a specimen that agrees with the protologue. The sand dune habitat is similar to that described as the original locality. *Pronectria tenacis* is distinguished from other speces of *Pronectria* by the host lichen and the ascomata with purplish red papillae.

**Pronectria tenuispora** (D. Hawksw.) Lowen, Mycotaxon 39: 462, 1990.

■ Nectriella tenuispora D. Hawksw., Notes Roy. Bot. Gard. Edinburgh 36: 187. 1978.

This species was described and illustrated in Hawksworth (1978).

Pronectria terrestris Lowen & Diederich, Mycologia 82: 790. 1990.

This species was described and illustrated in Lowen & Diederich (1990).

Pronectria tincta (Fuckel) Lowen, Mycotaxon 39: 462, 1990.

≡ Cryptodiscus tinctus Fuckel, Fungi rhenani exs., Fasc. 4: 1836. 1867.

= Calonectria tincta (Fuckel) Rehm, Ann. Mycol. 8: 302.
1910.

[≡ Nectriella coccinea Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 177. 1869 [1870], non N. coccinea (Pers.: Fr.) Fr. 1849].

[= Calonectria fuckelii (Sacc.) Rehm, Tranzschel & Serebrianikow, Mycotheca Rossica Fasc. 2: 68, 1910, non C. fuckelii (Nitschke) Sacc., Michelia 1: 310, 1878].

= Calonectria fuckelii (Sacc.) Rehm f. everniae Rehm, in Motouschek, Centralbl. Bakteriol., Abth. 2, 42: 105. 1915.

Ascomata immersed in ectal excipulum and thallus, scattered or in groups of up to 15, obpyriform, 170–220  $\mu m$  high  $\times$  170–230  $\mu m$  diam, at first pale red, then pale yellow, KOH–, papilla conical, 75–80  $\mu m$  high  $\times$  100–120  $\mu m$  diam. Ascomatal wall 10–16  $\mu m$  thick, of one region of thin-walled cells. Asci clavate, 60–80  $\times$  9–11  $\mu m$ , apex rounded, simple; ascospores biseriate. Ascospores fusiform, 17–22  $\times$  4–5.5  $\mu m$ , 1-septate, cells inequal, one cell wider than the other, hyaline, verruculose, with one to many guttules per cell.

ANAMORPH.- None known.

Habitat.— On thallus and ectal excipulum of Anaptychia ciliaris.

DISTRIBUTION. -- Europe.

Type Specimens.— SWITZERLAND. Neuchâtel: near Neuchâtel, in Jura, on thallus and apothecia of *Anaptychia ciliaris* [as *Hagenium ciliarum*], Spring 1870, P. Morthier, Fungi rhenani exs. 1836 (holotype of *C. tinctus* G; isotypes,

Herb. Barb. Boiss. FH – Höhnel, G; IMI, K). RUSSIA. Prov. Kursk: Schebekino, in thallus of 'Evernia prunastri', 15 July 1908, Serebrianikow, in Tranzschel & Serebrianikow, Mycoth. Ross. Fasc. 2, 68 (isotypes, BPI, FH, K, S, all as Calonectria fuckelii f. everniae).

ADDITIONAL SPECIMENS EXAMINED.— FINLAND. Tavastia australis: Tammela, Mustiala, 13 Oct 1888, P.A. Karsten 2740 (H: as Nectria fuckelii); ibid., on Physcia stellaris, 4 Oct 1888, P.A. Karsten 2739 (H; as Nectria fuckelii); ibid., Oct 1888, P.A. Karsten 2738 (H; as Nectria fuckelii). USSR. Prov. Kursk: Schebekino, in thallus, Aug 1908, Serebrianikow, Rehm Ascom., Fasc. 46: 1897 (FH – Höhnel, K, S, all as Calonectria tincta).

Notes.— *Pronectria tincta* is distinguished from other species of *Pronectria* by the host and the inequal, fusiform ascospores.

Pronectria verrucariae (Vouaux) Lowen, Mycotaxon 39: 462, 1990.

≡ Nectria verrucariae Vouaux, Bull. Trimestrial Soc. Mycol. France 28: 186. 1912.

Ascomata immersed in ascomata of the lichen, visible as black or sometimes orange spots due to the barely visible papilla, scattered or in groups of up to 6, ascomata falling out leaving orange, circular craters, obpyriform, 180–320  $\mu m$  high  $\times$  150–280  $\mu m$  diam, pale orange, KOH–, papilla truncate, 60  $\mu m$  wide. Ascomatal wall 16–20  $\mu m$  thick, of one region, of thinwalled, elongate cells. Asci clavate, 44–70  $\times$  8–12  $\mu m$ , with 2–4 ascospores, apex with a ring; ascospores uniseriate. Ascospores ovoid to ellipsoid–fusiform, one end often pointed, other end rounded, 16–21  $\times$  5–7  $\mu m$ ,

#### KEY TO THE SPECIES OF PRONECTRIA

1.	Ascospores 3- or more septate
1.	Ascospores 1-septate
2 (1) 2.	Ascospores 3-septate, elongate ellipsoid, $15.5-21\times5-6~\mu m$
3 (1) 3.	Ascomata becoming darker in KOH and yellow in lactic acid; ascospores uniseriate . 4 Ascomata not changing color in KOH or lactic acid; ascospores biseriate
4 (3). 4.	Ascospores $6.5-8\times5-6~\mu m$ , pale orange, verruculose; ascomata immersed in thallus of <i>Punctelia constantimontium</i>
4.	of Lobaria
<b>5</b> (3). <b>5.</b>	Ascomata having a red ostiolar area 6 Ascomata concolorous 8
6 (5).	Algicolous, in stipe of <i>Laminaria</i> sp.; ascomata white with red ostiolar area; ascospores $13-20 \times 7-9 \mu m$ , pale brown, verruculose
6.	Lichenicolous, in thallus or apothecia of lichenized fungi; ascomata orange with red ostiolar area

	Ascospores ellipsoid–ovoid, 10–16 × 5–6 μm, pale orange, verruculose; in thallus and apothecia of Collema
7.	Ascospores fusiform, $22-28(-33) \times 3.5-5 \mu m$ , hyaline, smooth; in thallus of <i>Peltigera</i>
<b>8</b> (5). <b>8.</b>	Ascospores smooth-walled; ascomata subglobose 9 Ascospores usually ornamented; ascomata obpyriform 12
9 (8).	Ascomata immersed in the thallus of <i>Hypogymnia physodes</i> ; ascomata yellow to bright orange with white hairs around the ostiole; ascospores $12-17 \times 4-6~\mu m$ , pale
9.	Ascomata immersed in the thallus of other lichens; ascomata pale pink, yellow, red or dark red, without hairs; ascospores hyaline
10 (9). 10.	Ascomata dark red; ascospores ellipsoid, 14–20(–22) × (2.5–)4–6 µm; on <i>Punctelia</i> , known from France and the eastern United States
11.	Ascomata pale pink to yellow; ascospores ellipsoid to ovoid, 12–17 × 5.5–7 µm; on <i>Thrombium</i> , known only from Luxembourg
12 (8). 12.	Ascomata orange-brown, immersed in the thallus of <i>Physcia aipolia</i> ; ascospores ellipsoid-ovoid, $12-14(-18)\times5.5-8(-10)$ µm, hyaline, echinulate <i>P. echinulata</i> Ascomata pale yellow, immersed in thalli of other lichens; ascospores ellipsoid to fusiform
13 (12). 13.	Ascomata in <i>Peltigera</i> spp
14 (13). 14.	Ascospores (17–)18–20(~30) $\times$ (4–)5.5–6(–8) $\mu$ m, hyaline, becoming pale orange, vertuculose
15 (13). 15.	Ascospores ovoid, $9-12\times4.5-5~\mu m$ , echinulate; in <i>Pertusaria P. pertusariicola</i> Ascospores ellipsoid or fusiform, more than 12 $\mu m$ long; in other lichens 16
	Ascospores ellipsoid, $12-18 \times 4.5-8$ µm, hyaline, slightly echinulate; ascomata dark red; in <i>Anaptychia</i> ; known from Europe
16.	Ascospores ellipsoid or fusiform, averaging more than 17 µm long, hyaline to pale orange, smooth to slightly roughened, slightly echinulate or verruculose; ascomata pale yellow to pale or bright orange
17 (16).	In ascomata of <i>Verrucaria</i> ; ascospores $16-21 \times 5-7$ µm, ellipsoid to fusiform, rarely
17.	ovoid, hyaline to pale orange, smooth to slightly roughened
<b>18</b> (17).	In thallus and apothecia of <i>Xanthoria</i> ; ascospores 17–24 × 4–5 µm, ellipsoid, of equal cells, hyaline to pale orange; known from the United Kingdom and Luxembourg  P. xanthoriae
18.	In thalline exciple of Anaptychia ciliaris; ascospores 17–22 × 4–5.5 μm, fusiform,

1-septate, often slightly constricted, at first hyaline, then pale orange, smooth to slightly roughened, with 1 to several guttules per cell.

ANAMORPH. - None known.

Habitat.— In thallus and perithecia of *Verrucaria*.

Distribution.— France, United States (New York).

HOLOTYPE.— FRANCE. Nord: Dunkirk, on dunes near the lighthouse, in thallus of *Verrucaria integra* on calcareous rocks, 20 May 1904, B. de Lesdain (Herb. Vouaux; isotype, FH – Höhnel).

ADDITIONAL SPECIMEN EXAMINED.— UNITED STATES. New York: Niagara County, Goat Island, near shore, north slope on retaining wall, in ascomata of *Verrucaria muralis* associated with *Caloplaca feracissima*, 1 Nov 1989, R.C. Harris 22856A (NY).

Notes.— *Pronectria verrucariae* is distinguished from other species of *Pronectria* by the ascomata immersed in ascomata and thallus of the *Verrucaria* host.

Pronectria xanthoriae Lowen & Diederich, Mycologia 82: 788. 1990.

This species was described and illustrated in Lowen & Diederich (1990).

PROTOCREOPSIS Doi, Bull. Natl. Sci. Mus., Tokyo, B. 2: 129. 1976.

Type: P. musicola Doi, a synonym of P. fusigera (Berk. & Broome) Doi.

[= Cryptothecium Penz. & Sacc., Malpighia 1: 388. 1897,

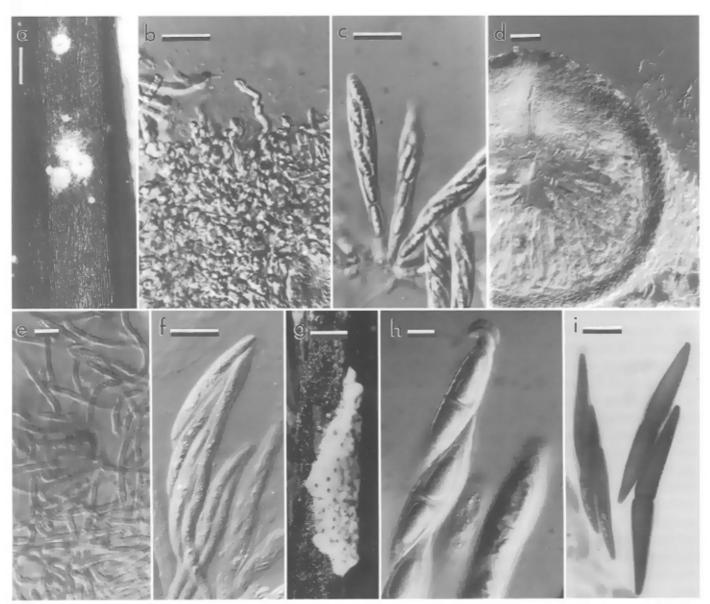


Plate 11. a–c. *Protocreopsis pertusa*. a. Ascomata on natural substratum. b. Ascomatal surface with hairs. c. Asci with ascospores. d–f. *Protocreopsis javanica*. d. Median section of ascoma. e. Hairs on ascomatal surface. f. Asci with ascospores. g–i. *P. fusigera*. g. Ascomata on natural substratum. h. Ascal apex and ascospores. i. Ascospores stained in cotton blue. a–c. G.J.S. 1288 – NY. d–f. Holotype of *P. palmicola* – TNS–F-192958. g–i. BPI 745424. h. TNS–F-226974, holotype of *P. musicola*. i. TNS–F-192691, holotype of *P. zingibericola*. Scale bars: a, g = 1 mm; b, c = 100 μm; d, i = 25 μm; e, h = 10 μm; f = 50 μm.

non Hübner 1851]. — Type: C. javanicum Penz. & Sacc. (

Calonectria javanica Höhn.), recognized as Protocreopsis javanica (Höhn.) Rossman & Samuels.]

Ascomata superficial on substrata, densely gregarious, less commonly solitary, surrounded by white to tan hyphae arising from ascomatal wall with few to many free ends visible, thus appearing Hypocrea-like, often extensive; ascomata hyaline to orange, KOH-; cells at ascomatal surface completely obscured by investing hyphae; ascomatal wall > 20 µm thick, comprising a single region of small, brick-like cells. Asci clavate to fusiform, apex simple or with an obscure ring; ascospores bi- to pluriseriate. Ascospores ellipsoid to fusiform, 1-septate, hyaline, typically striate, also smooth, punctate-striate, or tuberculate. Anamorph, where known, Acremonium-like. On dead monocotyle-donous substrata.

Notes.— Doi (1976) established *Protocreopis* for *Nectria*-like species having ascomata surrounded by a hyphal stroma. He included the type species, *P. musicola*, added two more species, *P. zingibericola* and *P. palmicola* (Doi, 1977), and later (Doi, 1978 a, b) revised the genus, adding several species and providing a key to all of them. The homonymous genus *Cryptothecium* Penz. & Sacc. was originally placed in the *Perisporieae* of the *Eurotiaceae*, although Rogerson (1970) included this genus in the *Hypocreales*. The type specimen of *C. javanicum* from PAD was examined, and, as suggested by the illustrations in Penzig & Saccardo (1897), this species was found to be an earlier name for *Protocreopsis palmicola* Doi.

Protocreopsis is most easily recognized by the ascomata that are completely enclosed in long, white to tan or green, flexuous hyphae. Species of Protocreopsis generally occur on monocotyledonous leaves, often on palms or Musaceae, that are decaying but still attached to living plants in tropical areas. Morphologically they are characterized by pallid ascomata surrounded by a hyphal stroma and striate ascospores. Species of Protocreopsis may be confused with those of Stilbocrea, a genus similar in having ascomata surrounded by an effused hyphal stroma or confluent hyphae or hairs. In Stilbocrea ascospores are generally spinulose, anamorphs are synnematous or pycnidial, and the species are corticolous, occasionally fungicolous, while in Protocreopsis ascospores are usually striate, although they may be smooth or tuberculate, anamorphs are Acremonium, and the species are herbicolous on monocots. Characteristics of the type species of Protocreopsis agree with those of species that have been placed in the Nectria subfalcata-group as defined by Samuels (1976) a, b). Although two species included by Samuels (1976)

a, b) in the Nectria subfalcata-group are now placed in Lasionectria, most members of that group are among the nine species of Protocreopsis.

Protocreopsis fusigera (Berk. & Broome) Doi, Bull. Natl. Sci. Mus., Tokyo, B. 4: 119. 1978. — Plate 4, c (see page 25); Plate 11, g-i.

- ≡ Hypocrea fusigera Berk. & Broome, J. Linn. Soc. 14:
  112. 1873.
- = Nectria subfalcata Henn., Hedwigia 41: 4. 1902.
- = Hypocrea bromeliicola Bat., Nascim. & Cif., Sydowia Beih. 1: 332. 1957.
- ≡ Nectria bromeliicola (Bat., Nascim. & Cif.) Samuels, Mem. New York Bot. Gard. 26: 32. 1976.
- ≡ Protocreopsis bromeliicola (Bat., Nascim. & Cif.) Doi, Bull. Natl. Sci. Mus., Tokyo, Ser. B. 4: 118, 1978.
- = Nectria heliconiae E. Müll. & Dennis, Kew Bull. 19: 383.
- = Protocreopsis musicola Doi, Bull. Natl. Sci. Mus., Tokyo, B. 2: 129, 1976.
- Protocreopsis zingibericola Doi, Kew Bull. 31: 552, 1976 [1977].

Anamorph. - Acremonium-like.

Mycelium spreading slightly from the ascomata, at first white and cottony, becoming roseous to buff, densely compacted, enveloping one to several ascomata in *Hypocrea*-like aggregates; hyphae surrounding the ascomata indefinite in length, flexuous, smooth, septate, branched, with many free ends, ca 5  $\mu$ m wide, with 1–2  $\mu$ m thick walls. Ascomata globose, 430–720  $\mu$ m diam, orange. Asci 125–185  $\times$  12–21  $\mu$ m, apex simple. Ascospores narrowly fusiform, 50–76  $\times$  6.5–9  $\mu$ m, without a sheath, smooth to finely striate.

ANAMORPH.— Conidiophores in culture unbranched, septate, monophialidic, 150–210 μm long, 4.5–6.5 μm wide at the base. Conidia fusiform to ellipsoid, 40–55 × 11–13 μm, basal abscission scar not recognizable, unicellular, hyaline, smooth, wall visibly thickened especially at the ends, held in a drop of hyaline liquid at the tip of each phialide.

Habitat.— On decaying debris of monocotyledonous plants, especially *Heliconia* and *Musa*.

DISTRIBUTION.— American tropics, Indonesia (North Sulawesi), western Pacific region; probably pantropical.

Types.— SRI LANKA (Ceylon). Peradeniya, on leaves of monocotyledonous plant, Nov 1867, No. 44 (K, holotype of *H. fusigera*); NEW BRITAIN ISLAND. Rabaul, on decayed stems and leaves of a species of *Zingiberaceae*, 1 Jan 1970, Y. Doi D.685, TNS-F-192961; holotype of *P. zingibericola*); same locality and substratum, 2 Jan 1970, Y. Doi D.708 (TNS-F-192959, paratype of *P. zingibericola*). PERU. Tingo María, about 700 m elev., on well-decayed leaf of *Musa*, 27 Jan 1976, Y. Doi D.2276 (TNS-F-226974, holotype of *P. musicola*).

Additional specimens examined are listed in Samuels (1976a) and Samuels et al. (1990).

ILLUSTRATIONS.— Batista & Ciferri (1957, Figs. 11, 12, as *H. bromeliicola*); Doi (1976, Fig. 7, as *P. musicola*); Doi (1978a, Fig. 4 as *P. bromeliicola*, Fig. 5); Müller & Dennis (1965, Fig. 4, as *N. heliconiae*); Samuels (1976a, Figs. 9S, 10, as *N. subfalcata*); Samuels *et al.* (1990, Fig. 27G, 28, as *N. subfalcata*).

Specimens illustrated.— PUERTO RICO. Luqillo Mts., Bisley Watershed 3, trail to stream, on *Heliconia* sp. at base of plant, 10 May 1995, S.M. Huhndorf 1379, D.J. Lodge 2273 (BPI 745424). ECUADOR. Prov. Pichincha, *ca* 19 km from Santo Domingo, on the new road from Santo Domingo to Quito, on herbaceous stem, elev. *ca* 860 m, 19 July 1975, K.P. Dumont-EC 682, S.E. Carpenter and P. Buriticà (NY, as *Nectria subfalcata*).

Notes.— The holotype of *Protocreopsis zingibericola* Doi is clearly *P. fusigera*, although the specimen consists of a few fragments of a leaf of *Zingiberaceae*. The ascomata appear immature, no asci were seen, but a few, very large conidia typical of *P. fusigera* are present.

Protocreopsis albofimbriata (Penz. & Sacc.) Doi, Bull. Natl. Sci. Mus., Tokyo, B. 4: 117. 1978.

≡ Nectria albofimbriata Penz. & Sacc., Malpighia 11:
513. 1897.

ANAMORPH: None known.

Mycelium tan, completely covering the aggregated ascomata; ascomata 250–350  $\mu$ m diam, hyphae arising from ascomatal wall, sinuous, septate, infrequently branched, with many free ends visible, 3–4  $\mu$ m wide, with ca 1  $\mu$ m thick walls. Asci (40–)55–75(–80)  $\times$  (9–)10–11(–12)  $\mu$ m, apex simple, 8-spored. Ascospores ellipsoid, 15.5–21  $\times$  4–6  $\mu$ m, equally 2-celled, not constricted at the septum, with many, low, ridge-like striations.

Habitat and distribution.— Known only from the type locality.

LECTOTYPE, designated here.— INDONESIA. Java: Tjibodas, on dead stems of (?) Elettaria sp., 6 Feb 1897, [? Penzig] 436a (PAD); same data. 6 Feb 1897. [? Penzig] 430 (PAD, syntype); same data. date unknown, [?Penzig] 172 (PAD, syntype).

ILLUSTRATIONS..— Doi (1978a, Fig. 3); Penzig & Saccardo (1904, Pl. 31, Fig. 1, as N. albofimbriata); Samuels et al. (1990, Fig. 27 A, B, as N. albofimbriata).

Notes.— Protocreopsis albofimbriata differs from P. pertusa in having larger ascospores with more pronounced, ridge-like striations. For discussion of this species, see Samuels et al. (1990).

Protocreopsis foliicola (Berk. & M.A. Curtis) Samuels & Rossman, comb. nov.

≡ Nectria foliicola Berk, & M.A. Curtis, J. Linn. Soc. Bot.
10: 378, 1869.

ANAMORPH: None known.

Mycelum completely enveloping groups of ascomata, hyphae straight, to 80  $\mu$ m long and free ends visible, ca 3  $\mu$ m wide, walls ca 0.5  $\mu$ m thick, tan. Ascomata 225–275  $\mu$ m diam, brown. Asci 60–80  $\times$  10–12  $\mu$ m, apex simple. Ascospores fusiform, straight or slightly curved or sigmoid, 21–27  $\times$  4–5  $\mu$ m, lacking a sheath, striate. Conidiophores not forming in culture. Ascomata forming in cultures derived from single ascospores.

Habitat.— On leaves of *Chusquea*, *Heliconia*, *Musa*, *Puya* and on peduncles of fruits of unidentified tree. Distribution.— Tropical America.

Type.— CUBA. On leaves of *Musa*, date not known, Wright 752 (K, holotype; FH, isotype).

Additional specimens examined are listed in Samuels (1976a).

ILLUSTRATIONS.— Samuels (1976a, Figs. 9D, 13, as N. foliico-la).

Notes.— Protocreopsis folicola is characterized by brown ascomata that are clothed in tan hyphal hairs. Despite isolation from several specimens, this species has never produced conidia in culture nor has an anamorph been observed associated with the teleomorph in nature.

Protocreopsis freycinetiae (Samuels) Samuels & Rossman, comb. nov.

≡ Nectria freycinetiae Samuels, New Zealand J. Bot. 14: 243. 1976 [as 'freycinetii'].

Anamorph: Acremonium-like.

Mycelium white, completely enveloping groups of ascomata, hyphae straight, 2–3  $\mu$ m wide, with 0.5–1  $\mu$ m thick walls, smooth, many free ends visible. Ascomata 260–370  $\mu$ m diam, orange. Asci 100–110  $\times$  11–17  $\mu$ m, apex simple. Ascospores fusiform, straight or slightly curved, 26–32  $\times$  6–7  $\mu$ m, lacking a sheath, striate. Conidiophores in culture unbranched, 4–6-septate, monophialidic, 60–90  $\mu$ m long, ca 3  $\mu$ m wide at the base, spinulose on phialides. Conidia ellipsoid, 6–10  $\times$  2–4  $\mu$ m, unicellular.

HABITAT AND DISTRIBUTION.— Known only from type specimen.

HOLOTYPE.— NEW ZEALAND. Auckland, Thames County, Coromandel Forest Park, Kauaeranga Valley, vic. Thames, on leaves of *Freycinetia banksii* [= *F. baueriana* subsp. banksii], 27 Aug 1974, G. J. Samuels 74–115 (PDD 32577). Culture CBS 573.76.

Notes.— This species is similar to P. foliicola, but the investing hyphae in the latter species are tan.

Protocreopsis javanica (Höhn.) Rossman & Samuels, comb. nov. — Plate 10 b; Plate 11, d-f.

[≡ Cryptothecium javanicum Penz. & Sacc., Malpighia 1: 388. 1897, genus illeg.]

Calonectria javanica Höhn., Sitzungsber. Kaiserl.
 Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 118: 1180. 1909.
 Protocreopsis palmicola Doi, Kew Bull. 31: 551. 1976.

Anamorph: None known.

Mycelium completely enveloping groups of ascomata, hyphae straight, roughened, 6–7  $\mu$ m wide, white to tan, loosely interwoven. Ascomata 300–400  $\mu$ m diam, pale yellow. Asci 55–70  $\times$  8–12  $\mu$ m, apex with a ring. Ascospores ellipsoid to fusiform or slightly sigmoid, 15–18  $\times$  4–5  $\mu$ m, lacking a sheath, striate, with many striae visible in one plane.

HABITAT .- On decaying palm leaves.

DISTRIBUTION.— Gabon, Indonesia, New Guinea, and Thailand.

Types.— INDONESIA. Java: Tjibodas, in foliis putrescentibus. 4 Mar 1897 (PAD, holotype of *Cryptothecium javanicum*). NEW GUINEA. Lae: near Markham Bridge, on rachides of decayed palm leaves, 20 Jan 1970, Doi D.786 (TNS–F-192958, holotype of *P. palmicola*; PNG, isotype, not examined).

Specimens examined,— GABON, Libreville: La Mondah, on dead palm parts, 8 Dec 1979, G. Gilles (BPI 745899): THAI-LAND, Nakorn Nayok Province: Khao Yai National Park, Phakrajai, on bamboo, 6 Aug 1997, G.J. Samuels (BPI 745855); same data (BPI 745867); same data, on palm (BPI 745858).

ILLUSTRATION .- Doi (1976, Fig. 1 A-G).

Notes.— Protocreopsis javanica resembles P. foliicola in the tan to brown coloration of the ascomata; however, ascospores of P. javanica are shorter than those of P. foliicola and the hyphae that envelop the ascomata of P. foliicola are smooth-walled. Protocreopsis javanica is the only member of the genus that has warted hyphae.

Protocreopsis pertusa (Pat.) Samuels & Rossman, comb. nov. — Plate 11, a-c.

■ Nectria pertusa Pat., in Patouillard & Lagerheim, Bull.
Soc. Mycol. France 11: 227, 1895.

= Nectria scitula Bres., Hedwigia 35: 299. 1896.

≡ Protocreopsis scitula (Bres.) Doi, Bull. Natl. Sci. Mus., Tokyo, B, 4: 116. 1978. = Nectria aemulans Rehm, Ann. Mycol. 7: 539, 1909.

ANAMORPH. - Acremonium-like.

Mycelium white but tan over individual ascomata, completely covering the ascomata; hyphae sinuous, smooth-walled, 2–2.5  $\mu m$  wide, with many short, free and hair-like ends around the ostiolar opening, walls visibly thickened, hyaline. Ascomata globose, 150–190  $\mu m$  diam, orange. Asci $70–80\times9–12$   $\mu m$ , apex simple. Ascospores ellipsoid,  $13–17\times4–5$   $\mu m$ , 3 or fewer striations visible in one plane of view. Conidiophores in culture unbranched, 0–1-septate, monophialidic, 20–30  $\mu m$  long, 2–3  $\mu m$  wide at the base: tip of phialide not flared. Conidia ellipsoid, 4–7  $\times$  1.5–2  $\mu m$ , unicellular.

Habitat.— On leaves of *Musa*, *Heliconia*, and palms, culms of bamboo, and rachides of tree ferns; on bark. Distribution.— Probably pantropical and subtropical.

Type.— ECUADOR. San George, on culms of *Chusquea* sp., July 1892, Lagerheim (FH, holotype of *N. pertusa*); BRAZIL. S. Catharina; Blumenau, on palm? leaf, date and collector not known, n. 9b, (lectotype of *N. scitula*, designated herein: S-herb Bresadola; isolectotype S-herb. Sydow).

ADDITIONAL SPECIMENS EXAMINED.— VENEZUELA. Amazonas. Cerro de la Neblina, elev. 1350 m, on dead fern rachis, 13 April 1984, G.J. Samuels 1288 (NY, VEN, filed as *Nectria pertusa*), and as listed in Samuels (1976 a, b, as *N. pertusa*) and Samuels *et al.* (1990, as *N. pertusa*). Culture CBS 568.76.

ILLUSTRATIONS.— Doi (1978a, Fig. 2, as *P. scitula*); Samuels (1976b, Fig. 2, as *N. pertusa*); Samuels *et al.* (1990, Figs. 27 C–F. as *N. cf. pertusa*).

Notes.— This is a common species in the American tropics. In a collection from Indonesia, Samuels *et al.* (1990) reported smaller ascospores than is usual for this species.

Protocreopsis pertusoides (Samuels) Samuels & Rossman, comb. nov.

■ Nectria pertusoides Samuels, New Zealand J. Bot. 14:
241, 1976.

Anamorph.— Acremonium-like.

Mycelium white, spreading over the substratum, completely enveloping groups of ascomata, hyphae straight to slightly sinuous, smooth-walled, 2–3  $\mu$ m wide, with 1–2  $\mu$ m thick, hyaline walls. Ascomata 220–375  $\mu$ m high × 220–280  $\mu$ m diam, orange. Asci 55–70 × 5.5–9  $\mu$ m, apex with an indistinct ring. Ascospores allantoid, ellipsoid to fusiform, 9–12 × 3–5  $\mu$ m, lacking a sheath, striate or punctate–striate. Conidiophores in culture un-

Plate 12. a-g. Protocreopsis viridis. a. Median section of ascomata on natural substratum, b. Close-up of ascomatal wall, c. Close-up of ascomatal apex. d. Ascomatal hairs, e. Asci with ascospores including remants of apical paraphyses in centrum. f-g. Asci with ascospores, g. stained in cotton blue to show median striations on ascospore wall. a-g. Holotype - BPI 745254. Scale bars:  $a = 100 \mu m$ ; b, c. f.  $g = 20 \mu m$ ;  $d = 50 \mu m$ ;  $e = 25 \mu m$ .

branched, 0–1-septate, monophialidic, smooth, 30–70  $\mu$ m long, 2–3  $\mu$ m wide at the base; tip of phialide not flared. Conidia ellipsoid to oblong, 6–7  $\times$  2–2.5  $\mu$ m, unicellular.

HABITAT.— On decaying herbaceous tissue, less frequently on bark.

DISTRIBUTION .- New Zealand.

HOLOTYPE.— NEW ZEALAND: Taranaki, Mt. Egmont Natl. Park, Stratford Mountain House, on dead leaves of *Cordyline indivisa*, G. J. Samuels 73-213 & C. S. Samuels (PDD 32032).

Additional specimens examined are listed in Samuels (1976b).

ILLUSTRATIONS.— Samuels (1976b; Figs. 3, 14, 26, as N. pertusoides).

Notes.— This species differs from *P. pertusa* in having smaller ascospores in which the striations are incomplete throughout the length, thus appearing punctate. Striations on ascospores of *P. pertusa* are always few in

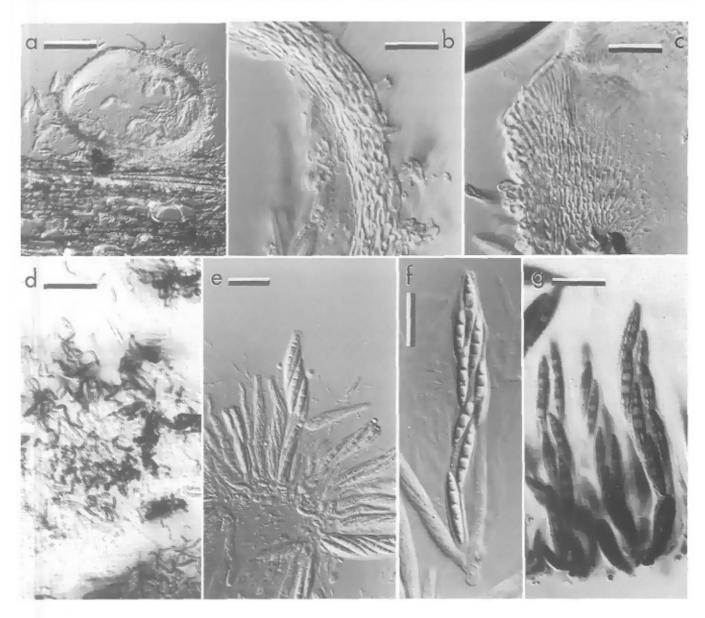
number and extend over the entire length of the ascospore.

Protocreopsis phormiicola (Samuels) Samuels & Rossman, comb. nov.

≡ Nectria phormiicola Samuels, New Zealand J. Bot. 14:
244. 1976.

Anamorph. - Acremonium-like.

Mycelium white, spreading over the substratum, completely covering individual ascomata; hyphae slightly sinuous, 2–3  $\mu m$  wide, walls 0.5–1  $\mu m$  thick, hyaline. Ascomata 150–220  $\mu m$  diam, yellow. Asci 60–70  $\times$  9–10  $\mu m$ , apex with an inconspicuous ring. Ascospores ellipsoid to fusiform–ellipsoid, 10–14  $\times$  3–4  $\mu m$ , surrounded by a sheath while still in the asci, tuberculate at maturity. Conidiophores in culture unbranched, 0–1-septate, monophialidic, smooth or spinulose, 33–45  $\mu m$ 



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long, 2–3  $\mu$ m wide at the base; tip of phialide not flared. Conidia ellipsoid, 5–7  $\times$  2–3  $\mu$ m, unicellular. Habitat and distribution.— Known only from the type locality.

HOLOTYPE.— NEW ZEALAND. Auckland: Waitemata County, Waitakere Ranges, vic. Piha, Marowhara Loop Track, on dead leaves of *Phormium tenax*, 17 Dec. 1974, J. M. Dingley *et al.* (PDD 32684; Samuels culture 74-133 = CBS 567.76 = ATCC 34049).

ILLUSTRATIONS.— Samuels (1976b, Figs. 5, 16, 28, as N. phormiicola).

Notes.— This species is unusual in the genus in having tuberculate ascospores that are surrounded by a gelatinous sheath while still in the asci.

Protocreopsis viridis Samuels, sp. nov. — Plate 12, a-g.

Perithecia solitaria vel pauca aggregata, hyphis viridibus, spinulosis vestita. Ascosporae (24–)26–29(–30) × 3.5–5.5 μm, anguste fusiformes, striatae, bicellulares.

ANAMORPH. None known.

Ascomata scattered and solitary or caespitose in groups of a few, non-stromatic, superficial, easily removed from the substratum, clothed in green hyphae, looking like masses of Trichoderma conidia, hyphae becoming brown in KOH, again green when followed by lactic acid. Ascomata subglobose, 250-300 µm high. 250-275 µm diam, non-papillate. Investing hyphae arising from ascomatal surface, strongly contorted, septate, branched, with few free ends visible, strongly spinulose, 2.5-3 µm wide, walls not thickened. Ascomatal wall ca 15 µm thick, of a single region of intertwined hyphae that appear cellular with individual cells ellipsoid, ca 5 µm long with walls ca 1.5 µm thick. Ascomatal apex a continuation of the wall below, the hyphae around the ostiolar opening forming a palisade of ca 3 µm wide cells with walls 1-1.5 µm thick. Asci narrowly clavate,  $85-120(-180) \times (9-)11-13(-15) \mu m$ , apex with an indistinct ring; ascospores pluriseriate. Ascospores narrowly fusiform, (24-)26-29(-30) × 3.5-5.5 µm, equally 1-septate, striate, striations ridgelike, few in number, staining in cotton blue/lactic acid.

Habitat.— On decaying palm leaf.

DISTRIBUTION.— Puerto Rico, known only from the type collection.

HOLOTYPE.— PUERTO RICO. Cordillera Central, Charioi Azul, off Rte. 184, elev. 550 m, on decaying palm leaf (*Praestoea monticola*), 25 Feb 1996, G. J. Samuels, D. J. Lodge & H. J. Schroers 8104 (BPI 745254).

#### KEY TO THE SPECIES OF PROTOCREOPSIS

	Ascospores more than 20 µm long
2. 2.	Ascospores 50–76 $\times$ 6.5–9 $\mu m$
	Enveloping hyphae green; ascospores (24–)26–29(–30) $\times$ 3.5–5.5 $\mu m$
	Enveloping hyphae tan; ascospores $2632\times67~\mu\text{m}$
5. 5.	Ascospores tuberculate
	Ascospores $9-12\times 3-5~\mu m$ , striations broken, punctate-striate, many visible in one plane of view
7. 7.	Only few (up to 3) striations visible in one plane of view
	Hyphae enveloping ascomata roughened, white to tan

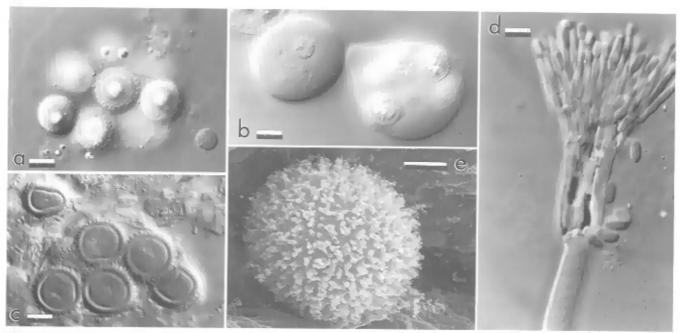


Plate 13. a-e. Roumegueriella rufula and anamorph, Gliocladium sp. a. Ascus with ascospores. b. Immature asci with ascospores. c. Ascus with ascospores in median focus to show wall ornamentation. d. Conidiophore with conidiogenous cells and developing conidia. e. SEM of ascospore. a-c. CBS 346.85. Scale bars: a-d = 10 μm; e = 5 μm.

Notes.— An attempt to isolate ascospores from the fresh specimen failed. No anamorph was associated with the ascomata on the host. This species is distinguished in *Protocreopsis* by the green hyphae that enclose the ascomata.

ROUMEGUERIELLA Speg., in Roumeguère & Spegazzini, Rev. Mycol. (Toulouse) 2: 18. 1880.

Type: R. muricospora Speg., a synonym of R. rufula (Berk. & Broome) Malloch & Cain.

= Lilliputia Boud. & Pat., Bull. Soc. Mycol. France 16: 144. 1900. — Type: L. gaillardii Boud. & Pat., a synonym of R. rufula (Berk. & Broome) Malloch & Cain.

= Lysipenicillium Bref., Unters. Gesammtgeb. Mykol. 14: 210. 1908. — Type: L. insigne Bref., a synonym of R. rufula (Berk. & Broome) Malloch & Cain.

Ascomata globose, soft, non-ostiolate, disintegrating at maturity, yellow to reddish-brown, of pseudoparenchymatous tissue; asci saccate, evanescent; ascospores globose, hyaline, ornamented. Anamorph *Gliocladium*-like. On dung and well-rotten debris.

Notes.— Spegazzini's generic description of Roumegueriella stated that the genus stands questionably between 'Sphaeropsideos et Hyphomycetes'; he apparently did not see asci in the type specimen and considered this to be an asexual fungus. However, Hughes (1951) and later Malloch & Cain (1972) reviewed the history of this genus noting that it is a cleistothecial ascomycete. Within the Hypocreales, Roumegueriella is one of six cleistothecial genera; it is most closely allied with another cleistothecial genus, *Heleo-coccum*, both of which were confirmed as members of the *Hypocreales* using molecular data (Rehner & Samuels, 1995). *Roumegueriella* includes two species.

The unispecific genus *Lilliputia* was originally described as a member of the *Tuberaceae* because of its cleistothecial ascomata. Malloch & Cain (1972) were the first to recognize that *L. gaillardii* is a synonym of *Roumegueriella rufula*.

Brefeld based his name Lysipenicillium upon Penicillium insigne without citing author and publication, simply stating: 'Eine Form von Penicillium ist als P. insigne fälschlich bezeichnet'. Although two later homonyms of this binomial exist, Brefeld was probably referring to P. insigne (G. Winter) Schröter based on Eurotium insigne G. Winter as listed below. Brefeld gave a clear description and illustration of Roumegueriella rufula including ascomata. The interpretation of Lysipenicillium as a possible synonym of Gliocladium as suggested by Raper & Thom (1949) is therefore not correct. All epithets described for this fungus in Gliocladium include the teleomorph and are therefore regarded as synonyms of R. rufula, while the anamorph strictly speaking has not been named.

Roumegueriella rufula (Berk. & Broome) Malloch & Cain, Canad. J. Bot. 50: 64. 1972. — Plate 13, a–e.

<sup>≡</sup> Chaetomium rufulum Berk. & Broome, Ann. Mag. Nat. Hist., Ser. 4, 11: 348. 1873.

<sup>≡</sup> Lilliputia rufula (Berk. & Broome) S. Hughes, Mycol. Pap. 42: 2. 1951.

= Eurotium insigne G. Winter, in Rabenh., Fungi Europaei no. 1732, 1874

≡ Lysipenicillium insigne Bref., Unters, Gesammtgeb. Mykol. 14: 210, 1908.

≡ Lilliputia insignis (G. Winter) Dennis & Wakefield, Trans. Brit. Mycol. Soc. 29: 145, 1946.

= Roumegueriella muricospora Speg., in Roumeguère & Spegazzini, Rev. Mycol. (Toulouse) 2: 18, 1880.

Cephalotheca francisci D. Sacc., Malpighia 12: 206, 1898.
 Lilliputia gaillardii Boud. & Pat., Bull. Soc. Mycol. France 14: 144, 1900.

= Mycogala macrospora Jaap, Verh. Bot. Ver. Brandenb. 52: 19. 1910.

= Gliocladium prolificum Bainier, Bull. Trimestriel Soc. Mycol. France 26: 385, 1910.

= Gliocladium borysseviczii Pidopl., Mikrobiol. Zhurn. 12(2): 36. 1950 (also Gribnaya Fl. grub. Kormov: 197. 1953).

Anamorph: Gliocladium-like.

Ascomata superficial, without a stroma, solitary or in groups of 2–3, irregularly globose, 450–640  $\mu m$  diam, dark yellow to reddish brown, non-ostiolate; ascomatal wall thin, of indistinct cells, becoming brittle and breaking down at maturity to expose the ascospores. Interthecial elements lacking. Asci irregularly saccate,  $46-66\times24-35~\mu m$ , evanescent at maturity. Ascospores globose,  $16-23~\mu m$  diam, hyaline, smooth when young, becoming densely echinulate.

HABITAT.— On various kinds of detritus including goose dung, damp paper, mushroom compost, nematodes, decaying seaweed, and rotting grass clippings.

DISTRIBUTION.— Belgium, England, France, Germany, India, Ireland, Japan (Udagawa et al., 1994), Mexico, Taiwan (Yaguchi et al., 1997), United States (California, Maine).

Types.— GERMANY. 'Auf Gänsemist in meinem Pilz-Cultur-Kasten. Halle a/S. im Juli 1873', Rabenhorst, Fungi Europaei no. 1732 (BPI. isotype of Eurotium insigne, none with good ascomata): Schleswig-Holstein: Reinbek bei Bergedorf, 'auf abgeschnittenem, faulendem Gras, 25 X. 1908, leg. Otto Jaap,' Jaap, Fungi Selecti Exs. no. 3961 (BPI, isotype of Mycogala macrospora). BELGIUM. Malmedy, 'in foliis et ramentis dejectis putrescentibus prope Malmedyanum et recentissime, aut. 1879, proxime Toloxam lectis' (isotype of Roumegueriella muricospora, not seen). FRANCE. Angers, 'dans la tannée ancienne d'une serre à Palmiers, inter frustulis caldario Andegavensi, Aprili 1900,' Revisio Reliquiae Libertianae (FH – Patouillard 4575, holotype of Lilliputia gaillardii).

ADDITIONAL SPECIMENS EXAMINED.— IRELAND. Rathmines Co., Dublin, on [herbivore] dung, 23 May 1935, S.W. Webb (BPI, as *Mycogala macrospora*). SWITZERLAND. Isolated from female *Globodera rostochiensis* buried in soil (CBS 346.85).

ILLUSTRATIONS.— Bainier (1910, Pl. 21, anamorph only); Brefeld (1912, Taf. VII, Figs. 1–7, as Lysipenicillium insigne); Hughes (1951, Fig. 12, as Lilliputia rufula; Pl. I, Fig.

5–9, as Chaetomium rufulum. Cephalotheca francisci, Eurotium insigne, Gliocladium prolificum, and Lilliputia gaillardii); Rabenhorst (1874, as E. insigne); Udagawa et al. (1994, Fig. G).

Notes.— Hughes (1951) and Malloch & Cain (1972) accounted for the various synonyms of *Roumegueriella rufula*. Hughes (1951) illustrated the variability in ascospore size and presented a detailed account of the specimens of the fungus known up to that time. The anamorph was described in detail by Bainier (1910) as *Gliocladium prolificum*.

A second species was described in *Roumegueriella* as *R. pulchella* Udagawa *et al.* (1994). Although similar to *R. rufula*, *R. pulchella* is differentiated by the small, ellipsoid ascospores,  $6.5-8\times4-5~\mu m$ , and the lack of an anamorph.

SELINIA P. Karst., Meddeland. Soc. Fauna Fl. Fenn. 1: 57, 1876

[≡ Hypocreopsis G. Winter, Hedwigia 14: 26, 1875, non P. Karst. 1873]. — Type: S. pulchra (G. Winter) P. Karst. ≡ Hypocreopsis pulchra G. Winter.

≡ Winteria Sacc., Michelia 1: 281, 1878. Type: Winteria pulchra '(G. Winter)' Sacc., a nomenclatural synonym of S. pulchra (G. Winter) P. Karst.

≡ Seliniana O. Kuntze, Revis, Gen. Pl. 2: 869, 1891. —
Type: Seliniana pulchra '(G. Winter)' O. Kuntze, a nomenclatural synonym of S. pulchra P. Karst.

Ascomata immersed in stromata, up to five immersed in each stroma, stromata of two parts, external part red-dish-brown, of non-descript, small, heavily pigmented cells, internal tissue of *textura epidermoidea*, thin-walled. Ascomata becoming erumpent, ostiolate, with a white, granular opening. Asci clavate. 8-spored. Ascospores ellipsoid, unicellular, hyaline, thick-walled, smooth. Anamorph phialidic. On dung.

Notes.— Selinia was described as a new name for Hypocreopsis G. Winter 1875, a later homonym of Hypocreopsis P. Karst. 1873. Winter considered his new genus to be similar to Hypocrea but differentiated by the few ascomata in each stroma, the distinct form of the asci and ascospores, and the presence of numerous paraphyses. Although paraphyses are described, they were not seen in the specimens examined of S. pulchra. Despite the soft-textured, light to bright-colored ascomata, several characteristics of the genus Selinia are unusual for members of the Hypocreales, specifically the thick-walled ascospores with nerve-like markings

on the inner wall and gelatinous epispore and occurrence on dung. In addition to *Selinia pulchra*, three other species are accepted in the genus as discussed by Khan & Krug (1989).

Selinia pulchra (G. Winter) P. Karst., Meddeland. Soc. Fauna Fl. Fenn. 1: 57. 1876. — Plate 14, a–f.

[≡ Hypocreopsis pulchra G. Winter, Hedwigia 14: 26. 1875, non Hypocreopsis P. Karst. 1873].

≡ Winteria pulchra (G. Winter) Sacc., Michelia 1: 281.
1878.

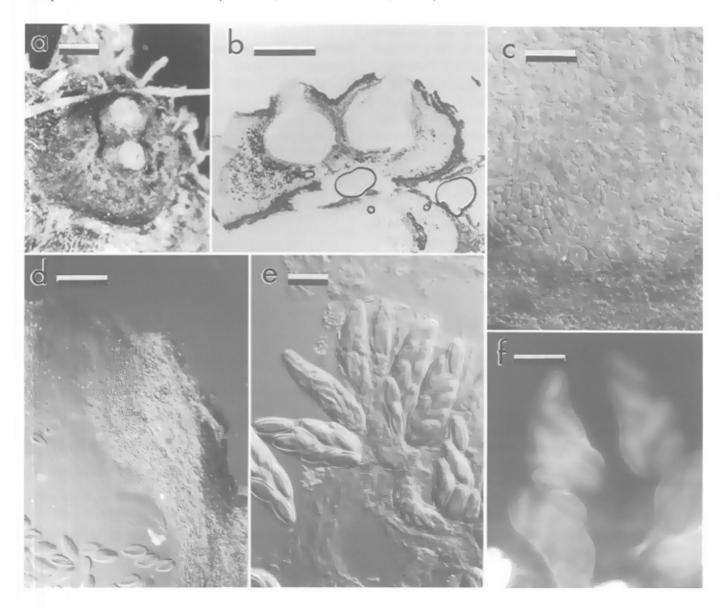
≡ Seliniana pulchra (G. Winter) O. Kuntze, Revis. Gen. Pl. 2: 869. 1891.

Stroma evident as small, less than 1 mm diam, amorphous, tar-like, blackened areas scattered on the substratum, consisting of two parts, external part rufous, ca 50 µm thick, of non-descript, small, heavily pigmented cells, several cell-layers thick; internal tissue textura epidermoidea, cells 7–10 µm diam, thin-walled, compact; ascomatal wall ca 40 µm thick, of about 10

layers of fusiform to rectangular cells,  $15-20~\mu m \log \times 4-7~\mu m$  diam, walls  $1.5-2.5~\mu m$  thick, hyaline; merging at the exterior with the surrounding stroma. Ascomata immersed in stroma, evident as a white, granular margin around the ostiolar opening, up to five ascomata in each stroma, apex protruding, venter remaining immersed in the stroma; papilla of vertically elongate cells, becoming progressively narrower toward the ostiolar canal, periphyses arising from and merging with elements of the papilla. Paraphyses not seen among the mature asci. Asci clavate, apex simple, 8-spored. Ascospores ellipsoid,  $45-56(-60)\times 20-24~\mu m$ , hyaline, smooth.

Habitat.— On dung of herbivores including cow, deer, hare, horse, opposum, and sheep.

DISTRIBUTION.— Argentina (Ranalli & Mercuri, 1995), Canada (Ontario); Chile (Udagawa, 1980), Denmark (Læssøe, 1995), Germany, Netherlands (von Arx & Müller, 1955; Larsen, 1971), Japan (Udagawa, 1980), New Zealand (Bell, 1975, 1983), United States (Florida, Maine).



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# KEY TO THE SPECIES OF SELINIA

1. 1.	Ascospores hore than 45 μm long	2 13
2. 2.	Ascospores $45-50\times 20-25~\mu m$ S. intermedial Ascospores $48-64\times 20-26~\mu m$ S. pulchi	ia
	Ascospores 25–30 × 12–13 μm	

Type.— ENGLAND: Shrewsbury, on sheep dung, W. Phillips, G. Winter, issued as C.B. Plowright, Sphaeriacei Britannici 100 (B – authentic specimen of *Hypocreopsis pulchra*).

Specimen examined.— UNITED STATES: Florida, off Mill-hopper Road, 10 mi. NW of Gainesville, on cow dung in pasture, 7 May 1970, J.W. Kimbrough (FLAS-F 48-978).

ILLUSTRATIONS.— von Arx & Müller (1955, Figs. 1, 2); Bell (1983, Fig. 35), Dennis (1978, Pl. 31D); Læssøe (1995, p. 52–53); Udagawa (1980, Figs. 8, 21).

Notes.— According to the original description, the type specimen of *Selinia pulchra* was found 'auf trocknem Schafkoth am Galgenberg bei Halle a. S.' No specimen agreeing with these data has been located. Plowright, Sphaeriacei Britannici no. 100 was apparently examined by Winter and is considered authentic material. Although unusual in having large, thick-walled, non-septate ascospores, developmental studies by Mercuri (pers. comm.) revealed the presence of apical paraphyses characteristic of hypocrealean fungi. Ranalli & Mercuri (1995) report an anamorph and were able to produce fertile ascomata in culture.

Three additional species are included in *Selinia*, *S. africana* Khan & Krug, *S. antartica* Speg., and *S. intermedia* Speg., as reviewed by Khan & Krug (1989). *Selinia intermedia* may be a synonym of *S. pulchra*.

STILBOCREA ['Stilbocera'] Pat., Bull. Soc. Mycol, France 16: 186, 1900.

Type: S. dussii Pat., a synonym of S. macrostoma (Berk. & M.A. Curtis) Höhn.

Ascomata immersed in a hyphal stroma that forms a continuous or discontinuous layer. Ascomata globose to ellipsoid or ovoid, pale yellow to orange, becoming red-brown or dark olive-green with age, KOH-, walls relatively thick, more than 25 µm. Asci narrowly clavate to cylindrical, 8-spored. Ascospores ellipsoid to ellipsoid-fusiform, 1-septate, hyaline, verrucose, echinulate or conspicuously spinulose. Anamorph Acremonium-like or Stilbella. On dead woody dicotyledonous and monocotyledonous substrata as well as decaying ascomycetous stromata.

Notes.— Patouillard described the genus Stilbocrea as being like Hypocrea but having a Stilbum anamorph and he included one species. Seifert (1985) studied the type specimens of Stilbocrea dussii and found Nectria macrostoma to be an earlier name. The genus Stilbocrea includes two additional species, namely S. gracilipes, with an even more darkly pigmented synnematal base than the dark green one of S. macrostoma, and S. impressa, with an Acremonium-like anamorph.

Stilbocrea macrostoma (Berk. & M.A. Curtis) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 118: 1185. 1909. — Plate 4, d (see page 25).

■ Nectria macrostoma Berk. & M.A. Curtis, J. Linn. Soc.,
Bot. 10: 378, 1868.

 ≡ Hypocreopsis macrostoma (Berk. & M.A. Curtis) E. Müll., Beitr. Kryptogamenfl. Schweiz 11(2): 650. 1962.

 = Hypocrea corticioides Berk. & Broome, J. Linn. Soc. Bot. 14: 111. 1873.

= Sphaerostilbe variabilis Berk. & Broome, J. Linn. Soc., Bot. 14: 115. 1875.

= Sphaerostilbe hypocreoides Kalchbr. & Cooke, Grevillea 9:26. 1880.

≡ Stilbocrea hypocreoides (Kalchbr. & Cooke) Seaver.
Mycologia 2: 62. 1910.

= Hypocrea carteri Berk. & Broome, Grevillea 12: 79. 1883.
≡ Hypocreopsis carteri (Berk. & Broome) Sacc., Syll. Fung. 9: 981, 1891.

= Stilbocrea dussii Pat., Bull. Soc. Mycol. France 16: 186.

= Sphaerostilbe henningsii Ferd. & Winge, Bot. Tidsskr. 29: 12, 1908.

[

Sphaerostilbe hypocreoides Hennings, Hedwigia 41: 4. 1902. non Kalchbr. & Cooke 1880].

= Sphaerostilbe intermedia Ferd. & Winge, Bot. Tidsskr. 29: 12. 1908.

≡ Stilbocrea intermedia (Ferd. & Winge) Seaver, Mycologia 2: 63, 1910.

= Sphaerostilbe placenta Theissen, Ann. Mycol. 9: 55. 1911. = Stilbocrea jenkiana [sic] Viégas, Bragantia 4: 97. 1944.

Anamorph: Stilbella aleuriata (Berk. & M.A. Curtis) Seifert, Stud. Mycol. 27: 54. 1985. ≡ Stilbum aleuriatum Berk. & M.A. Curtis, Grevillea 3:
63, 1874.

≡ Botryonipha aleuriata (Berk. & M.A. Curtis) O. Kuntze, Rev. Gen. Pl. 2: 845, 1891.

= Stilbum cinnabarinum Mont., Ann. Si. Nat., Bot. Sér. 2, 8: 360. 1837 (nom. rej.).

≡ Botryonipha cinnabarina (Mont.) O. Kuntze, Rev. Gen.
Pl. 2: 845, 1891.

≡ Stilbella cinnabarina (Mont.) Wollenw., Angew. Bot. 8:
195. 1926.

= Stilbum connatum Kalchbr. & Cooke, Gevillea 9: 22. 1880.
≡ Botryonipha connata (Kalchbr. & Cooke) O. Kuntze,
Rev. Gen. Pl. 2: 845. 1891.

Isaria aggregata Cooke & Massee, Grevillea 19: 48, 1890.
 Stilbum corallinum Cooke & Massee, Grevillea 19: 91, 1891.

= Stilbum subiculosum Pat., Bull. Trimestriel Soc. Mycol. France 20: 138, 1904.

= Stilbum intermedium Sacc. & Trott., Syll. Fung. 22: 477. 1913, as anamorph of Sphaerostilbe intermedia Ferd. & Winge.

= Stilbum vanderystii Sacc. & Trott., Syll. Fung. 22: 477. 1913, as anamorph of Sphaerostilbe henningsii Ferd. & Winge.

Stromata convex, round, ellipsoid or irregular in outline, at first white, becoming orange or pink and finally grey, smooth, punctate or pubescent, 1-15 mm long, 1-10 mm diam, 375-1250 µm thick, of textura intricata, hyphae 3-5 μm wide. Ascomata up to several hundred embedded in a single layer in the stroma, visible as a papilla protruding through the surface of the stroma, orange when young, becoming red-brown to dark olive-green with age, globose to subglobose, KOH-, 250-375 µm high × 200-300 µm diam, collapsing cupulate or laterally; papilla about 70-90 µm diam, of a palisade of vertically oriented, parallel hyphae, 2.5-3 μm wide; periphyses 1-1.5 μm wide. Ascomatal wall of a single, 10-30 µm thick region of compressed, ellipsoid to fusiform, hyaline cells,  $5-17 \times 2-3 \mu m$ , with walls slightly thickened, becoming less thickened towards the centrum; wall surrounded by textura intricata, hyphae 2.5–3  $\mu$ m wide. Asci cylindrical, 70–110  $\times$ 5-11 μm, with an apical ring, 8-spored, ascospores uniseriate. Ascospores ellipsoid to slightly fusiform,  $(8.5-)10-14 \times 4-6 \mu m$ , 1-septate, not constricted or slightly constricted at the septum, hyaline, verruculose to verrucose.

Anamorph.— Synnemata of two types, called A-synnemata and B-synnemata by Seifert (1985). A-synnemata solitary, gregarious, crowded or caespitose, arising from the teleomorph stroma, cylindrical–capitate, subulate–capitate, sometime clavate, slender to robust, curved, nodding or straight, unbranched or sometimes with a single branch, in some specimens with up to 10 inequivalent branches near the apex, smooth to granulose, stipe at first white, becoming orange or orange-pink, 250–2000  $\mu$ m tall  $\times$  50–225(–375)  $\mu$ m wide. B-synnemata solitary, gregarious to 3–4 caespi-

tose, emerging from a stroma at the periphery or several mm away from the stroma, subulate–capitate, cylindrical–capitate, or clavate, stipe grey, black or greybrown, shiny, sometimes granulose toward the apex, 1–2 mm tall, 50–200 μm wide. Conidiophores branching once, monoverticillate, stipe 1.5–2 μm wide. Phialides cylindrical, subulate, straight or curved, lateral and terminal or in terminal whorls of 3–4, 10–25 μm long × 1–1.5 μm wide, periclinal thickening sometimes obvious. Conidial mass hemispherical, globose or ellipsoid, orange, pink-orange, yellow-orange or red-brown, 75–350(–550) μm diam. Conidia oblong-ellipsoid, cylindrical or obovate, 3–6(–13) × 1–2(–2.5) μm. Descriptions modified from Seifert (1985).

Habitat.— On bark and decaying wood of dicotyledonous plants, often on other ascomycetes.

Distribution.— Pantropical and subtropical, occasionally temperate.

Types.— CUBA. On bark, date unknown, Wright 517 (K, holotype of *N. macrostoma*); SRI LANKA (Ceylon). South of the Island, on bark, July 1868, Herb. Berkeley No. 645 (K, holotype of *H. corticioides*); INDIA. Bombay, on bark, H.J. Carter, Herb. Berkeley, No. 8318 (K, holotype of *H. carteri*). Additional specimens examined from Indonesia are listed in Samuels *et al.* (1990). Culture CBS 542,95.

ILLUSTRATIONS.— Müller & von Arx (1962, Fig. 256, as Hypocreopsis macrostoma), Seifert (1985, Figs. 12–13, as N. macrostoma), Samuels et al. (1990, Fig. 29, as N. macrostoma).

Specimen illustrated.— UNITED STATES. Mississippi: Wilkinson Co., Clark Creek Natural Area, vicinity of pond, on twigs of dicotyledonous tree, 18 Aug 1996, G.J. Samuels, G.J.S. 96-175 ≡ CBS 101601, M. Blackwell (BPI 744508).

Notes.— Seifert (1985) presented a complete account of *Nectria macrostoma* and its anamorph, *Stilbella aleuriata*, with documentation of their numerous synonyms.

Stilbocrea gracilipes (Tul. & C. Tul.) Samuels & Seifert, comb. nov.

≡ Sphaerostilbe gracilipes Tul. & C. Tul., Sel. Fung. Carpol. 1: 131. 1861.

≡ Nectria gracilipes (Tul. & C. Tul.) Wollenw., Angew. Bot. 8: 198, 1926.

= Nectria dealbata Berk. & Broome, J. Linn. Soc., Bot. 14: 117, 1873.

= Nectria hypocreoides Berk, & Cooke, Grevillea 12: 81, 1884.

= Sphaerostilbe tetraspora Pat. & Gaill., Bull. Soc. Mycol. France 4: 115, 1888.

= Sphaerostilbe cinerascens Hennings, Ann. Jard. Bot. Buitenzorg, Sér. 11, 8: 63, 1909.

[= Sphaerostilbe ochracea Syd., Ann. Mus. Roy. Congo Belg., ser. 5, 3: 15. 1909, non Pat. 1903].

= Sphaerostilbe vanderystii Hennings, in Beeli, Bull, Jard, Bot, Nat, Belg, 8: 77, 1922.

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= Allantonectria creonectrioides Chardón, J. Dept. Agric. Porto Rico 14: 241. 1930.

Anamorph: Stilbella clavulata (Mont.) Seifert, Stud. Mycol. 27: 85. 1985.

≡ Stilbum clavulatum Mont., Ann. Sci. Nat. Bot., Sér. 2, 18: 248. 1842.

Additional synonyms listed in Seifert (1985).

Stroma white, grey or brown, erumpent, composed of textura intricata of hyaline hyphae, 3–9 µm wide, with smooth, to 0.5 µm thick walls. Ascomata 10–30(–50) caespitose, rarely solitary, seated on or slightly immersed in the stroma, globose to ellipsoid, 200–375 µm diam, sometimes collapsed cupulate, pale yellow, orange or pink when young, becoming brown with age, finally black, with an orange to black spot around the ostiole, KOH–, smooth-walled to granulose; papilla about 60–100 µm diam, of vertically oriented parallel, 1–2 µm wide hyphae, with slightly thickened walls; periphyses 1 µm wide. Ascomatal wall in surface view of

monilioid hyphae, 4–8  $\mu$ m wide, which ultimately fragment into globose to ellipsoid cells, 5–10  $\mu$ m diam, accumulating black pigment; in section 25–50  $\mu$ m thick, of two regions: outer region of *textura intricata* to *textura angularis*, 15–35  $\mu$ m thick, cells hyaline, 3–6  $\times$  3–8  $\mu$ m, becoming incrusted with black pigment with age, walls up to 0.5  $\mu$ m thick; inner region of *textura prismatica*, 10–20  $\mu$ m thick, of thin-walled, hyaline cells, 5–12  $\times$  3  $\mu$ m. Asci cylindrical, 60–90  $\times$  5–7  $\mu$ m, apex simple, 8-spored, ascospores uniseriate. Ascospores ellipsoid, often flattened on one side, 9.5–15(–17)  $\times$  3.5–6  $\mu$ m, 1-septate, not or slightly constricted at the septum, hyaline, spinulose.

Anamorph.— Synnemata scattered, gregarious, crowded, 2–10 caespitose, erumpent through the bark, emerging from a white basal subiculum or ascomatal stroma, cylindric-capitate, subulate-capitate to spathulate, straight, nodding or sinuous, usually slender, unbranched, or in some collections dichotomously to highly branched, stipe black, grey, or grey-brown,

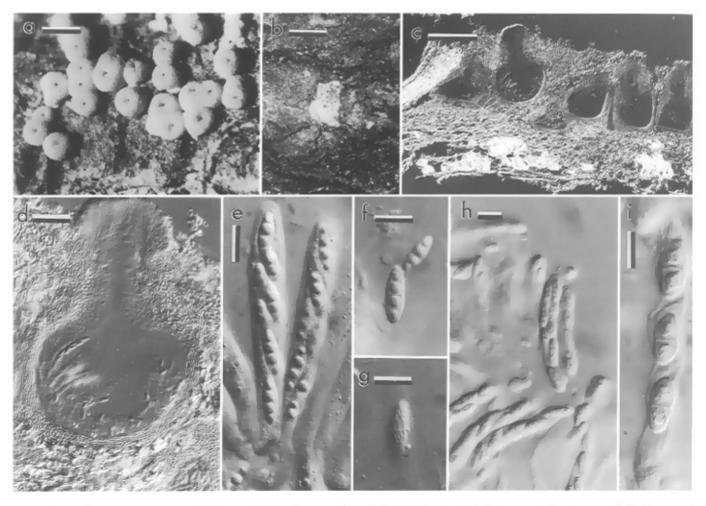


Plate 15. a. Stilbocrea impressa. Ascomata on natural substratum. b–g. Valsonectria simpsonii. b. Ascomata in stroma partially immersed in substratum. c. Median section of ascomata. d. Close-up of median section of ascomata. e. Asci with ascospores. f. Ascospores. g. Ascospore focussed out of median plane to show surface ornamentation. h. Valsonectria boldoae, asci with ascospores. i. Valsonectria pulchella, close-up of ascus with ascospores. a. BPI 745018. b–g. BPI 802564. h. LPS holotype. i. LPS holotype. Scale bars: a = 500 μm; b = 1 mm; c = 100 μm; d = 50 μm; e–i = 10 μm.

smooth to hirsute at the base, upper parts granulose, grey to white, 500-4000 µm tall × 50-250 µm wide in the middle, up to 450 µm wide at the base. Conidiophore branching once or twice monochasial. Phialides subulate or cylindrical, straight or curved, lateral, lateral and terminal, or in terminal whorls of 3(-6), 10-25  $\mu$ m long  $\times$  1–5–3  $\mu$ m wide at the base, sometimes proliferating percurrently to form a new phialide, 1-1.5 um wide at the conidiogenous aperture, collarettes sometimes slightly flared, periclinal thickening usually obvious. Conidial mass orange or green, at times yellow, red, or brown, hemisphaerical to globose, ovoid or ellipsoid, 60-625 µm diam. Conidia ellipsoid, oblong-ellipsoid or ovoid, straight or slightly curved.  $4-7(-8) \times 2-3(-3.5)$  µm, walls slightly thickened. Description modified from Seifert (1985).

Habitat.— On dead wood and bark of dicotyledonous plants and palms.

DISTRIBUTION.— Pantropical and warm temperate regions; anamorph common in temperate North America.

Types.— Holotype of Sphaerostilbe gracilipes at PC, fide Seifert (1985), not examined for this study. COLOMBIA. Dept. Valle de Cauca: Hacienda El Hatico between Cerrito and Palmira, on dead bark, 23 May 1929, C. E. Chardón & J. A. B. Nolla, 735 (CUP, holotype of Allantonectria creonectrioides). Cultures: CBS 657.83, 658.83, 523.85, 531.85, 532.85.

Additional specimens examined from Indonesia are listed in Samuels et al. (1990).

ILLUSTRATIONS.— Tulasne & Tulasne (1865, Pl. XIV, Figs. 14–19, as *Sphaerostilbe gracilipes*); Samuels *et al.* (1990, Fig. 30, as *N. gracilipes*), Seifert (1985, Figs. 26, 27, as *N. gracilipes*).

Stilbocrea impressa (Mont.) Samuels, comb. nov. — Plate 10, c, e; Plate 15a.

≡ Hypocrea impressa Mont., Ann. Sci. Nat. Bot., Sér. 4, 3: 143. 1855.

≡ Clintoniella impressa (Mont.) Sacc. & P. Syd., Syll. Fung. 16: 588. 1902.

Anamorph: Acremonium-like.

# KEY TO THE SPECIES OF STILBOCREA

Stroma 1-2 mm diam, about 350 µm high, erumpent through the bark at a single point, then spreading out, discrete, scattered, each with a single upper layer of 2-30 ascomata; stromal surface white to dull buff, slightly furfuraceous, hyphae highly branched, 3.5 µm wide, with many free ends, basal stroma of loose hyphae, occasionally associated with a black, pyenidial fungus. Ascomata globose, 280-500 µm diam, brown, KOH-; ascomatal wall 20-30 µm thick, of three regions: outer region ca 10 µm thick, of tightly compacted pseudoparenchyma, cells ca 5 µm diam, walls slightly thickened; middle region 15-20 µm thick, of loosely joined hyphae with cells  $10-15 \times 2-3 \mu m$ ; inner region ca 7  $\mu$ m thick, of compacted, fusoid cells, 5-7 × ca 1 μm. Periphyses 20-30 μm long, tapering from ca 1 μm in the lower part to the apex, stout toward outer regions. Asci clavate, 65-141 × 16-25 µm, apex simple, 8spored, ascospores biseriate or irregular. Ascospores ellipsoid to ellipsoid-fusiform, ends slightly pointed,  $15-22 \times 7-10$  µm, equally 1-septate, hyaline, with a sheath that at maturity appears prominently spinulose. Anamorph.— Buff-colored sporodochia formed on PDA after eight weeks. Sporodochia scattered, hemispherical, 1-2 mm diam, 45-55 μm thick, basal tissue of tightly compacted hyphae. Conidiophores arising as lateral branches from basal hyphae forming abundantly in sporodochia or from surface of agar, densely packed into a compact palisade, loosely and more or less verticillately branched, macronematous, mononematous,  $25-37 \mu m \log \times 2-3 \mu m$  at the base to  $1-1.5 \mu m$  at the apex, straight, smooth, aseptate or 1-septate, not conspicuously thickened or flared at the apex, proliferating percurrently to form a second full-length phialide, conidia held in hyaline to buff drops of liquid. Phialides straight, smooth, tapering gradually and uniformly from 2-3  $\mu$ m at the base to ca 1  $\mu$ m at the apex, apex with periclinal thickening, not flared, many phialides proliferating percurrently to form a second phialide. Conidia oblong, straight or slightly curved, with apical end often broader than the basal end,  $3.5-7.5 \times 1.5-3$ um, unicellular, hyaline, smooth, lacking an obvious

basal abscission scar.

- Ascomata orange to red-brown or dark olive-green; ascospores (8.5–)10–14 × 4–6 μm, verrucose to verruculose
   S. macrostoma

Habitat.— On small, dead twigs with bark still present.

Distribution.— Northern South America (Brazil, French Guiana, and Guyana)

HOLOTYPE. — FRENCH GUIANA. On bark, Leprieur, Crypt. Guyan. 517 (PC).

ADDITIONAL SPECIMENS EXAMINED.— BRAZIL. Amazonas, 0.3 km S of central portion of Serra Araca and 8 km E of Rio Javari. ca 1 h walk SE from camp, on twigs with bark, elev. 60 m. 00°49′ N, 63°19′ W. 6 Mar 1984, G.J. Samuels 84-256 (NY). FRENCH GUIANA. Remire, 15 km from Cayenne, trail to Vidal old farm, 52°18′ W, 4°52′ N, on dead twigs, 12 Feb. 1988, A. Rossman 3198, cult. G.J. Samuels 88-8 ≡ CBS 101600 (BPI, CAY); St. Laurent, near Mana, along road to Jahoury. white sand forest. 53°51′ W, 5°34′ N, on dead wood, 18 Nov 1986, A. Rossman 4020, cult. G.J. Samuels 86-570 (BPI 745018, CAY).

Notes.— Doi (1975a) described and illustrated a *Hypocrea* as *H. impressa* but, based on a comparison of his description with the type specimen of *H. impressa*, Doi's specimen is of *Hypocrea*, not *Stilbocrea*.

TRICHONECTRIA Kirschst., Verh. Bot. Vereins Prov. Brandenburg 47: 60. 1906 [1907].

Type: T. aculeata Kirschst., a synonym of Trichonectria hirta (Bloxam) Petch.

Ascomata solitary, superficial, without stroma; globose to subglobose, white, yellow, pale orange to pale red-dish-brown, occasionally pink when fresh, KOH-, not collapsing when dry, wall less than 40 µm thick, of small, non-descript cells; with thick-walled, straight, solitary hairs, or glassy, hyaline appendages in *T. hyalocristata*, arising from the surface of the ascomatal wall. Asci clavate, broadly clavate or broadly cylindrical. Ascospores ellipsoid, fusiform, broadly to long fusiform or cylindrical, 1- to multiseptate, hyaline, smooth-walled. Anamorph, where known, *Acremonium*-like. On decaying algae, mosses, lichenized and non-lichenized fungi.

Notes.— Trichonectria was described as being similar to Calonectria and Ophionectria, differentiated by the delicate, prosenchymatous ascomata covered with stiff hairs. The type specimen of the type species, T. aculeata, was apparently destroyed and Hawksworth (1978) neotypified that name with the type of T. hirta, thus assuring the synonymy of these two species. Trichonectria is characterized by a pallid, thin-walled ascomata having hyaline, thick-walled hairs and are often associated with lichens, bryophytes, and fungi.

Döbbeler (1978) described one additional species in Trichonectria, T. pellucida, from living leaves of a liverwort in Brazil, Samuels (1988) added four species, three of which are fungicolous, and Scheuer (1988, 1989) described a species having glassy, flattened, hyaline appendages around the ostiolar region that occurs on overwintered leaves of Carex. At present, seven species are included in *Trichonectria*. Species of *Trichonectria* show affinities to species of *Nectriopsis* and *Paranectria* in having small, pallid, thin-walled ascomata and occurring primarily on algae, bryophytes, lichens or other fungi.

Trichonectria hirta (Bloxam) Petch, Naturalist (Hull) 1937; 282, 1937.

≡ Nectria hirta Bloxam, in Currey, Trans. Linn. Soc. London 24: 158. 1884.

≡ Calonectria hirta (Bloxam) Sacc., Michelia 1: 307.
1878.

≡ Lasionectria hirta (Bloxam) Cooke, Grevillea 12: 112.
1884.

= Calonectria vermispora Massee & Crossland, Naturalist (Hull) 1904; 4, 1904.

≡ Dialonectria vermispora (Massee & Crossland) Massee & Crossland, Fungus Flora Yorkshire p. 214. 1905.

= Trichonectria aculeata Kirschst., Verh. Bot. Vereins Prov. Brandenburg 47: 60, 1906 [1907].

≡ Calonectria aculeata (Kirschst.) Weese, Centralbl. Bakteriol., Abth. 2, 42: 595, 1914.

= Trichonectria rosella Höhn., Sitzungsber, Kaiserl, Akad. Wiss., Math.-Naturwiss, Kl., Abt. 1, 127: 624, 1918.

Ascomata solitary or sparsely aggregated, superficial on the substratum, without a stroma or with a sparse hyphal subiculum radiating from the base of the ascomata, hyphae hyaline, thin-walled, 1.5-2 µm wide. Ascomata globose to subglobose, 230-300 μm high × 240-300 µm diam, laterally pinched or not collapsing when dry, white to pale yellow or pale saffron, KOH-, without papilla, with numerous, long, straight, solitary hairs projecting from the upper part of the ascomata. hairs  $43-100 \mu m \log \times 10-12 \mu m$  wide at the base. with 1.5-3 µm thick walls, unthickened toward the apex. Ascomatal wall 25-40 µm thick, of two regions: outer region 20-35 µm thick, of hyaline, thin-walled, elongate cells, 6-12 × 3-4 μm; inner region 3-7 μm thick of hyaline, thin-walled, elongate cells. Asci broadly cylindrical, 60-100 × 11-15 μm, apex simple. 8-spored, ascospores pluriseriate. Ascospores long fusiform to cylindrical, vermiform or sigmoid, 45-85 × 5-8 μm. 11-21-septate, forming cuboid to subcuboid cells, hyaline, smooth.

Habitat.— On very rotten, decorticated wood, often associated with granular lichen thalli.

DISTRIBUTION.— Austria, Belgium, Denmark (T. Læssøe, pers. comm.), England, and Germany.

HOLOTYPE. — England. Twycross, Leicestershire, on decaying rails, associated with *Lecidiea uliginosa* (K).

Additional specimens examined as cited in Rossman (1983).

ILLUSTRATIONS.— Dennis (1978, Pl. 31G); Döbbeler (1978, Fig. 21, as *T. rosella*); Hawksworth (1978, Fig. 7); Petch (1938, Fig. 26); Rossman (1983, Fig. 46).

# KEY TO THE SPECIES OF TRICHONECTRIA

1.	Ascomata with glassy, flattened, hyaline appendages around the ostiolar region; ascospores fusiform, 9.5–12.5 × 2–2.5 μm; on overwintered leaves of <i>Carex</i> ; Austria and Great Britain
1.	Ascomata with straight hairs, not on overwintered leaves of Carex
	Ascospores more than 25 $\mu$ m long
	Ascospores long fusiform to cylindrical, with broadly rounded ends, $45-85 \times 5-8$ µm, on old wood associated with granular lichen thalli; Europe
	On leaves of $Erythroxylum$ ; ascomatal hairs 25–35 $\mu$ m long; ascospores fusiform (12.5–)13.5–16.5(–17) × (2.5–)3–3.5(–4) $\mu$ m; Brazil
	On hymenium of inoperculate discomycetes; hairs 30–60 $\mu$ m long; ascospores narrowly ellipsoid to fusiform, (10–)11.5–13.5(–14.5) × 2–3 $\mu$ m; Venezuela
	Hairs 75–150 $\mu$ m long; ascospores fusiform, (12–)14.5–18.5(–24.5) × (2.5–)3–4 $\mu$ m; on stromata of <i>Diatrype stigma</i> ; North America

ADDITIONAL SPECIES OF TRICHONECTRIA:

Trichonectria albidopilosa (Rogerson & Samuels) Samuels, Mem. New York Bot. Gard. 48: 11. 1988. This species was described and illustrated in Rogerson & Samuels (1985, as *Nectria albidopilosa*).

Trichonectria erythroxylifolii Samuels, Mem. New York Bot, Gard. 48: 11. 1988.

This species was described and illustrated in Samuels (1988).

Trichonectria horrida Samuels, Mem. New York Bot. Gard. 48: 11. 1988.

This species was described and illustrated in Samuels (1988).

Trichonectria hyalocristata Scheuer, Mycol Res. 93: 117, 1989.

This species was described and illustrated in Scheuer (1988, 1989).

**Trichonectria pellucida** Döbbeler, Mitt. Bot. Staatssamml. München 14: 119. 1978.

This species was described from living leaves of a liverwort in Brazil by Döbbeler (1978). **Trichonectria rectipila** Samuels, Rogerson & M.E. Barr, Mem. New York Bot. Gard. 48: 11. 1988. This species was described and illustrated in Samuels (1988) and is known from the eastern United States,

VALSONECTRIA Speg., Anales Soc. Ci. Argent. 12: 211. 1881.

Type: V. pulchella Speg.

= Endocreas Samuels & Rogerson, Stud. Mycol. 31: 145. 1989. — Type: E. lasiacidis Samuels & Rogerson, recognized as V. lasiacidis (Samuels & Rogerson) Samuels & Rossman. Stroma immersed in the substratum, becoming partially erumpent, pale yellow, pseudoparenchymatous. Ascomata immersed in the stroma, globose to subglobose, yellow, KOH—, ostiolate. Asci clavate to cylindrical. Ascospores narrowly ellipsoid, ellipsoid to fusiform, equally 1-septate, hyaline or yellow-brown, smooth or coarsely striate. Anamorph, where known, Acremonium-like. On living and dead woody substrata and bamboo-like grasses.

Notes.— Spegazzini (1881) established Valsonectria for a species having Nectria-like ascomata immersed in

a valsoid stroma and lightly pigmented, one-septate ascospores. Petrak & Sydow (1936) examined the type specimen of *V. pulchella*, provided a detailed description, and concluded that the genus belonged in the *Hypocreales*. Müller & von Arx (1962) examined the type specimen and presented a description with illustrations that show the ascomata immersed at different depths in the stroma, although, based on our examination of the type specimen, the ascomata form a single layer in the substratum.

Endocreas Samuels & Rogerson was described as a hypocrealean genus similar to Nectriella (Samuels & Rogerson, 1989). In a recent publication Seifert & Samuels (1997) recognized the synonymy of Endocreas with Valsonectria and transferred the type species of Endocreas to Valsonectria. Valsonectria is distinguished from other genera in the Bionectriaceae by ascomata immersed in a well-developed stroma that is itself immersed in the substratum, and hyaline to yellow-brown, smooth or coarsely striate ascospores. At present, four species are included in Valsonectria.

Valsonectria pulchella Speg., Anales Soc. Ci. Argent. 12: 211. 1881. — Plate 10, e.

Stromata evident as ellipsoid, brightly colored, slightly raised bumps on bark, about 1 mm long, forming a single layer in the substratum, hyphae loosely disposed, 2-3 µm wide, hyaline, thin-walled, branched, septate. Ascomata immersed in stromal periphery, globose, yelnon-papillate, ostiolate. Asci cylindrical,  $88.5-97.5 \times 7.5 \,\mu\text{m}$ , apex simple, 8-spored, ascospores uniseriate. Ascospores ellipsoid to fusiform, 11-14 × 6-7 µm, equally 2-celled, yellow-brown, with wall up to I µm thick, coarsely striate, numerous, short, the striations resulting from accretion to the spore surface. Anamorph associated with mature ascomata, forming on the stromal surface. Phialides narrowly cylindrical,  $7-15 \times 1.5-2$  µm, widest at the base, collarette not flared, periclinal thickening not visible. Conidia unicellular, hyaline, without a recognizable basal abscission scar, slightly curved.

Habitat and distribution.— Known only from the type specimen.

HOLOTYPE.— ARGENTINA. Flores: Buenos Aires, on decaying branches of *Melia azedarach*, Apr 1881, C. Spegazzini (LPS 1217).

Notes.— The holotype specimen consists of a few, small woody twigs and most material is immature.

Valsonectria lasiacidis (Samuels & Rogerson) Samuels & Rossman, Mycologia 89: 515. 1997.

≡ Endocreas lasiacidis Samuels & Rogerson, Stud. Mycol. 31: 145. 1989.

Stromata immersed, becoming partially erumpent and rupturing the epidermis, linear parallel to the long axis of the host culm, pale yellow, KOH-, pseudoparenchymatous, cells 6-10 μm diam, with ca 1 μm thick walls, arranged in files perpendicular to the surface of the host; each stroma enclosing several ascomata; stromal tissue lacking below ascomata, ascomata seated directly on host tissue; hyphae invading epidermal cells of host. Ascomata globose, 145-155 µm high × (132-)165-185 µm diam, non-papillate or with a short papilla, remaining immersed within the stroma, pale yellow, KOH-. Ascomatal wall 10-15 µm thick, of a single region of small, ellipsoid to flattened cells. Ascomatal apex formed of hyphal elements 1 µm or less wide, merging with the periphyses at the interior and with surrounding stromal cells at the exterior. Sterile filaments persisting or not among mature asci, 3-4 µm wide, constricted at the refractive septa. Asci clavate, (48-)53-66 × (8-)9-12(-13.5) μm, with simple apex; 8-spored, ascospores completely filling each ascus or up to 20 µm of the ascal base empty. Ascospores narrowly ellipsoid to fusiform,  $(13.5-)15-17.5(-20) \times 2.5-3.5(-4.5) \mu m$ , equally 1-septate, not constricted at the septum, hyaline. smooth, becoming coarsely striate with age.

Habitat.— On dead culms of Lasiacis ligulata (Poaceae).

DISTRIBUTION. - French Guiana.

Types.— FRENCH GUIANA. Saül, elev. 200 m, Feb 1986. G.J. Samuels 3785 (NY, holotype); Upper Marouini River, ca 3 h walk W of river, ca 1 km E of Roche Koutou. 02°55′ N, 54°03′ W, elev. 15–350 m, 18 Aug 1987, G.J. Samuels 5866 (NY, paratype). Culture CBS 179.88,

Notes.— Although unique in many characteristics, the sparsely developed stroma, pallid, discrete ascomata, shape, septation, and striation of the ascospores, and the occurrence on dead herbaceous material suggest affinities with the genus *Bionectria*.

Valsonectria boldoae Speg., Revista Fac. Agron. Univ. Nac. La Plata, Ser. 2, 6: 98, 1910. — Plate 10, d.

Ascomata immersed, caespitose in groups of up to 10 but not obviously stromatic, globose, about 375–400 μm diam, orange, KOH–, smooth, non-papillate, with a viscid ostiolar opening. Ascomatal wall 25–30 μm thick, of a single region of intertwined hyphal cells. Ascomatal surface of textura epidermoidea. Asci cylindrical, 28–40 × 6–7.5 μm, apex simple, 8-spored, as-

cospores biseriate. Ascospores narrowly ellipsoid to oblong,  $8-11 \times 2-3$  µm, equally 1-septate, straight or slightly curved, hyaline, smooth-walled.

Habitat and distribution.— Known only from the type specimen.

HOLOTYPE.— CHILE. Talchuano, on Boldoa fragrans, Jan 1909, C. Spegazzini (LPS 1754).

Notes.— A large packet contains two smaller packets each with a handwritten label. One has a drawing of two perithecia, an ascus with allantoid ascospores, and some unicellular, allantoid ascospores, and is labelled 'Nectriovalsa boldoaea Speg. (n.sp.) cum Mattirolia (Diatrype) vitellina (Montagne) Speg.' This specimen is regarded as the holotype. The second packet has a drawing of allantoid spores and is labelled 'Diatrype [enteracantha] Berk. vii. p. 47.' No hypocreacealean fungus was found in that packet.

Valsonectria simpsonii Samuels & Seifert, Mycologia 89: 512. 1997. — Plate 15, b–g.

Stromata inconspicuous to 1 mm diam, white, subcortical, erumpent, ostiolar openings appearing as pale orange dots, stromata of hyphae 5–6  $\mu$ m wide, thickwalled, densely compacted especially at the surface, stroma lacking below the ascomata. Ascomata immersed in groups of 30–50, ca 275  $\mu$ m high × 190  $\mu$ m diam. Ascomatal wall ca 15  $\mu$ m thick, of a single region of ellipsoidal cells with 1.5  $\mu$ m thick walls. Asci cylindrical to narrowly clavate, 50–73 × 6–9  $\mu$ m, apex thickened, 8-spored. Ascospores ellipsoidal to fusiform, (9.5–)10.5–13 (–14) × 3.5–4.5  $\mu$ m, equally 2-

celled with two guttules in each cell, becoming slightly constricted at the septum, yellow-brown, coarsely striate while in the asci.

Anamorph. -- Synnemata in nature 150-625 µm tall. 30-160 µm wide, sometimes almost sessile, gregarious or caespitose, more or less cylindrical or clavate, unbranched, or often with several median to apical branches, white to straw-colored, with brownish bands. Conidiophore branching monoverticillate, occasionally with a basal dichotomous branch; metulae, when present, 15–25  $\times$  1.2-2  $\mu$ m; phialides 15-37  $\times$  0.5-1.5  $\mu$ m, hyaline, cylindrical to slightly subulate, often curved, sinuous and uneven in outline, in whorls of 2-4, sometimes regenerating percurrently to produce a new conidiogenous aperture, conidiogenous aperture 0.5-1 µm wide, periclinal thickening obvious with phase contrast, collarette inconspicuous, cylindrical, about 1 µm long. Conidial mass slimy, in nature, sparse, pale salmon-colored, up to 500  $\mu m$  diam. Conidia ellipsoidal or ovate, 3.5-6  $\times$  1.5-2.5 μm, sometimes slightly truncate at the base.

Habitat.— On living branch of *Elaeagnus pungens* Thunb.

DISTRIBUTION.— France (Pyrénées Atlantiques), known only from the type locality.

HOLOTYPE.— FRANCE. Pyrénées Atlantiques: Île de Sauveterre, on *Elaeagnus pungens*, 31 Oct 1992, F. Candoussau 252 (BPI 802564; ex-type culture G.J.S. 93-9 ≡ CBS 101602, also DAOM).

Notes.— Valsonectria simpsonii can be distinguished from the other three species in the genus on the basis of the host, ascospore characters, and the white, smooth synnematous anamorph with branching monoverticillate conidiophores.

#### KEY TO THE SPECIES OF VALSONECTRIA

- On living or dead wood; ascospores less than 15 μm long, hyaline or yellow-brown, smooth or coarsely striate